

The rice bran constituent triclin potently inhibits cyclooxygenase enzymes and interferes with intestinal carcinogenesis in *Apc^{Min}* mice

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Abstract

While brown rice is a staple dietary constituent in Asia, rice consumed in the Western world is generally white, obtained from brown rice by removal of the bran. Rice bran contains the flavone triclin, which has been shown to inhibit colon cancer cell growth. We tested the hypothesis that triclin interferes with adenoma formation in the *Apc^{Min}* mouse. Mice received triclin (0.2%) in their American Institute of Nutrition 93G diet throughout their postweaning life span (4–18 weeks). Consumption of triclin reduced numbers of intestinal adenomas by 33% ($P < 0.05$) compared with mice on control diet. We explored whether triclin may exert its effect via inhibition of cyclooxygenase (COX) enzymes. Its effect on COX activity was assessed in purified enzyme preparations *in vitro* and its ability to reduce prostaglandin E₂ (PGE₂) levels in human colon-derived human colon epithelial cell (HCEC) and HCA-7 cells *in vitro* and in *Apc^{Min}* mice *in vivo*. Triclin inhibited activity of purified COX-1 and COX-2 enzyme preparations with IC₅₀ values of ~1 μmol/L. At 5 μmol/L, it reduced PGE₂ production in HCEC or HCA-7 cells by 36% ($P < 0.01$) and 35% ($P < 0.05$), respectively. COX-2 expression was reduced by triclin weakly in HCEC and unaffected in HCA-7 cells. PGE₂ levels in the small intestinal mucosa and blood of *Apc^{Min}* mice that had received triclin were reduced by 34% ($P < 0.01$) and

40% ($P < 0.05$), respectively, compared with control mice. The results suggest that triclin should be further evaluated as a putative colorectal cancer chemopreventive agent. [Mol Cancer Ther 2005;4(9):1287–92]

Introduction

The isolation of constituents of the diet for preclinical and clinical evaluation of their potential cancer chemopreventive properties is a promising strategy in the discovery of novel cancer chemopreventive interventions (1). Examples of chemopreventive agents that have been found in this fashion are resveratrol and epigallocatechin gallate, contained in red grapes and tea, respectively. Epidemiologic evidence suggests that consumption of whole grain foods is inversely associated with incidence of intestinal adenomatous polyps (2), and the potential chemopreventive effect of whole grains on polyp development has been tentatively related to the presence of anticarcinogenic constituents rather than fiber. Rice, *Oryza sativa*, is the staple food of over half the population of the world. The unpolished brown (bran containing) variety, consumption of which is high in Asia, contains several agents suspected of possessing cancer chemopreventive properties, e.g., triclin (4',5,7-trihydroxy-3',5'-dimethoxyflavone; for structure, see Fig. 1), which are completely absent from the white variety. Rice consumed in the Western world is mainly the white variety, obtained from brown rice by removal of the bran. Dietary differences such as this may explain why the incidence of colorectal cancer in some parts of Asia is much lower than in the Western world (3). Very little is known about the pharmacology of triclin other than that it possesses anticarcinogenic activity in human-derived colon and breast cancer cells *in vitro* (4) and antineoplastic properties in mice with the P388 leukemia (5). Mechanisms by which triclin may exert chemopreventive or antineoplastic activities are unknown.

Rodent models of colorectal carcinogenesis are useful in the discovery of cancer chemopreventive agents because they allow the assessment of effect of interventions on incidence, multiplicity, or volume of preneoplastic (aberrant crypt foci, adenomas) and/or neoplastic lesions. Genetically driven models targeting the *APC* gene, such as the *Apc^{Min}* mouse, a model of the human heritable condition familial adenomatous polyposis coli (6), are particularly relevant to individuals carrying these germline mutations. Although correlations between rodent and human data with respect to efficacy and safety are imperfect, a retrospective analysis of results in rodents and humans obtained with inhibitors of cyclooxygenase (COX) enzymes in colorectal cancer prevention shows reasonable consistency between species (7). In the light of these considerations, we tested the hypothesis

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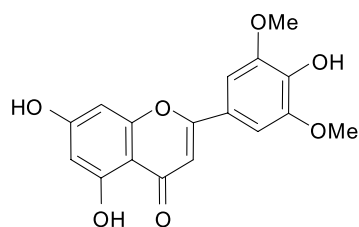


Figure 1. Structure of triclin.

that consumption of triclin attenuates adenoma development in the *Apc^{Min}* mouse. The ability of the selective COX-2 inhibitor celecoxib to decrease adenoma development in the *Apc^{Min}* mouse (8) led to its evaluation in familial adenomatous polyposis coli patients (9), the results of which in turn engendered approval of this agent by the Food and Drug Administration as a treatment for familial adenomatous polyposis coli. COX enzymes catalyze the metabolism of arachidonic acid to prostaglandins, such as prostaglandin E₂ (PGE₂), biomolecules that contribute to the initiation, promotion, or progression of intestinal malignancies (10). Interference with COX activity is considered a crucial mechanism through which nonsteroidal anti-inflammatory drugs and COX-2 inhibitors retard intestinal adenoma development in humans (9, 11) and also in *Apc^{Min}* mice (8, 12). In this context, it is pertinent to note that nonsteroidal anti-inflammatory drugs are of particular interest as cancer chemopreventive agents because they can reduce malignancies in the human colorectum, while in the *Apc^{Min}* mouse model they target predominantly small intestinal adenomas with little effect on colonic polyps. As we found that triclin interferes with intestinal carcinogenesis in *Apc^{Min}* mice, we tested the hypothesis that inhibition of COX activity is a mechanism by which triclin retards adenoma development. Overall, the study was designed to contribute to the body of evidence that will eventually help decide whether triclin should be advanced to clinical evaluation as colorectal cancer chemopreventive intervention.

Materials and Methods

Materials and Animals

Tricin was custom-synthesized by Syncom (Groningen, the Netherlands) for the National Cancer Institute Chemoprevention Branch. Its purity (>99%) was checked by high-performance liquid chromatography analysis (13). C57BL/6J Min/+ (*Apc^{Min}*) mice were bred in the Leicester University Biomedical Services facility using animals originally obtained from The Jackson Laboratory (Bar Harbor, ME).

Experiments in *Apc^{Min}* Mice

Experiments were carried out under animal project license PPL 40/2496, granted to Leicester University by the United Kingdom Home Office. The experimental design was vetted by the Leicester University Local Ethical Committee for Animal Experimentation and met the standards required by the United Kingdom Coordinating Committee on Cancer Research guidelines (14). An

Apc^{Min} breeding colony was established, and the *Apc^{Min}* genotype was confirmed by PCR (6). Groups of 15 to 22 mice (male-to-female ratio, 0.5:1 and 1.2:1 in control and intervention groups, respectively) received standard American Institute of Nutrition 93G diet or American Institute of Nutrition diet supplemented with triclin (0.2% in the diet, translating into a dose of ~6 mg per mouse per day) from ages 4 to 18 weeks. The choice of dose was based on the fact that dietary doses of naturally occurring polyphenols that attenuate colorectal carcinogenesis in preclinical models are generally of this order of magnitude, e.g., curcumin in *Apc^{Min}* mice (15, 16). Animals were killed by cardiac exsanguination (halothane anesthesia), and the intestinal tract was removed and flushed with PBS. Plasma was obtained by centrifugation of blood. Intestinal tissue was prepared as "Swiss roll" and fixed, processed, sectioned, and stained with H&E before microscopic examination. Multiplicity, location, and size of adenomas were recorded as described before (15).

Measurement of COX Activity

Purified COX-2 and COX-1 enzyme preparations from sheep placenta and sheep seminal vesicles, respectively, were purchased from Alexis Biosciences (Lausen, Switzerland). COX activity was assessed in incubations with COX-1 or COX-2 enzyme using a chemiluminescent enzyme activity kit (Assay Designs, Ann Arbor, MI) in which the COX peroxidase reaction is monitored with a cosubstrate generating a chemiluminescent species on COX peroxidation.

Measurement of PGE₂ Levels and COX-2 Protein in Cells

HCA-7 cells, derived from a human mucinous adenocarcinoma of the colon (17), and transformed, but nonmalignant human colon epithelial cells (HCEC), were provided by Drs. S. Kirkland (Hammersmith Hospital, Imperial College, London, United Kingdom) and A. Pfeifer (Nestec, Ltd., Research Centre, Lausanne, Switzerland), respectively. Cells from subculture 20 to 30 were seeded in 90 mm Petri dishes (Nunc, Fisher Scientific, Loughborough, United Kingdom) and grown in DMEM containing Glutamax I, glucose (4.5 g/L), and 10% (v/v) FCS (Life Technologies, Paisley, United Kingdom). HCEC cells were cultured in dishes coated with Vitrogen 100 (10 μL/mL, Collagen Corp., Palo Alto, CA), human fibronectin (2.5 μg/mL; Sigma, Poole, United Kingdom), and bovine serum albumin (50 μg/mL, Life Technologies). HCEC cells (0.6 × 10⁶ cells per Petri dish) were plated, left for 12 hours, and then incubated for 24 hours with tetradecanoyl phorbol acetate (to induce COX-2 expression), arachidonic acid, and triclin essentially as described before for curcuminoids (18). HCA-7 cells (2 × 10⁶ per dish) were incubated with triclin for 24 hours. Aliquots (1 mL) of either cell supernatant were removed for determination of PGE₂ by immunoassay, which was carried out using a PGE₂ immunoassay kit (R&D Systems, Abingdon, United Kingdom). Quantitation was performed using a FLUOstar OPTIMA plate reader (BMG Labtechnologies GmbH, Germany).

For measurement of the effect of triclin on cellular COX-2 protein levels, cells were incubated as described above, but for 6 hours instead of 24 hours; results obtained after 24 hours were similar to those observed for 6 hours. COX-2 was determined by Western blot analysis as previously described (19) using a polyclonal antibody against COX-2 (Santa Cruz Biotechnology, Inc., Santa Cruz, CA). Equal protein loading and transfer were monitored by probing for α -tubulin (Santa Cruz Biotechnology). Semiquantitation, including correction in relation to the respective α -tubulin band, was by densitometric analysis using a Gene Gnome densitometer (Syngene Bio Imaging Systems, Frederick, MD).

Measurement of Levels of PGE₂ and Tricin in Murine Tissue

The small intestine of *Apc^{Min}* mice was removed and mucosa was obtained by scraping with a spatula. Mucosa was homogenized and the homogenate (25% in water) was subjected to solid phase extraction (C₁₈ reverse phase columns, Varian, Oxford, United Kingdom), which involved preconditioning (10 mL ethanol), equilibration (10 mL water), and washing of the column (water, 15% ethanol and hexane, 10 mL each) and analyte elution (ethyl acetate 10 mL). PGE₂ in samples of cellular supernatant, plasma, or column eluate of mucosal homogenate was quantitated using the above PGE₂ immunoassay kit.

Levels of triclin in plasma and intestinal mucosa of *Apc^{Min}* mice, which had received dietary triclin (0.2%), were measured by high-performance liquid chromatography using a method described before (13).

Statistical Evaluation

Evaluation of significance of values compared with the appropriate controls was done by one-way ANOVA with subsequent Tukey's pairwise comparison.

Results

Effect of Tricin on Intestinal Adenoma Development in *Apc^{Min}* Mice

We explored whether dietary triclin affects intestinal adenoma development. *Apc^{Min}* mice received triclin with

their diet from weaning for 14 weeks. Histopathologic analysis of the small intestine showed focal proliferative lesions ranging from small to large areas of glandular hyperplasia (i.e., elongated or branched intestinal glands lined by hyperchromatic cells, often with mucin depletion) and polypoid adenomas, without significant differences in morphology between control mice and mice on the interventions. Numbers of small intestinal adenomas in mice on triclin were reduced by 33% compared with mice on control diet (Fig. 2A). Tricin did not retard numbers of adenoma in the colon (Fig. 2B). A detailed analysis of small intestinal polyp location revealed that polyp reduction by triclin was significant in the proximal intestine (Fig. 2C). As to its effect on differentially sized adenomas, the retarding activity of triclin was significant in large polyps (>3 mm; Fig. 2D). The body weight of mice, which had received triclin, was not significantly different from that of mice on control diet (20), suggesting that the effect on adenoma development was not the consequence of reduced food intake.

Effect of Tricin on COX Activity, PGE₂ Levels in Cells and Mouse Tissues, and COX-2 Levels in Cells

We tested the hypothesis that triclin inhibits COX activity in purified enzyme preparations *in vitro*, in intact colon cells, and in *Apc^{Min}* mice *in vivo*. The effect of triclin on purified enzyme activity was assessed using a kit in which a cosubstrate of the COX peroxidase reaction generates a chemiluminescent species. Tricin inhibited the activity of COX-1 and COX-2 enzymes similarly with an IC₅₀ of ~1 μ mol/L (Fig. 3). The corollary of exposure to triclin for levels of PGE₂ generated by COX-2 catalysis was determined in human colon-derived HCEC cells, in which COX-2 was induced by exposure to phorbol ester, and in HCA-7 cells, which constitutively overexpress COX-2 but hardly any COX-1 (21). At 5 μ mol/L, triclin reduced PGE₂ production in HCEC and HCA-7 cells by 36% and 35%, respectively, compared with cells unexposed to triclin (Fig. 4A and B). Next, the hypothesis was tested that triclin down-regulates cellular COX-2 expression as assessed by Western blot analysis. Tricin did not affect the expression of COX-2 in HCA-7 cells, while in

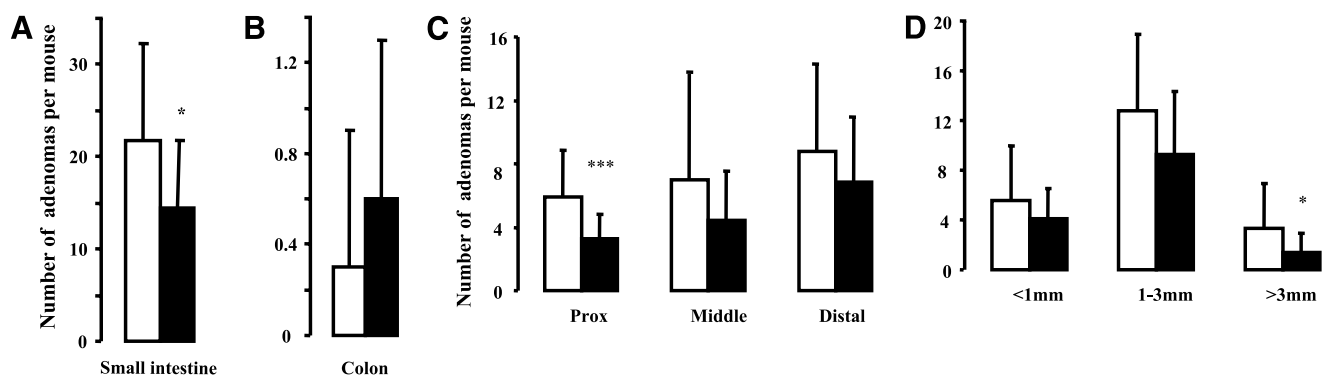


Figure 2. Effects of dietary triclin (0.2%) on adenomas in *Apc^{Min}* mice. Numbers of adenomas (**A–C**) in the small intestine (**A**), the colon (**B**), specific sections in the small intestine [proximal (*prox*), middle, and distal, **C**]. **D**, multiplicity of small (<1 mm diameter), medium-sized (1–3 mm), or large (>3 mm) adenomas. *Open columns*, control mice ($n = 15$); *closed columns*, mice in triclin ($n = 21$). *Columns*, mean; *bars*, SD. Asterisks indicate that values are significantly different from controls (* $P < 0.05$, *** $P < 0.005$). For details of animal experimentation, see Materials and Methods.

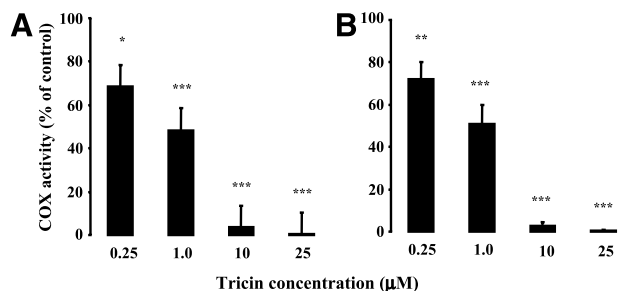


Figure 3. Effect of triclin on activity of purified COX-2 (A) and COX-1 (B) enzyme. Measurement was by a kit in which a cosubstrate of the COX peroxidase reaction generates a chemiluminescent species, as per the protocol of the vendor. Enzyme activity in the control incubates omitting triclin was 100%. Columns, mean (for 0.25 μmol/L, $n = 4$; for 1, 10, and 25 μmol/L in the case of COX-2, $n = 8$; in the case of COX-1, $n = 7$); bars, SD. Asterisks indicate that values were significantly different from controls (* $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$). For details of enzyme assay, see Materials and Methods.

HCEC cells 5 to 40 μmol/L triclin reduced COX-2 protein levels by between 19% and 30% (Fig. 4C and D). Tricin did not alter the expression of COX-1 assessed in HT-29 cells (result not shown). PGE₂ levels in the small intestinal mucosa and blood of mice, which had received triclin, were reduced by 34% and 40%, respectively, compared with mice on control diet (Fig. 5). The source of PGE₂ in the blood of these mice was probably exclusively COX-1, while the PGE₂ measured in the mucosa was presumably generated by adenomatous COX-2 in addition to COX-1 residing in normal and neoplastic tissue.

Steady-state levels of triclin in *Apc^{Min}* mice, which had received triclin (0.2%) with their diet, were 278 ± 126 nmol/g in the small intestinal mucosa (mean ± SD, $n = 10$), but only 0.44 ± 0.12 μmol/L in plasma (mean ± SD, $n = 8$). In the light of the enzyme inhibition data shown in Fig. 3, the intestinal levels of triclin were clearly sufficient to cause substantial inhibition of COX enzymes, while plasma levels were probably just high enough to account for modest inhibition. Furthermore, we cannot discard the possibility that metabolism of triclin to glucuronide or sulfate conjugates affords species that may have contributed to COX inhibition.

Discussion

The results presented here suggest for the first time that triclin possesses adenoma-retarding activity in the *Apc^{Min}* mouse model, and that it may exert its effect, at least in part, by COX inhibition. The latter notion is borne out by the following experimental evidence: (a) triclin potently inhibited the activity of purified COX enzymes at concentrations that were easily achieved in the intestinal mucosa of *Apc^{Min}* mice after triclin consumption, (b) it decreased the generation of PGE₂ in colon cells, which either express COX-2 constitutively (HCA-7) or after stimulation with phorbol ester (HCEC), while down-regulating COX-2 protein expression only weakly (HCEC) or not at all (HCA-7); and (c) triclin consumption decreased PGE₂ levels in murine plasma and small

intestinal mucosa. Intriguingly, the decrease in mucosal PGE₂ observed here is of an order of magnitude comparable with that caused by the nonsteroidal anti-inflammatory drug sulindac (0.016% in drinking water, ~0.5 mg per mouse per day) in *Apc^{Min}* mice (12). The results shown here suggest that triclin is a potent COX inhibitor with little effect on COX-2 protein expression. In contrast, flavonoid analogues of triclin, such as apigenin and quercetin, hardly inhibit COX enzyme activity, although they have been shown to markedly down-regulate COX-2 expression in cells when applied in the 5 to 50 μmol/L concentration range (22, 23). The inhibition of COX enzyme activity by triclin was apparently isoenzyme-nonselective, which renders it more comparable with nonsteroidal anti-inflammatory drugs, such as sulindac and aspirin, rather than to coxibs. The

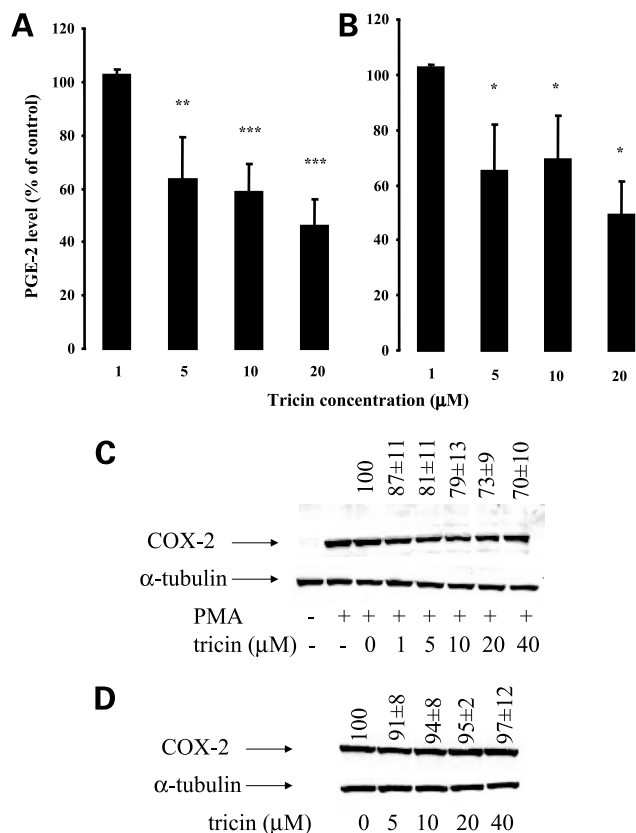


Figure 4. Effect of triclin on PGE₂ generation (A and B) and COX-2 protein expression (C and D) in phorbol ester-stimulated HCEC colon cells (A and C) and HCA-7 colon carcinoma cells (B and D). PGE₂ and COX-2 levels were measured after incubation of cells with triclin for 24 h (A and B) or 6 h (C and D) by immunoassay and Western blotting, respectively, as described under Materials and Methods. Control HCEC and HCA-7 cells (not exposed to triclin) contained 304 ± 66 and 930 ± 398 ng PGE₂/mg cellular protein, respectively (= 100%). Columns, mean of three to five separate experiments (A and B), each conducted in duplicate; bars, SD. Asterisks indicate that values were significantly different from controls (* $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$). Blots in (C) and (D) are from one experiment representative of three; band densities in percentage of control (mean ± SD, $n = 3$) indicated above the blots are the mean ± SD of three separate experiments.

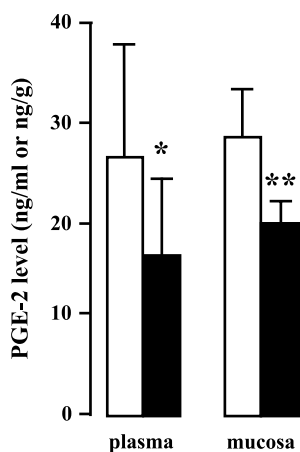


Figure 5. Effect of triclin on PGE₂ levels in plasma and small intestinal mucosa of *Apc*^{Min} mice, which had consumed triclin (0.2% in the diet) for their lifetime. *Open columns*, control mice; *closed columns*, treated mice. PGE₂ levels were measured by immunoassay as described under Materials and Methods. *Columns*, mean ($n = 9-10$); *bars*, SD. Asterisks indicate that values were significantly different from controls (* $P < 0.05$, ** $P < 0.01$).

fact that triclin reduced adenoma numbers particularly in the proximal small intestine could be hypothetically explained by preferential absorption in that segment, but this notion needs experimental verification. The overall incidence of adenomas in the colon in our experiment was very low (fewer than one lesion per mouse), and triclin did not affect colonic adenomas.

In a preliminary experiment, we found that supplementation of the murine diet with rice bran (30%), a dietary source of triclin, also interfered with small intestinal carcinogenesis in *Apc*^{Min} mice.³ Rice bran, which constitutes ~15% of brown rice, has been suggested to contain ~4.5 μ g triclin per 10 g bran (4). Rice bran contains a variety of nonnutrient components other than triclin with potential cancer chemopreventive activity that occur at concentrations exceeding that of triclin (reviewed in ref. 4). The low content of triclin in rice bran renders it difficult to judge to what extent it may contribute to the overall chemopreventive efficacy of bran.

Unwanted side effects of nonsteroidal anti-inflammatory drugs and of COX-2 inhibitors, the latter of which have recently received considerable attention (24), may ultimately militate against their extensive use as cancer chemopreventive agents in humans. Therefore, the search for novel and toxicologically innocuous alternative interventions is highly propitious and diet-derived phytochemicals provide an attractive focus for this search. A preliminary toxicity evaluation suggests that triclin possesses a favorable safety profile vis-à-vis other flavonoids, such as genistein and quercetin (20). This finding, together with the results shown above, strongly

³R.D. Verschoyle, H. Cai, W.P. Steward, and A.J. Gescher, unpublished observation.

supports further preclinical work to explore the suitability of evaluating triclin in humans with intestinal polyps. In such trials, the COX enzyme-inhibitory potency of triclin may be exploitable as a pharmacodynamic marker of its efficacy.

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References

- Kelloff GJ, Crowell JA, Steele VE, et al. Progress in cancer chemoprevention: development of diet-derived chemopreventive agents. *J Nutr* 2000;130:S467–71.
- Witte JS, Longnecker P, Bird CL, et al. Relation of vegetable, fruit and grain consumption to colorectal adenomatous polyps. *Am J Epidemiol* 1996;144:1015–25.
- Mohandas KM, Desai DC. Epidemiology of digestive tract cancers in India. V. Large and small bowel. *Indian J Gastroenterol* 1999;18:118–21.
- Hudson EA, Dinh PA, Kokubun T, et al. Characterization of potentially chemopreventive phenols in extracts of brown rice that inhibit the growth of human breast and colon cancer cells. *Cancer Epidemiol Biomarkers Prev* 2000;9:1163–70.
- Lee KH, Tagahara K, Suzuki H, et al. Antitumor agents. 49. Triclin, kaempferol-3-*O*- β -*D*-glucopyranoside and (+)-nortrachelogenin, antileukemic principles from *Wikstroemia indica*. *J Nat Prod* 1981;44:530–5.
- Luongo C, Moser AR, Gledhill S, Dove WF. Loss of *Apc*(+) in intestinal adenomas from Min mice. *Cancer Res* 1994;54:5947–52.
- Hawk ET, Levin B. Colorectal cancer prevention. *J Clin Oncol* 2005;23:378–91.
- Jacoby RF, Seibert K, Cole CE, et al. The cyclooxygenase-2 inhibitor celecoxib is a potent preventive and therapeutic agent in the Min mouse model of adenomatous polyposis. *Cancer Res* 2000;60:5040–4.
- Steinbach G, Lynch PM, Phillips RKS, et al. The effect of celecoxib, a cyclooxygenase-2 inhibitor, in familial adenomatous polyposis. *N Engl J Med* 2000;342:1946–52.
- Gupta RA, DuBois RN. Colorectal cancer prevention and treatment by inhibition of cyclooxygenase-2. *Nat Rev Cancer* 2001;1:11–21.
- Giardiello FM, Hamilton SR, Krush AJ, et al. Treatment of colonic and rectal adenomas with sulindac in familial adenomatous polyposis. *N Engl J Med* 1993;328:1313–6.
- Boobol SK, Dannenberg AJ, Chadburn A, et al. Cyclooxygenase-2 overexpression and tumor formation are blocked by sulindac in a murine model of familial adenomatous polyposis. *Cancer Res* 1996;56:2556–60.
- Cai H, Steward WP, Gescher AJ. Determination of the putative cancer chemopreventive flavone triclin in plasma and tissues of mice by HPLC with UV-visible detection. *Biomed Chromatogr*. In press 2005.
- Workman P, Twentymann P, Balkwill F, et al. United Kingdom Coordinating Committee on Cancer Research (UKCCCR) guidelines for the welfare of animals in experimental neoplasia. 2nd ed. *Br J Cancer* 1998;77:1–10.
- Perkins S, Verschoyle RD, Hill KA, et al. Chemopreventive efficacy and pharmacokinetics of curcumin in the Min/+ mouse, a model of familial adenomatous polyposis. *Cancer Epidemiol Biomarkers Prev* 2002;11:535–40.
- Mahmoud NN, Carothers AM, Grunberger D, et al. Plant phenolics decrease intestinal tumors in an animal model of familial adenomatous polyposis. *Carcinogenesis* 2000;21:921–7.
- Marsh KA, Stamp GWH, Kirkland SC. Isolation and characterization of multiple cell types from single human colonic carcinoma: tumorigenicity of the cell types in a xenograft system. *J Pathol* 1993;170:441–50.
- Ireson C, Orr S, Jones DJL, et al. Characterization of metabolites of the chemopreventive agent curcumin in human and rat hepatocytes

and in the rat *in vivo* and evaluation of their ability to inhibit phorbol ester-induced prostaglandin E-2 production. *Cancer Res* 2001;61:1058–64.

19. Plummer SM, Holloway KA, Manson MM, et al. Inhibition of cyclooxygenase 2 expression in colon cells by the chemopreventive agent curcumin involves inhibition of NF- κ B activation via the NIK/IKK signalling complex. *Oncogene* 1999;18:6013–20.

20. Verschoyle RD, Greaves P, Cai H, et al. Preliminary safety evaluation of the putative cancer chemopreventive agent triclin, a naturally occurring flavone. *Cancer Chemother Pharmacol*. In press 2005.

21. Sharma RA, Gescher A, Leuratti C, et al. Link between malondialde-

hyde and its DNA adduct levels in human colonic cells of differing cyclooxygenase-2 activities. *Carcinogenesis* 2001;22:1557–60.

22. Liang YC, Huang YT, Tsai SH, et al. Suppression of inducible cyclooxygenase and inducible nitric oxide synthase by apigenin and related flavonoids in mouse macrophages. *Carcinogenesis* 1999;20:1945–52.

23. Raso GM, Meli R, Di Carlo G, et al. Inhibition of inducible nitric oxide synthase and cyclooxygenase-2 expression by flavonoids in macrophage J774A.1. *Life Sci* 2001;68:921–31.

24. Frantz S. Vioxx risk could signify trouble in class. *Nat Rev Drug Discov* 2004;3:899–900.

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