Matrix Metalloproteinase Inhibitors in Cancer Therapy: Turning Past Failures Into Future Successes

Arthur Winer, Sylvia Adams, and Paolo Mignatti

Abstract

The matrix metalloproteinases (MMP) are a family of proteolytic enzymes that degrade multiple components of the extracellular matrix. A large body of experimental and clinical evidence has implicated MMPs in tumor invasion, neoangiogenesis, and metastasis, and therefore they represent ideal pharmacologic targets for cancer therapy. From the 1990s to early 2000s, synthetic inhibitors of MMPs (MMPI) were studied in various cancer types. Unexpectedly, despite strongly promising preclinical data, all trials were unsuccessful in reducing tumor burden or improving overall survival; in addition, MMPIs had unforeseen, severe side effects. Two main reasons can explain the failure of MMPIs in clinical trials. It has now become apparent that some MMPs have antitumor effects; therefore, the broad-spectrum MMPIs used in the initial trials might block these MMPs and result in tumor progression. In addition, although MMPs are involved in the early stages of tumor progression, MMPIs were tested in patients with advanced disease, beyond the stage when these compounds could be effective. As more specific MMPIs are now available, MMP targeting could be reconsidered for cancer therapy; however, new trials should be designed to test their antimetastatic properties in early-stage tumors, and endpoints should focus on parameters other than decreasing metastatic tumor burden.

Future Successes

Introduction

Cancer remains a leading cause of mortality worldwide, in some estimates accounting for more deaths than coronary artery disease or stroke. In the United Stated, over 1.5 million new cases were diagnosed in 2016, leading to 595,000 deaths (1). Patients die of metastatic disease; therefore, prevention of metastasis and treatment of micrometastatic disease is most important to improve cure rates. The mechanisms by which tumors metastasize are complex and involve numerous interactions between tumor cells and their microenvironment. A malignant cell invades into the surrounding tissue, enters the vasculature, and extravasates at distant sites. Proteolytic enzymes are essential for this process, degrading the extracellular matrix (ECM) and allowing for tumor dissemination (2–4). While hundreds of proteinase genes have been identified, the matrix metalloproteinases (MMP) have been heavily implicated in metastatic spread (5).

The MMPs are a family of 24 endopeptidases that control the physiologic turnover of the ECM. High levels of MMP correlate with unfavorable prognosis in multiple cancers (5). Therefore, clinical trials of synthetic MMP inhibitors (MMPI) were performed during the late 1990s and early 2000s (6–8). However, these studies failed due to lack of efficacy and severe side effects. This review will discuss the preclinical data that indicated the potential efficacy of MMPIs in cancer, the clinical trials and what led to their failure, and offer a perspective on potential trial designs.

History and Biology of MMPs

The MMPs mediate the constant remodeling the extracellular matrix. While their substrates include collagens, gelatins, proteoglycans and elastin, they have wide-reaching effects on many other proteins (2). The first vertebrate MMP described was the collagenase associated with the resorption of the tadpole tail, in 1962 (9). Human collagenase (now known as MMP-1) was identified in the skin 5 years later, and similar enzymes were further characterized across species (10). Initially MMPs were categorized on the basis of their substrate specificity (e.g., collagenase). However, as further MMPs were discovered, it became evident that many substrates are degraded by multiple MMPs, and each MMP can degrade multiple substrates; therefore, a sequential numbering system was adopted reflecting the order in which MMPs were discovered. The MMPs have also been classified per their structure and function into eight groups that comprise secreted and membrane-bound MMPs (membrane-type or MT-MMPs; ref. 11). As MMPs cleave numerous substrates, their activity heavily impacts the extracellular environment and, left unchecked, their action can be disastrous (12). Their activity is therefore strictly regulated to prevent excessive ECM degradation. MMP synthesis is first controlled at the level of transcription and translation, and posttranslational modifications also regulate MMP activity (11). Like all extracellular proteinases, MMPs are secreted as proenzymes, orzymogens, rendered inactive by the interaction between the zinc ion in the catalytic domain and a cysteine-sulphydryl group in the N-terminal (pro) domain. Activation requires

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removal of this interaction, a mechanism termed "cysteine switch" which can occur either after secretion or intracellularly by prohormone convertases (furin). Following the cysteine switch, MMPs are only partially activated; complete activation is achieved by a process of autocalysis, in which the proteinase cleaves its prodomain. The enzyme can further degrade and inactivate itself, a mechanism of regulation in multiple MMPs.

MMP proteolytic activity is further controlled by specific protein inhibitors, the tissue inhibitors of metalloproteinases (TIMP), comprising a family of four proteins (TIMP-1 to 4) that reversibly bind to the MMP catalytic site in a stoichiometric manner (2, 13). MMPs can also be inhibited by nonspecific inhibitors including α2-macroglobulin, thrombospondin-1, and -2 (13).

**MMPs in Malignancy**

**Role in metastatic spread**

MMPs exert profound effects on the extracellular microenvironment and are therefore highly regulated in normal physiology. Invasive malignancies can "deregulate" these proteinases to spread beyond their microenvironment in the complex, multistep metastatic process (5). Highly motile, invasive tumor cells egress from the primary tumor in either a collective pattern in which cell–cell interactions are closely maintained and cells move in broad sheets, or in a streaming pattern in which cells maintain a loose connection moving along the same pathway (11). Single-cell migration also occurs, whereby cells move by adopting an amoeboid-like phenotype or a mesenchymal phenotype, a process that mimics the epithelial–mesenchymal transition (EMT) that occurs during embryo development. This transition involves a decrease in E-cadherin expression with a concomitant increase in expression of N-cadherin. Indeed, multiple tumors show decreased E-cadherin levels, which reflect a decrease in synthesis and/or degradation by several MMPs, including MMP-9, -10, and -15 (14, 15).

Whatever the mode of local invasion, tumor cells must breach histologic barriers, basement membrane, stroma, and vascular basal lamina, to move into the blood stream and spread to distant sites (Fig. 1). This process requires the degradation of their molecular components, and multiple studies have shown that MMPs play an important role (5, 13, 16–19). After entering the bloodstream, tumor cells invade again through the vascular basal lamina to extravasate into distant tissues. Multiple MMPs including MMP-2, -9, and -14 can degrade the basal lamina of capillary vessels and have been implicated in tumor cell extravasation (20). MMPs also have complex effects on growth factors and cytokines (13). Upregulation of the COX-2 pathway is associated with increased blood–brain barrier permeability and breast cancer cell entry into the CNS (21). Experimental studies of human
melanoma have shown that MMP-2 upregulates tumor cell secretion of VEGF-A, which in turn activates the vascular endothelium favoring melanoma cell interaction with the blood vessel lining and their extravasation (21, 22).

Once tumor cells extravasate, a metastatic niche must be set up to permit tumor cell growth in an unfavorable environment. MMPs promote this process through several mechanisms. Angiogenesis, the formation of capillary blood vessels from preexisting vasculature, involves multiple interactions between stroma and vascular cells. A number of MMPs, including MMP-1, -2, -7, -9, and -14 contribute to angiogenesis via several mechanisms (23). In addition to mediating the ECM degradation necessary for endothelial cell migration into the tumor to be vascularized, MMPs contribute to the release of proangiogenic factors such as VEGF, fibroblast growth factor-2 (FGF-2), and TGFβ from the ECM (2). These growth factors are sequestered in the stroma, and metastatic foci utilize MMPs to create a favorable metastatic niche by mobilizing these factors to support tumor growth.

ECM-degrading enzymes further influence metastatic cell survival by modulating apoptosis. MMP-7 confers a survival benefit to tumor cells by cleaving Fas ligand, removing it from the cell surface and preventing it from stimulating the Fas death receptor, a potent mediator of innate apoptotic pathways (24). By this mechanism, malignant cells evade apoptosis and may also gain resistance to chemotherapeutics (25). Other MMPs, such as MMP-14, also promote tumor progression through antiapoptotic interactions with the surrounding microenvironment (Fig. 1; ref. 26).

The immune system surveilles the body for tumor cells, recognizing and killing malignant cells by recruiting neutrophils, macrophages, and tumor-specific T lymphocytes. Cancer cells have developed multiple mechanisms utilizing MMPs to evade the immune system, thereby ensuring metastatic cell survival. Tumors utilize MMPs to cleave chemokines, preventing inflammatory cell chemotaxis and recruitment to involved tissues (27). In melanoma, MMP-2–conditioned dendritic cells prime naïve CD4 T cells to differentiate toward the Th2 cell pathway, thereby skewing the immune response (28). MMP-1, -2, and -9 downregulate interleukin receptor on the surface of T cells, further dampening immunity and promoting tolerance of cancer (29). TIMP-2 downregulation has also been implicated in suppressing local immune function, allowing cancer cells to escape (30).

**MMP expression and modulation in cancer**

MMP overexpression has been well documented in multiple types of solid tumors (31). High levels of MMPs have been correlated with poor overall survival in virtually all solid malignancies (13, 31–33). Studies have also shown significant associations between tumor aggressiveness and elevated MMP expression. For example, distant metastases from breast cancer have been correlated with high levels of multiple MMPs including MMP-1, -7, -9, -11, and -13 (34). MMP-13 levels are also increased in lung and prostate malignancies (32, 33). MMP-9 overexpression has been strongly associated with poor prognosis in multiple malignancies including breast, lung, colon, gastric, pancreatic, and prostate cancer (33, 35–38). However, despite hundreds of observational studies in humans correlating high MMP levels with metastatic spread or recurrence, only MMP-11 (stromelysin) has thus far become part of a prognostic assay, the OncotypeDX platform, a clinically validated 21-gene array for prognosticating recurrence and guiding therapy in early hormone receptor–positive, HER2-negative breast cancer (39).

TIMP levels also change as tumors become more aggressive, and ultimately TIMP deregulation contributes to metastatic spread (40). However, while TIMP downregulation is expected to favor tumor progression, evidence shows that more complex mechanisms are in play; some TIMPs are in fact upregulated, while others are silenced. TIMP-1 overexpression is associated with unfavorable prognosis and early recurrence in multiple cancers including breast and prostate carcinoma (41, 42). Conversely, the lack of TIMP-1 expression predicts both a favorable prognosis and tumor responsiveness to chemotherapy in some cancers (43). In contrast to TIMP-1 overexpression correlating with poor survival in the metastatic setting, strong data support that multiple human cancers silence TIMP-3 as they spread (40). This effect seems to imply that TIMP-3 functions as a tumor suppressor gene and that by turning off its expression, tumors are allowed unchecked growth (44). Regardless of which TIMPs are upregulated or silenced, growing evidence shows that their deregulation contributes to metastatic spread of malignancy, and therefore represents a potential therapeutic target.

**Clinical Trials of MMPs**

Given the robust experimental and clinical evidence associating MMPs with tumor progression and poor prognosis, several MMPIs were synthesized and trialed from the late 1980s into the early 2000s for various cancer types (Table 1; ref. 45). One of the first drugs developed was batimastat, a small peptidomimetic molecule designed to mimic the most common MMP substrate, collagen. Batimastat showed broad-spectrum inhibition of virtually all MMP family members. Preclinical data indicated a promising antitumor effect of the drug; however, early trials showed that its water insolubility resulted in low oral bioavailability (46). Although several phase I studies showed efficacy with direct injection of the drug into the pleural or peritoneal space of patients with malignant effusions or ascites, significant toxicity, including pain, pyrexia, transaminitis, dyspnea, cough, and nausea, was observed. Therefore, further testing was not pursued, given the development of a more readily orally bioavailable drug, marimastat (47, 48).

Marimastat was developed as a next-generation oral analogue with a similar mechanism of action as batimastat. It too showed much promise in the preclinical setting, and reached phase II and III clinical trials in the metastatic setting for multiple solid tumor types including pancreatic, lung, breast, colorectal, brain, and prostate cancer (7, 8, 49–51). Despite the breadth of these trials, they uniformly failed to demonstrate a survival benefit. Many patients also had a negative impact on their quality of life due to a debilitating “musculoskeletal syndrome” consisting of joint pain, stiffness, and inflammation, which forced the discontinuation of the drug in several patients (52). One trial which evaluated the drug for unresectable gastric carcinoma did show a modest survival benefit at 2 years (9% in the treatment arm vs. 3% in the placebo group), but again with significant musculoskeletal toxicity (53).

The musculoskeletal syndrome seen in patients treated with batimastat and marimastat has been attributed to the inhibition of two members of the ADAM (a disintegrin and metalloproteinase) family of proteinases, ADAM-10 and -17. These
ADAMS are also termed "sheddases" as they cleave the membrane-bound precursor of tumor necrosis factor-α (TNFα), shedding the active form into the circulation. However, ADAM-10 and -17 are also responsible for the degradation of TNFα receptors, serving as a regulatory mechanism for TNFα action (54). Inhibiting the activity of these proteinases therefore disrupts this balance as receptors remain upregulated and activated TNFα molecules are able to bind to their unregulated receptors, contributing to the musculoskeletal symptoms seen in these patients. Significant fibrosis has also been described in subjects treated with marimastat, due to MMP-1 inhibition. The inhibition of this enzyme prevents interstitial type 1 collagen remodeling, leading to excessive deposition in the ECM and fibrosis, which may have contributed to some of the severe side effects that led to the failure of marimastat (55).

Other more selective inhibitors that avoided inhibition of ADAM-10 and -17 were then trialed, including tanomastat, a small-molecule inhibitor of MMP-2, -3, -8, -9, and -13, prinomastat, which inhibits MMP-2, -3, -9, -13, and -14, and rebimastat, an inhibitor of MMP-1, -2, -3, -8, -9, -13, -14 (56). All these inhibitors were studied in the metastatic setting of ovarian, pancreatic, lung, breast, and prostate carcinomas (6, 57–63). Unfortunately, despite their narrower inhibitory action, these trials failed to demonstrate a positive effect on survival. While musculoskeletal toxicity was seen less often with these inhibitors, some studies still reported significant joint pain and swelling, as well as bone marrow suppression and venous thromboembolism. Ultimately, further trials of MMPIs were halted after these negative results were published in the mid-2000s (Table 1).

**Why did MMPIs block tumor progression in mice but not in man?**

Several reasons have been hypothesized to explain why, despite preclinical and clinical evidence implicating MMPs in tumor growth and metastasis, clinical trials of MMPIs were unsuccessful (5, 17, 19). First, the difference between human and murine...
Matrix Metalloproteinase Inhibitors in Cancer Therapy

Our knowledge of the biochemistry and biology of MMPs has grown considerably in the 15 years since the clinical trials of MMPIs were halted. New MMPs have been discovered and new roles of already known MMPs, including inflammation and protection against cancer, have been revealed (67). New molecular genetics techniques such as CRISPR-Cas9 and the availability of genetic tools for the tissue-and time-specific silencing of genes, make the generation of mouse models easier, faster and cheaper than 15 years ago. The combination of conditional MMP knock-out models with spontaneous tumor models can afford clear, unambiguous target validation before MMPIs are generated and brought into preclinical and clinical studies. Detailed analyses of MMP molecular structures have provided accurate information of the determinants of their substrate specificity, paving the path to the design of novel, highly selective and potent MMPIs based on differing mechanisms of action (69). These advances can allow us to overcome the limitations that potentially caused the failure of the clinical trials, and reconsider MMPi treatment of metastatic cancer in a new light.

The first-generation MMPis were designed to target the MMP catalytic site, which is highly conserved (i.e., very similar) in all members of the MMP family. This approach resulted in a generation of drugs that could effectively block MMP-mediated proteolysis but lacked the ability to selectively inhibit the specific MMP(s) associated with a given tumor. In the light of today's knowledge that some MMPs have antitumor effects in some types of cancers, considerable effort has been, and is being put into the

What Can We Do Next?

New, selective MMPis: patient selection based on individual MMP expression, novel clinical trial design

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design of MMPIs that are highly selective and possibly inhibit only deleterious functions of specific, tumor-associated MMPs. A turning point in this effort was determined over a decade ago by findings that MMP activity can be inhibited specifically by targeting molecular structures outside of the catalytic domain (so-called "exosites"; ref. 69). Unlike the catalytic domains, different exosites are present in different MMPs, and their targeting with synthetic, low-molecular weight compounds or antibodies can result in the selective inhibition of even a specific function of a single MMP. This approach has led to the generation of highly selective mAbs to MMP-9, AB0041 and AB0046, and the humanized version of AB0041, CS-5745, which have shown efficacy in a mouse xenograft model of colorectal carcinoma (69). A set of mAbs to exosites of MMP-14 (LEM-2/15, -2/63, and -1/58) have shown high selectivity for MMP-14. Importantly, LEM-2/15 specifically inhibits MMP-14 degradation of gelatin and collagen type I without affecting its capacity to activate proMMP-2, an important function of MMP-14. Conversely, another antibody to MMP-14 (9E5), which is also highly selective for this MMP, has no effect on MMP-14 proteolytic activity, but inhibits proMMP-2 activation, showing that specific MMP functions can be selectively inhibited (69).

Selective MMP inhibition can also be achieved by the use of "endogenous" or "intrinsic" MMP inhibitors. Like all extracellular proteases, MMPs are secreted as inactive proenzymes whose proteolytic activity is inhibited by the intramolecular interaction of the catalytic domain with the N-terminal "pro" domain. Removal of the prodomain by proteolysis or other mechanisms results in activation of the proMMP. Unlike the highly conserved catalytic domains, the pro domains differ from one MMP to another, a difference that can be exploited to generate specific protein inhibitors. Exogenous addition of the pro domains of the sheddases ADAM10 and ADAM17 results in selective inhibition of the respective enzyme, without cross-reactivity in spite of the high similarity of the two ADAMs (69).

Other approaches to the development of selective and specific MMPIs have used sophisticated biochemical techniques such as protein engineering and directed evolution to improve the inhibitory activity of antibodies and TIMPs. Anti-MMP-14 antibodies that effectively reduce tumor growth and metastasis in preclinical models have been generated by selection of a phage display library of single-chain variable fragments (scFv), followed by protein engineering to increase their affinity and inhibitory activity. A mAb to MMP-14, DX-2400, was selected by screening a human Fab-phage library for candidates binding selectively to the MMP-14 catalytic domain. DX-2400 is a high-affinity, highly selective inhibitor of MMP-14 that retards tumor growth and metastasis in several in vivo mouse models of breast cancer and melanoma, both as single agent and in combination with paclitaxel or bevacizumab. mAb fragments (scFv) have also been developed to MMP-1, MMP-2, and MMP-3 by a combination of phage display library screening and combinatorial mutagenesis (69).

Protein engineering has been used to generate TIMP variants that specifically inhibit MMP-14. Point mutations in the sequence of TIMP-2 increase binding to and inhibition of MMP14 by 9–14 folds. TIMP mutants with inhibitory activity toward MMPs that were not their native targets were developed by protein engineering, directed evolution and computational design. By these methods, a mutant of TIMP-2 was generated, which selectively blocks MMP-14 activity with an inhibition constant of 0.9 pmol/L, the strongest inhibitor of this MMP thus far generated (69).

Thus, a number of novel MMPIs have been and continue to be engineered with the high affinity, specificity and selectivity that earlier-generation MMPIs lacked. These features can circumvent not only the potentially deleterious inhibition of protective MMPs but also avoid the onset of the musculoskeletal syndrome.

Novel molecular biology techniques afford relatively fast and inexpensive analysis of MMP expression in very small biological samples or even single-cells. Diagnostic bioptic material can provide sufficient tumor tissue to analyze the expression of the MMPs expressed by the individual patient's tumor. Relatively low numbers of tumor cells can be identified in peripheral blood and analyzed for MMP expression; tumor cell DNA can also be detected in the circulation, providing a potential surrogate of metastasis. These techniques can allow identification of the MMP(s) produced by a tumor, rapid assessment of the treatment efficacy, and therefore a precision medicine approach to anti-MMP treatment.

However, to effectively test MMPIs, a fundamental shift in clinical trial design is necessary. Currently, investigational cancer drugs are first tested in advanced cancer patients with overt metastatic disease. As MMPIs act most effectively (and almost exclusively) in the pre- and perimetastatic setting, these clinical trials can only be ineffective. To effectively study MMPIs, new trials should be designed, incorporating early-stage patients in the premetastatic setting.

Traditionally, in neoadjuvant and adjuvant trials recurrence-free survival or freedom from metastatic disease is the required primary endpoint for approval of therapies in the early disease setting. These trials are costly as they require enrollment of many patients and outcome readouts take years or even decades. Therefore, to meaningfully study the effectiveness of systemic cytotoxic drugs, antitumor efficacy is first determined in the metastatic setting, then the drug is moved to neoadjuvant stages and surrogates of early response, such as pathologic complete response (pCR), are assessed. This strategy has proven useful, for instance, in aggressive breast cancers, where pCR rates correlate with freedom from metastases and survival, supporting testing of cytotoxic compounds in this setting.

As MMPIs are not expected to decrease tumor size alternative surrogates must be tested, such as decrease in circulating tumor cells (as indicator of decrease in micrometastases) and/or decrease in tumor-associated MMP activity. MMPIs should therefore be studied in two stages of a phase II trial. In stage one, surrogate markers should be used as an endpoint instead of recurrence-free survival, to demonstrate target inhibition (which could also direct dose finding) and possibly early effectiveness. For example, a preclinical model in mice used markers such as change in target mRNA expression to show drug effectiveness, and correlated this outcome with a decrease in bone metastases (70). In the case of MMPI therapy, target MMP activity could be used as surrogate marker. A decrease in circulating tumor cells is also a possible endpoint and several studies in lung, breast and castration-resistant prostate cancer have correlated a decrease in this marker and an improvement in metastatic burden (71). If stage one using a surrogate marker shows a positive outcome, the trial could then move on to stage two, expanding in size and evaluating a more traditional and clinical endpoint such as freedom from metastatic disease (Fig. 2A).

Unlike previous trials of MMPIs, which primarily studied patients with stage IV disease, new trials should enroll patients
with high-risk disease that is not yet clinically or pathologically metastatic, or patients with high-risk precursor lesions. The drug should be given prior to surgery, in the so called “window-of-opportunity” between the time of diagnosis and surgical excision, or postoperatively in the adjuvant setting as microscopic residual disease may not have developed the mutations necessary to fully metastasize (Fig. 2B). While clinical trials with MMPIs have not been conducted in the premetastatic setting, there are ongoing trials of other drugs with a similar design. For example, the ongoing D-Care study is investigating denosumab, a drug already approved for the prevention of pathologic fracture in breast cancer patients with osseous metastasis, in the neoadjuvant or adjuvant setting for patients with stage II or III breast cancer at high risk of recurrence. Primary outcome includes bone-metastasis free survival, which, if positive, would be a successful confirmation of this novel trial design and outcome (72).

As discovered in the past decade, inhibition of tumor growth is largely dependent on the individual MMP targeted, and the mechanism of action of novel MMPIs must be more specific than earlier generations’. Specific inhibitors have recently been and are currently being developed. The mAbs discussed above are perhaps the most promising of the new MMPIs. Ideally, personalized therapies can be envisaged in which only MMPs expressed by the individual tumor are targeted. Such an approach is now feasible thanks to multiple techniques that allow the analysis of gene expression in few, or even single cells that can be identified by laser capture microscopy.

Since the failure of the last trials in the mid-2000s, compounds have been shelved and trials have been on hold. Given the enormous costs of drug development, most manufacturers have been hesitant to reopen the door on trialing MMPIs and a search of active clinical trials in the US yields few results (73). While a few pharmaceutical companies are beginning to trial highly selective MMPIs, these trials are still being conducted in the metastatic setting, and it remains unclear what benefit may be gained by this approach (74, 75). Clearly, a culture shift is needed if the true effects of MMPIs are to be revealed. A first step may be to perform “window of opportunity” trials in early cancers, identifying and validating biomarkers of enzymatic inhibition and metastasis as proxy for clinical success.

Disclosure of Potential Conflicts of Interest
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