

Minireview

Wnt/Planar cell polarity signaling: A new paradigm for cancer therapy

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Abstract

The evolutionarily conserved and developmentally important Wnt signaling pathway has traditionally been regarded as a critical player in tumorigenesis through the canonical Wnt/ β -catenin cascade. Nevertheless, accumulating evidence based on recent research has revealed the previously unacknowledged role of noncanonical Wnt/planar cell polarity (PCP) signaling in cancer progression, invasion and metastasis, and angiogenesis. This review describes the PCP signaling pathway and its ever-expanding components and modulators, highlights the most recent studies that provide insight into the link between PCP signaling and cancer, and, finally, proposes a model by which PCP signaling may promote cancer development. This review underscores the emerging theme that deregulated PCP signaling contributes to tumorigenesis, providing new potential targets for cancer therapy. [Mol Cancer Ther 2009;8(8):2103–9]

Introduction

The Wnt signaling pathway is highly conserved in eukaryotes and is one of the most important signaling pathways. It is used extensively during animal development to regulate diverse processes including cell proliferation, differentiation, polarity, and migration, which are fundamental to embryogenesis. Additionally, deregulation of Wnt signaling has been implicated in a broad range of pathological processes including degenerative diseases and cancer (1). Despite the complexity of this pathway, a combination of developmental, genetic, and biochemical analyses have greatly enriched our understanding of the Wnt pathway, and the list of Wnt signaling branches and

components has exploded (2). It is evident that multiple extracellular, cytoplasmic, and nuclear regulators intricately modulate Wnt signaling. Wnt signals are transduced by at least two distinct pathways: the well-established canonical Wnt/ β -catenin pathway and the β -catenin independent noncanonical Wnt pathway. Noncanonical Wnt signaling is very diverse and is still evolving into more and more branches (3). Among them, Wnt/ Ca^{2+} signaling and Wnt/planar cell polarity (PCP) signaling are relatively better characterized. In Wnt/ Ca^{2+} signaling, Wnt-Frizzled (Fzd) binding activates phospholipase C via G proteins, leading to the increase of intracellular Ca^{2+} levels and activation of downstream effectors including protein kinase C (PKC). The Wnt/ Ca^{2+} pathway plays an important role in development and is implicated in cancer (4). For example, Wnt5a mediates melanoma metastasis via the induction of epithelial to mesenchymal transition (EMT) through the PKC/ Ca^{2+} cascade (5).

Wnt/PCP signaling is the first described and most extensively studied among the several noncanonical Wnt pathways (6, 7). *Drosophila* serves as an excellent model system for PCP signaling because planar cell polarity occurs visibly in several external structures, including the precisely aligned hairs on wing cells, the perfectly arranged ommatidia in the facet eye, and the bristles on the thorax. In fact the term “planar cell polarity” is derived from the study of tissue polarity necessary to generate polarization within the plane of the epithelium, along an axis perpendicular to the apical-basal axis of the cell (8). Subsequently, planar cell polarity was also shown in several cellular processes in vertebrates such as convergent extension (CE) movements of mesenchymal cells during gastrulation, ordered arrangement of hairs of mammalian skin and cilia of respiratory tract, orientation of stereocilia in the organ of Corti, and orientation of axon extension (9).

PCP Signaling Pathway

Genetic screens in *Drosophila* and subsequent work in vertebrates have helped establish that the PCP pathway is evolutionarily conserved and constitutes three sequential steps (10). Initially, upstream PCP components provide a long range signal for the global direction of polarity across the entire tissue. These components include Four-jointed, Dachous, Fat and Atrophin (11). This directional signal is then interpreted by a set of core PCP components to establish planar polarity within individual cells along the axis of polarity. Fzd, Dishevelled (Dvl), Celsr, Vangl, and Prickle

Received 3/26/09; revised 5/15/09; accepted 6/8/09; published OnlineFirst 8/11/09.

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doi:10.1158/1535-7163.MCT-09-0282

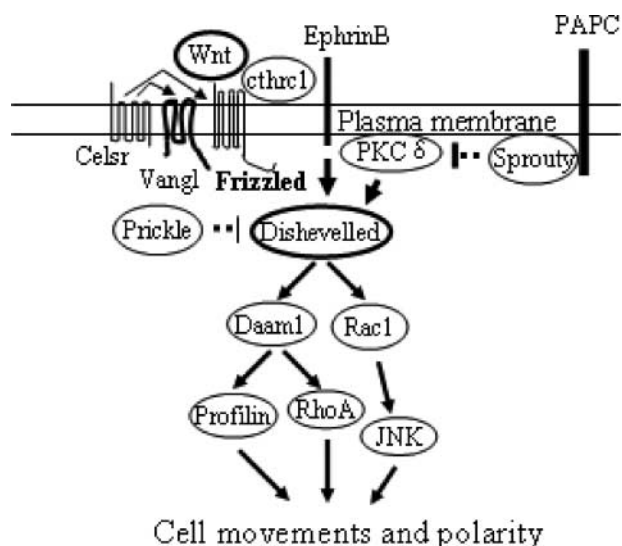


Figure 1. PCP signaling in vertebrates. Noncanonical-Wnt (Wnt5a, Wnt11) binds 7-transmembrane Fzd receptor and activates the recruitment of cytoplasmic scaffold protein Dvl to the plasma membrane. Downstream of Dvl, diverse pathways regulate different aspects of cytoskeleton reorganization in cell movements and polarity. Daam1 binds Dvl by its carboxyl terminus and RhoA by its amino terminus, leading to the formation of Dvl-RhoA complex. Profilin1 is a new interacting partner of Daam1 and localized with Daam1 to actin stress fibers in response to Wnt signal. Dvl-Rac1 complex activates JNK. PCP signaling is regulated at multiple levels. 7-transmembrane protocadherin Celsr regulates the asymmetrical localization of Fzd and 4-transmembrane protein Vangl, contributing to the maintenance of planar polarity. The cytoplasmic tail of Vangl binds and recruits Dvl to plasma membrane, where Prickle binds Dvl and antagonizes Dvl recruitment by Fzd, thus inhibiting PCP signaling. Cthrc1, a secreted glycoprotein, stabilizes the Wnt-Fzd complex to promote PCP signaling. EphrinB1 also positively regulates PCP signaling. The cytoplasmic tail of EphrinB1 binds Dvl and activates RhoA. Dvl recruitment by Fzd is also regulated by kinases including PKC δ , which promote PCP signaling. In contrast, cytoplasmic protein Sprouty antagonizes PCP signaling by inhibiting PKC δ activation, Dvl recruitment by Fzd, and downstream RhoA activation. Protocadherin PAPP can sequester and inhibit Sprouty via direct binding of Sprouty with its cytoplasmic tail, serving as a positive modulator of PCP signaling.

are listed in this group. Finally, tissue-specific downstream PCP effectors convert the upstream signals mediated by upstream and core PCP components into specific morphogenetic programs in individual tissues to generate the distinct planar polarity phenotypes. Daam1, Rho, Rac, Rho kinase, JNK, and Profilin are the primary members of this group that are engaged in the regulation of cytoskeleton to execute cell polarity and cell movements. Rho GTPases including Rho and Rac are extremely important in mediating downstream effects of Wnt signals (12). Taken together, PCP pathway components transduce extracellular polarity cues and integrate a myriad of extracellular and intracellular inputs to induce intracellular cytoskeleton rearrangements and impact cell behaviors (Fig. 1).

Fzd mediates the polarity cue both in *Drosophila* and in vertebrates. Although no Wnt proteins seem to be engaged in PCP signaling in *Drosophila* (2), in vertebrates PCP is mediated by noncanonical Wnts, such as Wnt5 and Wnt11, which do not signal by means of β -catenin (13, 14). The remainder of this review will concentrate on

PCP signaling in vertebrates and how it relates to human cancer development. The PCP signaling cascade outlined in Fig. 1 and Table 1 provides an overview of vertebrate PCP components, their function in PCP signaling, and their involvement in cancer. In addition, it is worth noting that a variety of proteins have been identified to modulate PCP signaling in vertebrates and they are included in Table 1 as well (see also Fig. 1).

PCP Signaling and Cancer

With a growing number of molecules in PCP signaling identified, our understanding of PCP signaling has exploded in the past several years. The role of PCP signaling is critical in multiple embryonic processes including embryo morphogenesis and the formation of most embryonic tissues and organs (15–17). It is noteworthy that embryonic development shares many similarities with cancer development. In the 19th century, Lobstein and Cohnheim speculated that tumorigenesis recapitulates aspects of development (18). In 1982 Nusse and Varmus made the groundbreaking discovery that Wnt1 is proto-oncogene activated in mouse mammary tumors prior to the discovery of its role in embryonic development over the following two decades (19). In fact, conserved signaling pathways involved in embryogenesis including Wnt, Hedgehog, and Notch are dysregulated in tumorigenesis, supporting the theory that cancer is a manifestation of development gone awry (20). Although canonical Wnt/ β -catenin signaling is a critical player in tumorigenesis and β -catenin has gained recognition as an important target for cancer therapy (21), accumulating evidence draws our attention to the previously unappreciated role of PCP signaling in cancer development (22, 23). Summarized below are recent studies suggesting that PCP signaling is closely linked to cancer and plays diverse roles in tumorigenesis.

Role of PCP Signaling in Tumor Metastasis

Metastasis accounts for about 90% of all deaths from cancer. As tumors progress, cancer cells develop the ability to invade through surrounding tissues and basement membranes to form secondary tumors at sites distinct from the primary tumor. Metastatic tumor formation consists of a series of discrete processes. An improved understanding of molecular basis of metastasis is of vital importance to develop effective targeted therapy (24). Although the molecular mechanisms involved in metastasis are still incompletely understood, the factors associated with cell adhesion and cell migration are commonly accepted to be critical in tumor invasion and metastasis (25, 26). Given the central role of PCP signaling in the modulation of cell adhesion, motility, and movement in diverse developmental morphogenetic contexts, it comes as no surprise that dysregulated PCP signaling is implicated in tumor metastasis.

Characterization of a variety of PCP components and modulators has shown their involvement in metastasis. Wnt5a is a typical noncanonical Wnt that transduces β -catenin independent signals in most cases; although, it can

activate β -catenin signaling depending on receptor context (2). Wnt5a has been shown to promote metastasis of melanoma, gastric cancer, and breast cancer by activating Rac and JNK (27–29). Among the 10 members of the Fzd gene family, Fzd7 is unique in that it can activate different branches of Wnt signaling. In *Xenopus*, Fzd7 can activate noncanonical Wnt signaling in morphogenesis, and Fzd7-mediated PCP signaling regulates cell movements via activation of JNK and Rho (30–32). Fzd7 also mediates PCP signaling to regulate bone morphogenesis in chick (33). In humans, Fzd7 promotes migration of hepatocellular carcinoma and invasion of colon cancer cell lines, which seems to be mediated by noncanonical Wnt signaling (34–36). Recently, Fzd10 was shown to play a critical role

in the metastasis of synovial sarcoma. Mechanistically, Fzd10 activates the Dvl-Rac1-JNK axis of PCP signaling, with no effect on nuclear β -catenin accumulation and Tcf/Lef reporter activity (37). Downstream of Fzd, three Dvl members (Dvl1, Dvl2, and Dvl3) have been identified in human. A recent study investigating the clinical significance of Dvl expression in non-small cell lung cancer (NSCLC) specimens showed that the expression of Dvl1 and Dvl3 are significantly higher in nodal metastases than primary tumors. Intriguingly, although Dvl1 expression is correlated with β -catenin expression in the metastases, there is no correlation between Dvl3 and β -catenin expression in primary tumors or metastases. Corroborating these histological data, exogenous expression of Dvl1 and Dvl3 each promotes the

Table 1. Vertebrate PCP components and modulators, their role in PCP signaling, and their involvement in cancer

Gene	Structure	Function in PCP	Involvement in cancer
Upstream PCP components			
<i>Wnt 5a,11</i>	Secreted glycoprotein	Initiate PCP signaling	Promote or inhibit cancer (context dependent)
<i>Fat</i>	Protocadherin	Cell adhesion and signaling	Deletion or silencing in oral squamous carcinoma
<i>Dachsous</i>	Protocadherin	Cell adhesion and signaling	Tumor suppressor gene
<i>Four-jointed</i>	Golgi kinase	Phosphorylate cadherin domains of Fat and Dachsous	Unknown
<i>Atrophin</i>	Transcriptional corepressor	Bind cytoplasmic tail of Fat	Altered expression in neuroblastoma
Core PCP components			
<i>Frizzled</i>	7-Transmembrane receptor	Recruit Dvl to membrane upon binding Wnt	Upregulated in cancer
<i>Dishevelled</i>	Cytoplasmic protein	Recruited to membrane by Fzd	Upregulated in cancer
<i>Vangl</i>	4 -Transmembrane protein	Bind Dvl and PKC δ	High expression in metastatic tissue
<i>Celsr</i>	7-Transmembrane protocadherin	Regulate asymmetrical localization of Fzd	Unknown
<i>Prickle</i>	Cytoplasmic protein	Antagonize Dvl recruitment by Fzd	Downregulated in hepatocellular carcinoma
Downstream PCP effectors			
<i>Daam1</i>	Formin	Bridge Dvl and RhoA	Unknown
<i>RhoA</i>	Small GTPase	Regulate cytoskeleton downstream of Daam1	Upregulated in cancer
<i>Rac</i>	Small GTPase	Regulate cytoskeleton downstream of Dvl	Upregulated in cancer
<i>JNK</i>	S/T protein kinase	Regulate cytoskeleton downstream of Rac	Upregulated in cancer
<i>Rock</i>	Rho kinase	Regulate cytoskeleton downstream of Rho	Upregulated in cancer
<i>Profilin</i>	G-actin-binding protein	Bind DAAM1	Low expression in breast, pancreatic, and hepatic cancer
PCP Modulators			
<i>Paraxial protocadherin</i>	Protocadherin	Promote Dvl recruitment to membrane	Downregulated in breast cancer
<i>EphrinB1</i>	Transmembrane ligand for Eph	Bind Dvl and activate Rho	Promote cancer cell invasion
<i>PKCδ</i>	PKC kinase	Bind and activate Dvl for membrane recruitment	Breast cancer progression
<i>Sprouty1</i>	Cytoplasmic protein	Inhibit Dvl recruitment to membrane	Tumor suppressor gene
<i>Cthrc1</i>	Secreted glycoprotein	Form Cthrc1-Wnt-Fzd complex to promote PCP	Upregulated in invasive cancer
<i>Ptk7</i>	Transmembrane protein tyrosine pseudokinase	Cooperate with Fzd to localize Dvl	Expressed in cancer

invasive ability of lung cancer cells but has differential effects on β -catenin protein levels (38). This suggests that metastasis of NSCLS may be mediated not only by Dvl1 via the canonical Wnt pathway, but also by Dvl3 in a β -catenin independent manner. Further analysis is needed to clarify whether Dvl3 mediates NSCLS metastasis through PCP pathway or other noncanonical Wnt pathways like the Wnt/ Ca^{2+} pathway.

Van Gogh-like 1 (Vangl1) is the human homolog of *Drosophila* PCP gene van gogh/strabismus. Suppression of Vangl1 expression inhibits growth of hepatocellular carcinoma (39). Interestingly, later this 4-transmembrane protein was identified as a binding partner of metastasis suppressor KAI1/CD82 and renamed as KAI1 COOH-terminal interacting tetraspanin (KITENIN) (ref. 40). Vangl1 promotes mouse colon cancer and squamous cancer metastasis, and its expression is higher in tumor mucosa and metastatic tissues from gastric cancer patients and metastatic lymph nodes from head and neck squamous cell carcinoma (40, 41). Moreover, loss of Vangl1 by siRNA suppresses colon cancer metastasis in mice, confirming the metastasis-promoting function of Vangl1 (42). Vangl1 promotes colorectal cancer cell migration and invasion by forming a functional complex with Dvl and PKC δ to modulate cell motility (43). This study suggests that upregulation of Vangl1 takes over PCP signaling to promote tumor invasion and metastasis.

Eph receptors are the largest family of receptor tyrosine kinases. Their ligands, ephrins are membrane-anchored proteins that are classified as glycosylphosphatidylinositol-anchored ephrinA and transmembrane ephrinB. Eph/ephrin signaling plays critical roles not only in embryonic development, but also in the homeostasis of adult organs by regulating a variety of cellular processes such as cell adhesion versus repulsion, cell motility and migration, and cell-cell communication (44). This may be partly due to the unique feature of the Eph/ephrin system to initiate bidirectional signaling that impacts not only Eph receptor-expressing cells, but also ephrin ligand-expressing cells. In particular, reverse signaling transduced via the intracellular domain of ephrinB can regulate the interaction of the C-terminal PDZ-binding motif of ephrinB with other proteins affecting cytoskeletal organization and cell adhesion. Members of the ephrinB family are often overexpressed in cancer cells and are associated with angiogenesis and metastasis (45). Although several studies provide insight into how ephrinB1-mediated reverse signaling contributes to invasion of gastric and pancreas cancer cells (46, 47), recent evidence reveals that intracellular domain of ephrinB1 interacts with Dvl and modulates PCP signaling to control cell adhesion and movement in development (48–50). This opens up the possibility to explore the potential crosstalk between ephrinB and PCP signaling in the context of tumor cell invasion and metastasis.

Collagen triple helix repeat containing protein 1 (Cthrc1) is a secreted glycoprotein containing a short collagen-like motif with 12 Gly-X-Y repeats. Cthrc1 was first identified as a gene transiently expressed in the arterial wall upon injury, possibly supporting vascular remodeling by limiting collagen matrix deposition and promoting cell migration

(51). Cthrc1 is aberrantly upregulated in the majority of human solid tumors and seems to be associated with cancer invasion and metastasis (52, 53). Intriguingly, Cthrc1 was recently characterized as a novel Wnt coreceptor that specifically activates PCP signaling. Cthrc1 knockout synergizes with Vangl2 mutation leading to PCP defects in mice. At cellular level, Cthrc1 synergizes with Wnt5a or Dvl2 to activate PCP downstream effectors RhoA and Rac. Biochemical study shows that Cthrc1 promotes the clustering of Wnt/Fzd/Ror2 complex, providing convincing evidence that Cthrc1 is specifically engaged in PCP signaling (54). Because the molecular mechanism by which Cthrc1 promotes cell migration remains elusive, it is tempting to investigate whether abnormal modulation of PCP signaling by Cthrc1 contributes to invasiveness of Cthrc1-overexpressing tumor cells.

Role of PCP Signaling in Angiogenesis

Angiogenesis, the formation of new blood vessels, is essential not only for the growth of primary tumors but also for tumor metastasis. Angiogenesis facilitates metastasis by increasing the chance that tumor cells enter the blood circulation and by providing nutrients and oxygen for growth at the metastatic sites. Angiogenesis is a complex process involving endothelial cell proliferation along with a myriad steps in which endothelial cells divide, invade the basement membrane, migrate, and eventually undergo differentiation and capillary tube formation (55). Wnts have emerged as a novel class of angiogenic factors that promote angiogenesis, and recent evidence indicates that both Wnt/ β -catenin and noncanonical Wnt signaling are involved in angiogenesis (56, 57). Using a chemical genetic approach, the potent anti-angiogenic natural product fumagillin was identified as a specific inhibitor of noncanonical Wnt signaling, suggesting a link between noncanonical Wnt signaling and angiogenesis (58). Further study showed that inhibition of PCP signaling by the fumagillin analog TNP-470 disrupts endothelial cell growth, polarity, and migration. Furthermore, Dvl2 mutants capable of mediating canonical Wnt/ β -catenin but that are deficient in PCP signaling also disrupt endothelial cell growth, polarity, and migration. Interestingly, Δ DIX-Dvl2, a Dvl mutant that selectively abrogates Wnt/ β -catenin signaling, and the PCP downstream effectors Daam1, diversin, or inversin can rescue the defects caused by TNP-470. Moreover, Wnt5 mutant zebrafish with impaired PCP signaling display angiogenesis defects such as defective intersegmental vessels, reduced dorsal aortas, and reduced posterior cardinal veins (59). Taken together, these findings provide strong support for the emerging role of PCP signaling in angiogenesis.

Complex Role of PCP Signaling in Cancer Development

Although the large body of evidence discussed above delineates the significant involvement of PCP signaling in tumor invasion, metastasis, and angiogenesis, the role of PCP signaling in cancer development is still controversial.

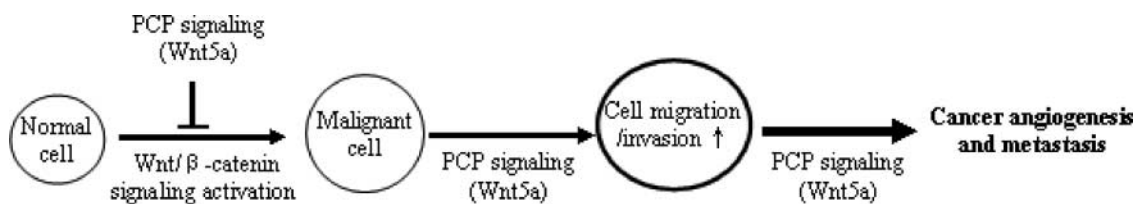


Figure 2. Complex roles of PCP signaling in cancer development. At early stages of cancer, aberrantly activated Wnt/ β -catenin signaling transforms normal cells. PCP signaling (mediated by Wnt5a) inhibits cancer progression by antagonizing Wnt/ β -catenin signaling. However, tumors develop as cells escape the inhibition of PCP signaling through downregulation of PCP signaling or upregulation of downstream effectors of Wnt/ β -catenin signaling. As tumors progress, PCP signaling gets activated and promotes tumor cell migration and invasion and supports angiogenesis, contributing to metastasis in late stages of cancer.

One would expect that PCP signaling can suppress tumorigenesis given the well-recognized upregulation of Wnt/ β -catenin signaling in tumorigenesis and the documented antagonism between canonical and noncanonical Wnt signaling (60). Indeed, several components of the PCP signaling pathway can suppress tumor progression, including the protocadherin family members Fat, Dachshous, and Paraxial protocadherin (PAPC). Although the basic outline emerges that Fat and Dachshous act as tumor suppressors by engaging downstream signaling networks to regulate transcription and control cell growth, it is still an open question to determine the extent to which control of cell polarity by Fat. PCP signaling is involved in the tumor suppression function of Fat (61). PAPC can promote PCP signaling by sequestration of Sprouty with its cytoplasmic tail (62). Given that Sprouty negatively regulates Ras signaling and is frequently downregulated in cancer (63), PAPC might enhance tumor development by antagonizing Sprouty and promoting PCP signaling. Contrary to this prediction, protocadherin-8 (PCDH8), the human ortholog of PAPC, has tumor suppression function and is inactivated through either mutation or epigenetic silencing in about one third of all breast carcinomas (64). It is conceptually simple to assign the tumor suppressor ability of PCDH8 to the adhesion activity mediated by the extracellular domain of PCDH8 given that most of PCDH8 mutations found in breast tumors reside at the extracellular cadherin repeat. However, one PCDH8 mutation is localized in the intracellular domain, implying that intracellular signaling may be also important for tumor suppression function of PCDH8 (64). Further investigation is needed to address the status of PCP signaling in PCDH8 inactivated breast cancer. Moreover, the tumor suppressor or oncogenic activity of PCDH8 should also be surveyed in other tumors given that another protocadherin protocadherin-PC is implicated in the progression of prostate cancer (65).

Noncanonical Wnt5a acts as an oncogene or tumor suppressor gene in a context-dependent manner. Antisense Wnt5a mimics Wnt1-mediated mammary epithelial cell transformation (66). Wnt5a is downregulated in hematopoietic malignancies, thyroid carcinoma, breast cancer, colorectal cancer, and hepatocellular carcinoma, and in some cases there is an inverse correlation between Wnt5a level and β -catenin level (67–72). Taken together, these studies support the tumor suppressor function of Wnt5a by antagonizing

Wnt/ β -catenin signaling. Nevertheless, in other contexts, especially in cancers of advanced stages, Wnt5a is upregulated and has oncogenic properties promoting tumor invasion, metastasis, and angiogenesis as discussed in previous sections. In this aspect, PCP signaling is analogous to transforming growth factor β (TGF β) signaling, which plays multiple roles in cancer development with early tumor suppressive effects through growth inhibition but later tumorigenic effects, including increased tumor cell motility and invasion and induction of angiogenesis (73). Therefore, it is appealing to propose a model in which PCP signaling has biphasic role in carcinogenesis by acting early as tumor suppressor but later stimulating cancer progression via the regulation of tumor invasion, metastasis, and angiogenesis (Fig. 2).

Modulation of PCP Signaling for Cancer Therapy

This review makes the first attempt to comprehensively cover the impact of PCP signaling on different features of cancer development and outlines an emerging picture that PCP signaling plays important roles in tumorigenesis. Therefore, it is critical for us to take advantage of this new understanding to develop novel diagnostic and therapeutic approaches for cancer. In fact, some components and modulators of PCP signaling such as Vangl1, EphrinB, and Cthrc1 have the potential to serve as diagnostic biomarkers for advanced metastatic tumors, given that their expression levels are higher in malignant tumors and are correlated with reduced patient survival rates (41, 45, 52, 53). In addition, inhibition of the function of PCP signaling molecules Vangl1 by siRNA may suppress colon cancer progression and metastasis (42), supporting the targeting of PCP signaling components as promising cancer therapies. Compared with canonical Wnt/ β -catenin signaling, which plays a crucial physiologic role not only in embryonic development but also in the maintenance of adult tissue homeostasis, current research suggests that PCP signaling functions primarily in embryo morphogenesis. Thus, modulation of PCP signaling in cancer patients is less likely to interfere with homeostasis and represents a better avenue for cancer therapy.

Nevertheless, several issues must be sorted out in order to conceptualize effective therapeutic strategies for targeting PCP signaling. First, unlike the field of canonical

Wnt/ β -catenin signaling in which a straightforward readout system has been used to identify small molecule compounds targeting this pathway (74), the field of PCP signaling lacks a well-established pathway readout system, hindering the utilization of small molecule screening to discover compounds that specifically modulate PCP signaling for cancer therapy. Fortunately, TNP-470 was serendipitously identified as the first small molecule capable of specifically inhibiting PCP signaling (58, 59). This example provides guidance for future small molecule screening for PCP signaling modulators using zebrafish high resolution phenotyping resources until a better PCP pathway readout system is developed. Second, some components of PCP signaling have multifaceted functions. For example, unrelated to its function in PCP signaling, Fat has tumor suppressor function through regulation of transcription and cell growth. Undoubtedly, it will benefit cancer patients if we can target downstream effects of Fat PCP signaling while preserving its tumor suppression function. Third, tumors have recognized abilities to acquire resistance to therapeutics, so combined therapy is absolutely necessary. It is well acknowledged that PCP signaling can crosstalk with other pathways including canonical Wnt/ β -catenin signaling, bone morphogenetic proteins (BMP) signaling, and fibroblast growth factor (FGF) signaling to regulate embryo morphogenesis (15). Therefore, it is worth investigating their crosstalk in the context of cancer development to develop potent cancer therapy strategies.

In conclusion, we are now in a good position to approach cancer from the perspective of developmental biology, which will continue to provide valuable insights into the mechanisms and roles of PCP signaling. Realizing that the role of PCP signaling in cancer is a budding area, there is no doubt that in-depth dissection of the role of PCP signaling in cancer development will uncover novel and powerful therapeutic approaches for cancer treatment in the coming years.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

Acknowledgments

I thank Dr. Rachel K. Miller for critical reading of the manuscript.

References

- Logan CY, Nusse R. The Wnt signaling pathway in development and disease. *Annu Rev Cell Dev Biol* 2004;20:781–810.
- van Amerongen R, Mikels A, Nusse R. Alternative wnt signaling is initiated by distinct receptors. *Sci Signal* 2008;1:re9.
- Semenov MV, Habas R, Macdonald BT, He X. SnapShot: Noncanonical Wnt Signaling Pathways. *Cell* 2007;131:1378.
- Kohn AD, Moon RT. Wnt and calcium signaling: β -catenin-independent pathways. *Cell Calcium* 2005;38:439–46.
- Dissanayake SK, Wade M, Johnson CE, et al. The Wnt5A/protein kinase C pathway mediates motility in melanoma cells via the inhibition of metastasis suppressors and initiation of an epithelial to mesenchymal transition. *J Biol Chem* 2007;282:17259–71.
- Simons M, Mlodzik M. Planar cell polarity signaling: from fly development to human disease. *Annu Rev Genet* 2008;42:517–40.
- Zallen JA. Planar polarity and tissue morphogenesis. *Cell* 2007;129:1051–63.
- Nubler-Jung K. Insect epidermis: disturbance of supracellular tissue polarity does not prevent the expression of cell polarity. *Dev Genes Evol* 1987;196:286–9.
- Goodrich LV. The plane facts of PCP in the CNS. *Neuron* 2008;60:9–16.
- Tree DR, Ma D, Axelrod JD. A three-tiered mechanism for regulation of planar cell polarity. *Semin Cell Dev Biol* 2002;13:217–24.
- Strutt H, Strutt D. Long-range coordination of planar polarity in *Drosophila*. *Bioessays* 2005;27:1218–27.
- Schlessinger K, Hall A, Tolwinski N. Wnt signaling pathways meet Rho GTPases. *Genes Dev* 2009;23:265–77.
- Moon RT, Campbell RM, Christian JL, McGrew LL, Shih J, Fraser S. Xwnt-5A: a maternal Wnt that affects morphogenetic movements after overexpression in embryos of *Xenopus laevis*. *Development* 1993;119:97–111.
- Heisenberg CP, Tada M, Rauch GJ, et al. Silberblick/Wnt11 mediates convergent extension movements during zebrafish gastrulation. *Nature* 2000;405:76–81.
- Wang Y, Steinbeisser H. Molecular basis of morphogenesis during vertebrate gastrulation. *Cell Mol Life Sci* 2009;66:2263–73.
- Karner C, Wharton KA, Jr., Carroll TJ. Planar cell polarity and vertebrate organogenesis. *Semin Cell Dev Biol* 2006;17:194–203.
- Barrow JR. Wnt/PCP signaling: a veritable polar star in establishing patterns of polarity in embryonic tissues. *Semin Cell Dev Biol* 2006;17:185–93.
- Rather LJ. The genesis of cancer: A study in the history of ideas. Baltimore (MD): Johns Hopkins University Press; 1978.
- Nusse R, Varmus HE. Many tumors induced by the mouse mammary tumor virus contain a provirus integrated in the same region of the host genome. *Cell* 1982;31:99–109.
- Kho AT, Zhao Q, Cai Z, et al. Conserved mechanisms across development and tumorigenesis revealed by a mouse development perspective of human cancers. *Genes Dev* 2004;18:629–40.
- Takemaru KI, Ohmitsu M, Li FQ. An oncogenic hub: β -catenin as a molecular target for cancer therapeutics. *Handb Exp Pharmacol* 2008;186:261–84.
- Katoh M. WNT/PCP signaling pathway and human cancer [review]. *Oncol Rep* 2005;14:1583–8.
- Kikuchi A, Yamamoto H. Tumor formation due to abnormalities in the β -catenin-independent pathway of Wnt signaling. *Cancer Sci* 2008;99:202–8.
- Chiang AC, Massague J. Molecular basis of metastasis. *N Engl J Med* 2008;359:2814–23.
- Christofori G. New signals from the invasive front. *Nature* 2006;441:444–50.
- Yamaguchi H, Wyckoff J, Condeelis J. Cell migration in tumors. *Curr Opin Cell Biol* 2005;17:559–64.
- Weeraratna AT, Jiang Y, Hostetter G, et al. Wnt5a signaling directly affects cell motility and invasion of metastatic melanoma. *Cancer Cell* 2002;1:279–88.
- Kurayoshi M, Oue N, Yamamoto H, et al. Expression of Wnt-5a is correlated with aggressiveness of gastric cancer by stimulating cell migration and invasion. *Cancer Res* 2006;66:10439–48.
- Pukrop T, Klemm F, Hagemann T, et al. Wnt 5a signaling is critical for macrophage-induced invasion of breast cancer cell lines. *Proc Natl Acad Sci U S A* 2006;103:5454–9.
- Medina A, Reintsch W, Steinbeisser H. *Xenopus* frizzled 7 can act in canonical and non-canonical Wnt signaling pathways: implications on early patterning and morphogenesis. *Mech Dev* 2000;92:227–37.
- Kinoshita N, Iioka H, Miyakoshi A, Ueno N. PKC δ is essential for Dishelved function in a noncanonical Wnt pathway that regulates *Xenopus* convergent extension movements. *Genes Dev* 2003;17:1663–76.
- Tanegashima K, Zhao H, Dawid IB. WGEF activates Rho in the Wnt-PCP pathway and controls convergent extension in *Xenopus* gastrulation. *EMBO J* 2008;27:606–17.
- Li Y, Dudley AT. Noncanonical frizzled signaling regulates cell polarity of growth plate chondrocytes. *Development* 2009;136:1083–92.
- Merle P, de la Monte S, Kim M, et al. Functional consequences of

frizzled-7 receptor overexpression in human hepatocellular carcinoma. *Gastroenterology* 2004;127:1110–22.

35. Vincan E, Swain RK, Brabletz T, Steinbeisser H. Frizzled7 dictates embryonic morphogenesis: implications for colorectal cancer progression. *Front Biosci* 2007;12:4558–67.
36. Ueno K, Hiura M, Suehiro Y, et al. Frizzled-7 as a potential therapeutic target in colorectal cancer. *Neoplasia* 2008;10:697–705.
37. Fukukawa C, Nagayama S, Tsunoda T, Toguchida J, Nakamura Y, Katagiri T. Activation of the non-canonical Dvl-Rac1-JNK pathway by Frizzled homologue 10 in human synovial sarcoma. *Oncogene* 2009;28:1110–20.
38. Wei Q, Zhao Y, Yang ZQ, et al. Dishevelled family proteins are expressed in non-small cell lung cancer and function differentially on tumor progression. *Lung Cancer* 2008;62:181–92.
39. Yagyu R, Hamamoto R, Furukawa Y, Okabe H, Yamamura T, Nakamura Y. Isolation and characterization of a novel human gene, VANGL1, as a therapeutic target for hepatocellular carcinoma. *Int J Oncol* 2002;20:1173–8.
40. Lee JH, Park SR, Chay KO, et al. KAI1 COOH-terminal interacting tetraspanin (KITENIN), a member of the tetraspanin family, interacts with KAI1, a tumor metastasis suppressor, and enhances metastasis of cancer. *Cancer Res* 2004;64:4235–43.
41. Lee JK, Bae JA, Sun EG, et al. KITENIN increases invasion and migration of mouse squamous cancer cells and promotes pulmonary metastasis in a mouse squamous tumor model. *FEBS Lett* 2009;583:711–7.
42. Lee JH, Cho ES, Kim MY, et al. Suppression of progression and metastasis of established colon tumors in mice by intravenous delivery of short interfering RNA targeting KITENIN, a metastasis-enhancing protein. *Cancer Res* 2005;65:8993–9003.
43. Kho DH, Bae JA, Lee JH, et al. KITENIN recruits Dishevelled/PKC δ to form a functional complex and controls the migration and invasiveness of colorectal cancer cells. *Gut* 2009;58:509–19.
44. Pasquale EB. Eph-ephrin bidirectional signaling in physiology and disease. *Cell* 2008;133:38–52.
45. Campbell TN, Robbins SM. The Eph receptor/ephrin system: an emerging player in the invasion game. *Curr Issues Mol Biol* 2008;10:61–6.
46. Tanaka M, Kamata R, Takigahira M, Yanagihara K, Sakai R. Phosphorylation of ephrin-B1 regulates dissemination of gastric scirrhous carcinoma. *Am J Pathol* 2007;171:68–78.
47. Tanaka M, Sasaki K, Kamata R, Sakai R. The C-terminus of ephrin-B1 regulates metalloproteinase secretion and invasion of cancer cells. *J Cell Sci* 2007;120:2179–89.
48. Tanaka M, Kamo T, Ota S, Sugimura H. Association of Dishevelled with Eph tyrosine kinase receptor and ephrin mediates cell repulsion. *EMBO J* 2003;22:847–58.
49. Lee HS, Mood K, Battu G, Ji YJ, Singh A, Daar IO. Fibroblast growth factor receptor-induced phosphorylation of ephrinB1 modulates its interaction with Dishevelled. *Mol Biol Cell* 2009;20:124–33.
50. Lee HS, Bong YS, Moore KB, Soria K, Moody SA, Daar IO. Dishevelled mediates ephrinB1 signalling in the eye field through the planar cell polarity pathway. *Nat Cell Biol* 2006;8:55–63.
51. Pygay P, Herault M, Wang Q, et al. Collagen triple helix repeat containing 1, a novel secreted protein in injured and diseased arteries, inhibits collagen expression and promotes cell migration. *Circ Res* 2005;96:261–8.
52. Turashvili G, Bouchal J, Baumforth K, et al. Novel markers for differentiation of lobular and ductal invasive breast carcinomas by laser microdissection and microarray analysis. *BMC Cancer* 2007;7:55.
53. Tang L, Dai DL, Su M, Martinka M, Li G, Zhou Y. Aberrant expression of collagen triple helix repeat containing 1 in human solid cancers. *Clin Cancer Res* 2006;12:3716–22.
54. Yamamoto S, Nishimura O, Misaki K, et al. Cthrc1 selectively activates the planar cell polarity pathway of Wnt signaling by stabilizing the Wnt-receptor complex. *Dev Cell* 2008;15:23–36.
55. Takeda A, Stoeltzing O, Ahmad SA, et al. Role of angiogenesis in the development and growth of liver metastasis. *Ann Surg Oncol* 2002;9:610–6.
56. Masckauchan TN, Kitajewski J. Wnt/Frizzled signaling in the vasculature: new angiogenic factors in sight. *Physiology (Bethesda)* 2006;21:181–8.
57. Cheng CW, Yeh JC, Fan TP, Smith SK, Charnock-Jones DS. Wnt5a-mediated non-canonical Wnt signalling regulates human endothelial cell proliferation and migration. *Biochem Biophys Res Commun* 2008;365:285–90.
58. Zhang Y, Yeh JR, Mara A, et al. A chemical and genetic approach to the mode of action of fumagillin. *Chem Biol* 2006;13:1001–9.
59. Cirone P, Lin S, Griesbach HL, Zhang Y, Slusarski DC, Crews CM. A role for planar cell polarity signaling in angiogenesis. *Angiogenesis* 2008;11:347–60.
60. Veeman MT, Axelrod JD, Moon RT. A second canon. Functions and mechanisms of β -catenin-independent Wnt signaling. *Dev Cell* 2003;5:367–77.
61. Reddy BV, Irvine KD. The Fat and Warts signaling pathways: new insights into their regulation, mechanism and conservation. *Development* 2008;135:2827–38.
62. Wang Y, Janicki P, Koster I, et al. Xenopus Paraxial Protocadherin regulates morphogenesis by antagonizing Sprouty. *Genes Dev* 2008;22:878–83.
63. Lo TL, Fong CW, Yusoff P, et al. Sprouty and cancer: the first terms report. *Cancer Lett* 2006;242:141–50.
64. Yu JS, Koujak S, Nagase S, et al. PCDH8, the human homolog of PAPC, is a candidate tumor suppressor of breast cancer. *Oncogene* 2008;27:4657–65.
65. Giannakopoulos X, Stagikas D, Peschos D, Batistatou A, Charalabopoulos K. Implication of protocadherin-PC in the progression of the advanced prostate cancer. *Exp Oncol* 2007;29:74–5.
66. Olson DJ, Gibo DM. Antisense wnt-5a mimics wnt-1-mediated C57MG mammary epithelial cell transformation. *Exp Cell Res* 1998;241:134–41.
67. Liang H, Chen Q, Coles AH, et al. Wnt5a inhibits B cell proliferation and functions as a tumor suppressor in hematopoietic tissue. *Cancer Cell* 2003;4:349–60.
68. Kremenevskaja N, von Wasielewski R, Rao AS, Schofl C, Andersson T, Brabant G. Wnt-5a has tumor suppressor activity in thyroid carcinoma. *Oncogene* 2005;24:2144–54.
69. Leris AC, Roberts TR, Jiang WG, Newbold RF, Mokbel K. WNT5A expression in human breast cancer. *Anticancer Res* 2005;25:731–4.
70. Ying J, Li H, Chen YW, Srivastava G, Gao Z, Tao Q. WNT5A is epigenetically silenced in hematologic malignancies and inhibits leukemia cell growth as a tumor suppressor. *Blood* 2007;110:4130–2.
71. Ying J, Li H, Yu J, et al. WNT5A exhibits tumor-suppressive activity through antagonizing the Wnt/ β -catenin signaling, and is frequently methylated in colorectal cancer. *Clin Cancer Res* 2008;14:55–61.
72. Liu XH, Pan MH, Lu ZF, et al. Expression of Wnt-5a and its clinicopathological significance in hepatocellular carcinoma. *Dig Liver Dis* 2008;40:560–7.
73. Akhurst RJ, Derynck R. TGF- β signaling in cancer—a double-edged sword. *Trends Cell Biol* 2001;11:S44–51.
74. Chen B, Dodge ME, Tang W, et al. Small molecule-mediated disruption of Wnt-dependent signaling in tissue regeneration and cancer. *Nat Chem Biol* 2009;5:100–7.

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Mol Cancer Ther 2009;8:2103-2109. Published OnlineFirst August 11, 2009.

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