**Preclinical Development**

**TAK-701, a Humanized Monoclonal Antibody to Hepatocyte Growth Factor, Reverses Gefitinib Resistance Induced by Tumor-Derived HGF in Non–Small Cell Lung Cancer with an EGFR Mutation**

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**Abstract**

Most non–small cell lung cancer (NSCLC) tumors with an activating mutation of the epidermal growth factor receptor (EGFR) are initially responsive to EGFR tyrosine kinase inhibitors (TKI) such as gefitinib but ultimately develop resistance to these drugs. Hepatocyte growth factor (HGF) induces EGFR-TKI resistance in NSCLC cells with such a mutation. We investigated strategies to overcome gefitinib resistance induced by HGF. Human NSCLC cells with an activating EGFR mutation (HCC827 cells) were engineered to stably express HGF (HCC827-HGF cells). HCC827-HGF cells secreted large amounts of HGF and exhibited resistance to gefitinib in vitro to an extent similar to that of HCC827 GR cells, in which the gene for the HGF receptor MET is amplified. A MET-TKI reversed gefitinib resistance in HCC827-HGF cells as well as in HCC827 GR cells, suggesting that MET activation induces gefitinib resistance in both cell lines. TAK-701, a humanized monoclonal antibody to HGF, in combination with gefitinib inhibited the phosphorylation of MET, EGFR, extracellular signal-regulated kinase, and AKT in HCC827-HGF cells, resulting in suppression of cell growth and indicating that autocrine HGF-MET signaling contributes to gefitinib resistance in these cells. Combination therapy with TAK-701 and gefitinib also markedly inhibited the growth of HCC827-HGF tumors in vivo. The addition of TAK-701 to gefitinib is a promising strategy to overcome EGFR-TKI resistance induced by HGF in NSCLC with an activating EGFR mutation.

**Introduction**

Somatic mutations in the kinase domain of the epidermal growth factor receptor (EGFR) are associated with a high rate of response to EGFR tyrosine kinase inhibitors (TKI) such as gefitinib (Fig. 1) and erlotinib in advanced non–small cell lung cancer (NSCLC; refs. 1–3). Despite the therapeutic benefit of EGFR-TKIs in NSCLC, however, most patients ultimately develop resistance to these drugs. A secondary T790M mutation of EGFR and amplification of the MET gene are major causes of acquired resistance to EGFR-TKIs (4–7). In addition, hepatocyte growth factor (HGF), a ligand of the MET oncoprotein (8, 9), induces gefitinib resistance in EGFR mutation–positive NSCLC by activating MET and downstream signaling (10).

HGF was originally identified as a mitogenic protein for hepatocytes (11). Both HGF and its MET receptor are expressed, and often overexpressed, in a broad spectrum of human solid tumors including lung, mesothelioma, breast, and brain cancer (12–16). HGF thus acts as an autocrine or paracrine growth factor for these tumor cells (17, 18). TAK-701 is a potent humanized monoclonal antibody to HGF that blocks various HGF-induced biological activities as well as inhibits tumor growth in an autocrine HGF-MET–driven xenograft model.6 To identify strategies or agents capable of overcoming resistance to EGFR-TKIs induced by HGF, we have now established sublines of the EGFR mutation–positive human NSCLC cell line HCC827 that stably express transfected HGF cDNA. With the use of these cells, we investigated the effects of TAK-701 on HGF-MET signaling and gefitinib resistance induced by cell-derived HGF both in vitro and in vivo.

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Materials and Methods

Cell culture and reagents

The EGFR mutant NSCLC cell lines HCC827 (del E746_A750) and the human glioblastoma cell line U87MG were obtained from the American Type Culture Collection. HCC827 GR5 (del E746_A750/MET amplified) was generated and characterized as described previously (6). We screened all sublines of HCC827 for the presence of EGFR mutations by direct DNA sequencing of exons 18 to 21 and MET amplification by fluorescence in situ hybridization analysis with a probe specific for MET and a control probe for the centromere of chromosome 7 as described previously (19, 20) for this study. All cells were passaged for ≤3 months before the renewal from frozen, early-passage stocks obtained from the indicated sources. Cells were regularly screened for mycoplasma with the use of a MycoAlert Mycoplasma Detection Kit (Lonza). HCC827 cells were cultured under a humidified atmosphere of 5% CO2 at 37°C in RPMI 1640 medium (Sigma) supplemented with 10% fetal bovine serum. U87MG cells were cultured in DMEM (Gibco) supplemented with 10% fetal bovine serum. TAK-701 was kindly provided by Takeda Pharmaceutical Co. Ltd., gefitinib was obtained from AstraZeneca, and PHA-665752 was from Tocris Bioscience.

Cell transfection

A full-length cDNA fragment encoding human HGF was obtained from U87MG cells by reverse transcription and PCR with the primers HGF-F (5-GCGGCCGCAGCACCATGTTGGTGACCAAA-3) and HGF-R (5-CGGGATCCCTATGACTGTGGTACCTTATAT-3). The amplification product was verified by sequencing after its cloning into the pCR-Blunt II-TOPO vector (Invitrogen). The HGF cDNA was excised from pCR-Blunt II-TOPO vector (Invitrogen) and transferred to the pQCXIH retroviral vector (Clontech). Retroviruses encoding HGF were then produced and used to infect HCC827 cells as described (21). Cells stably expressing HGF were then isolated by selection with hygromycin at 500 μg/mL (InvivoGen).

Enzyme-linked immunosorbent assay for HGF

Cells (5 × 10^6) were seeded in 6-well plates, cultured overnight in complete medium, and then incubated in serum-free medium for 24 hours, after which the latter medium was collected and assayed for HGF with a Human HGF Quantikine ELISA Kit (R&D Systems). A standard curve for the enzyme-linked immunosorbent assay (ELISA) was generated with the supplied reagents, and HGF concentration was determined as the average value from triplicate samples.

Cell growth inhibition assay

Cells were transferred to 96-well flat-bottomed plates and cultured for 24 hours before exposure for 72 hours to various concentrations of gefitinib, TAK-701, or PHA-665752, as indicated. Tetra Color One (5 mmol/L tetrazolium monosodium salt and 0.2 mmol/L 1-methoxy-5-methyl phenazinium methylsulfate; Seikagaku Kogyo) was then added to each well, and the cells were incubated for 3 hours at 37°C before measurement of absorbance at 490 nm with a Multiskan Spectrum instrument (Thermo Labsystems). Absorbance values were expressed as a percentage of that for untreated cells.

Annexin V binding assay

The binding of Annexin V to cells was measured with the use of an Annexin-V-FLUOS Staining Kit (Roche). Cells were harvested by exposure to trypsin-EDTA, washed with PBS, and centrifuged at 200 × g for 5 minutes. The cell pellets were resuspended in 100 μL of Annexin-V-FLUOS labeling solution, incubated for 10 to 15 minutes at 15°C to 25°C, and then analyzed for fluorescence with a flow cytometer (FACSCalibur) and Cell Quest software (Becton Dickinson).

Immunoblot analysis

Cells were washed twice with ice-cold PBS and then lysed with 1× cell lysis buffer (Cell Signaling Technology) consisting of 20 mmol/L Tris-HCl (pH 7.5), 150 mmol/L NaCl, 1 mmol/L EDTA (disodium salt), 1 mmol/L EGTA, 1% Triton X-100, 2.5 mmol/L sodium pyrophosphate, 1 mmol/L β-glycerophosphate, 1 mmol/L Na3VO4, leupeptin (1 μg/mL), and 1 mmol/L phenylmethylsulfonyl fluoride. The protein concentration of cell lysates was determined with a bicinchoninic acid protein assay kit (Thermo Fisher Scientific), and equal amounts of protein were subjected to SDS-PAGE on 7.5% gels (Bio-Rad). The separated proteins were transferred to a nitrocellulose membrane, which was then incubated with blocking One solution (Nacalai Tesque) for 20 minutes at room temperature before incubation overnight at 4°C with primary antibodies. Antibodies to phosphorylated EGFR (phosphotyrosine-1068), to phosphorylated MET (phosphotyrosine-1349), to phosphorylated or total forms...
of AKT, and to phosphorylated extracellular signal-regulated kinase (ERK) were obtained from Cell Signaling Technology; those to total ERK were from Santa Cruz Biotechnology; those to total EGFR and to total MET were from Zymed/Invitrogen; and those to β-actin were from Sigma. The membrane was then washed with PBS containing 0.05% Tween 20 before incubation for 1 hour at room temperature with horseradish peroxidase-conjugated secondary antibodies (GE Healthcare). Immunocomplexes were finally detected with ECL Western blotting detection reagents (GE Healthcare).

**Growth inhibition assay in vivo**

All animal studies were done in accordance with the Recommendations for Handling of Laboratory Animals for Biomedical Research compiled by the Committee on Safety and Ethical Handling Regulations for Laboratory Animal Experiments, Kinki University. The ethical procedures followed the requirements of the United Kingdom Coordinating Committee on Cancer Research guidelines (22). HCC827 cells were implanted s.c. into the right hind leg of 6-week-old female athymic nude mice (BALB/c nu/nu; CLEA Japan). Tumor volume was determined from caliper measurement of tumor length (L) and width (W) according to the formula $LW^2/2$. Treatment was initiated when tumors in each group of animals achieved an average volume of 300 to 400 mm$^3$. Treatment groups (each containing five mice) consisted of vehicle control, TAK-701 alone, gefitinib alone, and TAK-701 plus gefitinib. The mice were injected with TAK-701 (5 mg/kg) i.p. twice a week for 7 weeks; control animals received PBS as vehicle. Gefitinib (50 mg/kg) was administered by oral gavage daily for 49 days; control animals received a 0.5% (w/v) aqueous solution of hydroxypropylmethylcellulose as vehicle. Both tumor size and body weight were measured twice per week.
Statistical analysis

The data, presented as means ± SD or SE, were analyzed with Student's two-tailed t test, with \( P < 0.05 \) considered statistically significant.

Results

Cell-derived HGF induces gefitinib resistance in EGFR mutation-positive NSCLC cells

To investigate whether cell-derived HGF induces gefitinib resistance in NSCLC cells with an EGFR mutation, we established HCC827 cells (which are EGFR mutation positive) that stably express human HGF (HCC827-HGF1 and -HGF2 cells) or stably harbor the corresponding empty vector (HCC827-Mock cells). The secretion of HGF from these cell lines as well as from the parental (HCC827) cells and from an HCC827 subline with MET amplification (HCC827 GR5) was examined with the use of an ELISA. We found that HCC827-HGF1 and -HGF2 cells released large amounts of HGF into the culture medium, whereas the secretion of HGF from parental (HCC827), HCC827-Mock, or HCC827 GR5 cells was undetectable (Fig. 2A). To assess the effects of gefitinib on cell growth, we exposed these five cell lines to various concentrations of the drug and then measured cell viability. HCC827 GR5 as well as HCC827-HGF1 and -HGF2 cells showed a reduced sensitivity to gefitinib compared with HCC827 and HCC827-Mock cells, with median inhibitory concentrations of \( \sim 10 \) \( \mu \)mol/L apparent for the former cell lines compared with \( \sim 0.1 \) \( \mu \)mol/L for the latter (Fig. 2B). To investigate possible differences in signal transduction among these cell lines, we examined the effects of gefitinib on EGFR, MET, AKT, and ERK phosphorylation by immunoblot analysis (Fig. 2C). In the parental cells, gefitinib markedly inhibited the phosphorylation of EGFR, AKT, and ERK. In contrast, in the resistant cells (HCC827 GR5, and HCC827-HGF1 and -HGF2), gefitinib alone had no effect on AKT and ERK phosphorylation, although it substantially reduced the level of EGFR phosphorylation. These data suggest that sustained AKT and ERK signaling in the presence of gefitinib contributes to gefitinib

![Figure 3](https://example.com/figure3.png)

Figure 3. Effects of the combination of gefitinib and either TAK-701 or PHA-665752 on the growth of gefitinib-resistant NSCLC cells. A to C, HCC827 GR5 cells (A), HCC827-HGF1 cells (B), and HCC827-HGF2 cells (C) were cultured for 72 hours in medium containing 10% serum, various concentrations of gefitinib, and either PHA-665752 (500 nmol/L) or TAK-701 (50 \( \mu \)g/mL), after which cell viability was assessed. Data are means ± SD from three independent experiments. D, HCC827-Mock or HCC827-HGF2 cells were incubated in the absence or presence of gefitinib (1 \( \mu \)mol/L) or TAK-701 (50 \( \mu \)g/mL) for 48 hours in medium containing 10% serum. The proportion of apoptotic cells was then assessed by staining with Annexin V followed by flow cytometry. Data are means ± SD from three independent experiments. *\( P < 0.001 \); n.s., not significant.
resistance in HCC827-HGF1 and -HGF2 cells as well as in HCC827 GR5 cells.

**TAK-701 abrogates gefitinib resistance induced by HGF**

To investigate the roles of MET and HGF in gefitinib resistance in HCC827 GR5 as well as in HCC827-HGF1 and -HGF2 cells, we exposed the cells to the MET-TKI PHA-665752 or to TAK-701, a humanized monoclonal antibody to HGF, in combination with gefitinib. Combined treatment with PHA-665752 and gefitinib was previously shown to result in substantial growth inhibition in HCC827 GR5 (MET amplification–positive) cells (6). We found that the combination of gefitinib and TAK-701 did not affect the growth of HCC827 GR5 cells (Fig. 3A). In HCC827-HGF1 and -HGF2 cells, however, TAK-701 and PHA-665752 each restored the sensitivity of cell growth to inhibition by gefitinib (Fig. 3B and C).

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**Figure 4.** Effects of the combination of gefitinib and either TAK-701 or PHA-665752 on cell signaling in gefitinib-resistant NSCLC cells. HCC827 cells, HCC827 GR5 cells, and HCC827-HGF2 cells were incubated for 6 hours in medium containing 10% serum in the absence or presence of gefitinib (1 μmol/L), PHA-665752 (500 nmol/L), or TAK-701 (50 μg/mL), as indicated. Cell lysates were then prepared and subjected to immunoblot analysis with antibodies to phosphorylated or total forms of EGFR, MET, AKT, or ERK, or with those to β-actin.

**Figure 5.** Effects of the combination of TAK-701 and gefitinib on the growth of gefitinib-resistant NSCLC cells in vivo. Nude mice with tumor xenografts established by s.c. injection of HCC827-Mock (A) or HCC827-HGF2 (B) cells were treated for 7 weeks with vehicle (control), gefitinib (50 mg/kg), TAK-701 (5 mg/kg), or both drugs, as described in Materials and Methods. Tumor volume was determined at the indicated times after the onset of treatment. Data are means ± SE from five mice per group. P < 0.001 for comparison of gefitinib versus gefitinib plus TAK-701 in B.
In addition, staining with Annexin V revealed that gefitinib alone induced a marked increase in the frequency of apoptosis in HCC827-Mock cells but elicited a much smaller effect in HCC827-HGF2 cells (Fig. 3D). However, treatment with both gefitinib and TAK-701 triggered an increase in the number of Annexin V–positive HCC827-HGF2 cells similar in extent to that induced by gefitinib alone in HCC827-Mock cells. These results thus indicate that TAK-701 restores gefitinib-induced apoptosis in HCC827-HGF2 cells.

To examine the effects of gefitinib, PHA-665752, and TAK-701 on cell signaling in the parental, HCC827 GR5, and HCC827-HGF2 cell lines, we again did immunoblot analysis (Fig. 4). Consistent with previous observations (6), PHA-665752 in combination with gefitinib inhibited MET, AKT, and ERK phosphorylation in HCC827 GR5 cells. We further found that TAK-701 alone did not inhibit MET phosphorylation, and thus the combination of TAK-701 and gefitinib did not abrogate AKT and ERK phosphorylation in HCC827 GR5 cells. In HCC827-HGF2 cells, however, TAK-701 as well as PHA-665752 inhibited MET phosphorylation, and the combined treatment with TAK-701 and gefitinib fully suppressed ERK and AKT phosphorylation. These results indicate that HGF-induced gefitinib resistance is mediated by HGF-MET signaling and is abrogated by treatment with TAK-701 in HCC827-HGF2 cells.

Cell-derived HGF induces gefitinib resistance in NSCLC cells and TAK-701 restores the sensitivity of tumor growth to inhibition by gefitinib in vivo

To examine the possible induction of gefitinib resistance by tumor cell–derived HGF and the efficacy of combined treatment with TAK-701 and gefitinib in vivo, we generated xenografts in nude mice by injection of HCC827-Mock or HCC827-HGF2 cells. We found that, whereas gefitinib markedly inhibited the growth of HCC827-Mock xenografts (Fig. 5A), HCC827-HGF2 xenografts were substantially resistant to gefitinib (Fig. 5B). TAK-701 alone had a minimal effect on tumor growth in both HCC827-Mock and HCC827-HGF2 xenograft models. However, the combination of gefitinib and TAK-701 induced marked regression of HCC827-HGF2 xenografts. These results thus suggest that HGF produced by NSCLC tumors harboring an EGFR mutation induces gefitinib resistance, and that TAK-701 abrogates such HGF-induced gefitinib resistance in vivo.

Discussion

In the present study, we established HGF-overexpressing sublines of HCC827 cells and showed that these sublines are resistant to gefitinib both in vitro and in vivo. To investigate whether the resistance of HCC827-HGF cells to gefitinib is attributable to HGF-MET signaling, we examined the effects of the MET-TKI PHA-665752 and of TAK-701, a humanized monoclonal antibody to HGF, on signal transduction and cell growth. In both HCC827-HGF1 and HCC827-HGF2 cells as well as in HCC827 GR5 cells, which are positive for MET amplification, gefitinib alone did not inhibit AKT or ERK phosphorylation, whereas gefitinib in combination with PHA-665752 markedly suppressed the phosphorylation of these signaling molecules. Consistent with these results, PHA-665752 restored the sensitivity of cell growth to inhibition by gefitinib in HCC827-HGF cells as well as in HCC827 GR5 cells. These results indicate that the gefitinib resistance of these cell lines is mediated by MET signaling. TAK-701 has been shown to potently inhibit HGF binding to MET in cancer cells and xenograft models dependent on autocrine HGF-MET signaling. TAK-701 did not inhibit the phosphorylation of MET in HCC827 GR5 cells, suggesting that the activation of MET in these cells is not dependent on HGF. Indeed, we were not able to detect the secretion of HGF from HCC827 GR5 cells. In contrast, TAK-701 suppressed MET phosphorylation, and thus the combination of TAK-701 and gefitinib markedly inhibited both AKT and ERK signaling in HCC827-HGF cells, resulting in their growth inhibition. These results indicate that autocrine HGF-MET signaling contributes to gefitinib resistance in HCC827-HGF cells. Similar ligand-mediated gefitinib resistance has been described previously, with insulin-like growth factor having been found to rescue cells expressing wild-type EGFR from gefitinib-induced inhibition of cell growth (23). These observations suggest that ligand-dependent receptor tyrosine kinase (RTK) activation (by HGF or insulin-like growth factor), as well as ligand-independent RTK activation (by MET amplification), plays a pivotal role in the development of resistance to gefitinib. Further studies should reveal whether other ligand-RTK combinations contribute to gefitinib resistance.

We found that the baseline levels of both MET expression and MET phosphorylation in HCC827-HGF cells were lower than those in HCC827 GR5 cells (Fig. 2C), whereas HCC827-HGF cells were resistant to gefitinib to the same extent as HCC827 GR5 cells in vivo (Fig. 2B). These results suggest that phosphorylated MET activates downstream signaling through different pathways in HCC827 GR5 and HCC827-HGF cells. MET was recently shown to signal through ERBB3 in MET amplification–positive NSCLC cells (6) or through Grb2-associated binder 1 (Gab1) in NSCLC cells with HGF-induced gefitinib resistance (24). Further studies are required to investigate whether the biological properties of NSCLC cells or the abilities of drugs to overcome gefitinib resistance are affected by differences in RTK downstream signaling.

In our HCC827-HGF xenograft model, we showed that HGF secreted from EGFR mutation–positive NSCLC cells drives tumor growth even in the presence of gefitinib, and that combination therapy with TAK-701 and gefitinib was...
TAK-701 Reverses HGF-Induced Gefitinib Resistance

able to greatly inhibit the growth of HCC827-HGF tumors. These results indicate that interruption of HGF-MET signaling with TAK-701 represents a powerful strategy to abrogate gefitinib resistance induced by HGF derived from tumor cells. HGF was previously shown to be expressed predominantly by adenocarcinoma cells in NSCLC specimens, although a low level of HGF staining was also apparent in stromal cells (25). Furthermore, marked expression of HGF has been detected in most lung cancers with intrinsic or acquired resistance to gefitinib (10, 26). These data suggest that our autocrine model systems based on stable overexpression of HGF are clinically relevant and should prove useful for the establishment of strategies to overcome gefitinib resistance. HGF is also produced by stromal cells of various tumor types (13, 27, 28). Indeed, HGF derived from fibroblasts injected into nude mice together with EGFR mutation–positive NSCLC cells induced gefitinib resistance in the NSCLC cell lines in vivo (29). Further studies are required to clarify the major source of HGF that contributes to gefitinib resistance in patients with EGFR mutation–positive lung cancer. Given that TAK-701 inhibits HGF binding to MET, TAK-701 may reverse gefitinib resistance induced by HGF derived not only from tumor cells but also from stromal cells.

In conclusion, we have shown that autocrine activation of MET by HGF confers resistance to gefitinib, and that TAK-701, a humanized monoclonal antibody to HGF, restored sensitivity to gefitinib in tumors with HGF-induced gefitinib resistance. TKIs inhibit several signaling pathways and are therefore associated with a risk of high toxicity, whereas therapeutic antibodies are thought to be less toxic as a result of their high specificity. TAK-701 is currently undergoing phase I trials as a single agent in patients with advanced solid tumors, and our results now suggest that further studies of combination therapy with TAK-701 and gefitinib are warranted in NSCLC patients with HGF-induced EGFR-TKI resistance.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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