Penfluridol Represses Integrin Expression in Breast Cancer through Induction of Reactive Oxygen Species and Downregulation of Sp Transcription Factors

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Abstract

It was recently demonstrated the penfluridol inhibited breast tumor growth and metastasis and this was associated with downregulation of α6- and β4-integrins. In this study, we observed the penfluridol induced reactive oxygen species (ROS) and this was the primary mechanism of action. Penfluridol-mediated growth inhibition, induction of apoptosis, and inhibition of breast cancer cell migration was attenuated after cotreatment with glutathione. Penfluridol also downregulated Sp transcription factors Sp1, Sp3, and Sp4 through epigenetic downregulation of C-Myc and c-Myc-regulated miRNAs (miR27a and miR20a/miR17) and induction of the miR-regulated Sp transcriptional repressors ZBTB10 and ZBTB4. α6- and β4-integrins as well as α5- and β1-integrins are Sp-regulated genes that are also coregulated by the orphan nuclear receptor NRA41 and these integrins can be targeted by agents such as penfluridol that suppress Sp1, Sp3, and Sp4 and also by NRA41 agonists. Mol Cancer Ther; 16(1); 205–16. ©2016 AACR.

Introduction

Repositioning clinically approved drugs for cancer chemotherapy has several advantages including a more rapid drug approval process coupled with potential development of mechanism-based compounds that can be clinically used to target important pro-oncogenic pathways. This approach has been particularly successful with nonsteroidal antiinflammatory drugs (NSAID) and anti-diabetics such as metformin (1, 2). Phenothiazine-derived antipsychotic drugs such as thioridazine and chlorpromazine exhibit anticarcinogenic activity in several different cancer cell lines (3–9). More recent studies have demonstrated that penfluridol, another antipsychotic drug, also inhibits breast and pancreatic cancer cell growth (9, 10). For example, in pancreatic cancer a series of phenothiazene analogues induced apoptosis and inhibited clonogenic growth and colony formation, and more detailed studies with penfluridol indicated that induction of protein phosphatase 2A (PP2A) was a key effect of this compound (9). Penfluridol exhibited antitemastatic activity in triple-negative breast cancer cells and inhibited tumor growth and brain metastasis in three different in vivo models and the key elements were inhibition of α6- and β4-integrin expression (10). However, the mechanisms of the penfluridol-induced responses were not well defined, and this limits potential clinical applications of the compound.

Recent studies in this laboratory showed that β1- and β3-integrin expression in breast cancer cells is regulated by specificity protein 1 (Sp1) transcription factor (TF) in combination with the orphan nuclear receptor 4A1 (NRA41, Nur77, TR3) which acts as a nuclear cofactor (11). Many of the effects observed in breast and other cancer cell lines treated with penfluridol and other phenothiazine derivatives are similar to that observed after knockdown of Sp transcription factors Sp1, Sp3 or Sp4 or after treatment with agents that target Sp TFs (12). For example, knockdown of Sp1, Sp3, or Sp4 individually or combined decreased proliferation and migration/invasion of breast (MDA-MB-231 and SKBR3) and other cancer cell lines (12) and similar results were observed for drugs that repress Sp TF expression (13–18). Moreover, the effects of penfluridol and other phenothiazines on inhibition of several genes including cyclin D1, bcl-2, VEGF receptors, myc, and activation/cleavage caspase-3/PARP (3–10) have also been observed after Sp knockdown (10–18). It was recently reported that the antitemastatic activity of penfluridol in triple-negative breast cancer cells was related to downregulation of α6- and β4-integrin expression (10); however, because both integrin gene promoters are GC-rich, it is possible that Sp1 and other Sp TFs may regulate expression of α6- and β4-integrins as well as α5-integrin (19–21).

Therefore, we hypothesize that the mechanism of action of penfluridol as an antitemastatic agent for triple-negative breast cancer is due to downregulation of Sp TFs. This hypothesis was confirmed in this study which shows that penfluridol induces reactive oxygen species (ROS) in breast cancer cells and ROS-dependent downregulation of Sp1, Sp3, and Sp4 and Sp-dependent genes including α6-, α5-, β1-, and β4-integrins, which are also coregulated by NRA41 and decreased by NRA41 antagonists.
Materials and Methods

Cell lines and antibodies
Breast cancer (SKBR3, MDA-MB-231) cell lines were purchased from ATCC and were kept frozen until initiation of these studies. The cells were received at low passage (<15) and new frozen stocks were used every 6 to 8 weeks. The two cell lines were authenticated by Biosynthesis on February 3, 2015. Cells were maintained 37°C in the presence of 5% CO2 in DMEM/Ham F-12 medium with 10% FBS with antibiotic. DMEM was purchased from GenDepot. Penfluridol, N-acetylcysteine (NAC), catalase, and 36% formaldehyde were purchased from Sigma-Aldrich. Glutathione (GSH) reduced free acid was purchased from Millipore. Hematoxylin was purchased from Vector Laboratories. Apoptotic, Necrotic, and Healthy Cells Quantification Kit was purchased from Biotium. Antibodies were purchased as outlined in Supplementary Table S1.

Cell proliferation, luciferase and ROS assays, and Annexin V staining
Cell proliferation and ROS assays using the cell permeable fluorescent CM-H3DCFDA probe were carried out as described previously (refs. 11–13; also see Supplementary Methods), and changes in cell number were determined by Coulter Z1 cell counter. Annexin V staining used the Vybrant Apoptosis Kit according to the manufacturer’s protocol. The GC-rich promoter luciferase pGL3-pGC3-luc construct and transfection/luciferase assays were carried out as previously described (22).

Boydren chamber assay
SKBR3 and MDA-MB-231 cancer cells (3.0 × 10⁵ per well) were seeded in DMEM/Ham F-12 medium supplemented with 2.5% charcoal-stripped FBS and were allowed to attach for 24 hours. Cells were seeded and subsequently treated with varying concentrations of penfluridol for 24 hours (±GSH, 3 hours prior to treatment) and for subsequent 24 hours of cotreatment. Cells were trypsinized, counted, placed in 12-well 8.0 μm pore ThinCerts from Greiner Bio-one, allowed to migrate for 24 hours, fixed with formaldehyde, and then stained with hematoxylin. Equal numbers of cells were used for each assay and cells that migrated through the pores were then counted as described (11–13).

RT-PCR
miRNA was isolated using the mirVana miRNA Isolation Kit (Ambion) according to the manufacturer’s protocol. Quantification of miRNA (RNU6B and miR-17, miR-20a, and miR-27a) was done using the TaqMan miRNA Assay Kit (Life Technologies) according to the manufacturer’s protocol with real-time PCR. U6 small nuclear RNA was used as a control to determine relative miRNA expression.

Western blot analysis
SKBR3 and MDA-MB-231 cancer cells (3.0 × 10⁵ per well) were seeded in DMEM/Ham’s F-12 medium supplemented with 2.5% charcoal-stripped FBS and were allowed to attach for 24 hours. Cells were treated with varying doses of penfluridol for 24 hours. Cells were analyzed by Western blot analysis as described previously (11–13) and the catalase inhibition studies were determined as described previously (17).

siRNA interference assay
siRNAs used are outlined in Supplementary Table S1. siRNA experiments were conducted as described previously (11–13).

Chromatin immunoprecipitation
The chromatin immunoprecipitation (ChIP) assay was performed using the ChIP-IT Express Magnetic Chromatin Immunoprecipitation Kit (Active Motif) according to the manufacturer’s protocol. SKBR3 and MDA-MB-231 cells were treated with DMSO, DIM-C-pPhOH (15 or 20 μmol/L), DIM-C-pPhCO₂Me (15 or 20 μmol/L) for 24 hours, or penfluridol (5 μmol/L) for 3 or 6 hours. Cells were then fixed with 1% formaldehyde, and the cross-linking reaction was stopped by addition of 0.125 mol/L glycine. After washing twice with PBS, cells were scraped and pelleted. Collected cells were hypotonically lysed, and nuclei were collected. Nuclei were then sonicated to the desired chromatin length (~200 to 1,500 bp). The sonicated chromatin was immunoprecipitated with normal IgG, p300, Sp1, Sp3, Sp4, NR4A1, or RNA polymerase II antibodies and protein A-conjugated magnetic beads at 4°C for overnight. After the magnetic beads were extensively washed, protein-DNA cross-links were reversed and eluted. DNA was prepared by proteinase K digestion followed by PCR amplification. Primers used for detecting PCR products are listed in Supplementary Table S1. PCR products were resolved on a 2% agarose gel in the presence of RGB-4103 GelRed Nucleic Acid Stain.

Xenograft studies
Female athymic nude mice 4 to 6 weeks old were purchased, and MDA-MB-231 cancer cells (2.0 × 10⁵ cells) were suspended in Matrigel (1:1 ratio) and injected into the mammary fat pad of athymic nude mice. When tumors became palpable (150 to 200 mm³), mice were randomly assigned to control (corn oil vehicle) and penfluridol (10 mg/kg/day), and then treated every day for 19 days. Tumor volumes, tumor weights, and tumor lysates were determined and analyzed as previously described (13, 4).

Statistical analysis
Statistical significance of differences between the treatment groups was determined as previously described (11–13).

Results
Drugs that downregulate Sp TFs act through ROS-dependent and-independent pathways (18), and results in Fig. 1A show that different concentrations of penfluridol induced ROS in MDA-MB-231 and SKBR3 cells after treatment for 3 and 6 hours and similar results were observed after 24 hours (Supplementary Fig. S1A). Moreover, after cotreatment with 5 mmol/L GSH, the penfluridol-mediated induction of ROS as detected using the fluorescent probe CM-H₂DCFDA was significantly attenuated. The role of penfluridol-induced ROS on cell proliferation (Fig. 1B), Annexin V staining (apoptosis; Fig. 1C), and migration in a Boyden chamber assay (Fig. 1D) was also determined in MDA-MBA-231 and SKBR3 cells treated with penfluridol alone and in combination with GSH. Penfluridol inhibited cell growth and induced Annexin V staining; however, we now show for the first time that all of these responses were ROS-dependent and significantly attenuated after cotreatment with GSH. SKBR3 cells appeared to be more resistant to penfluridol than MDA-MB-231 cells and this may be due to several factors including...
constitutive ROS levels, GSH and reductant levels, and differential expression of drug-metabolizing and transport enzymes.

Results illustrated in Fig. 2A and B show that penfluridol decreased expression of Sp1, Sp3 (both high and low molecular weight forms) and Sp4 in MDA-MB-231 and SKBR3 cells and cotreatment with GSH attenuated these effects, and similar results were observed for NAC (Supplementary Fig. S1B). For some responses (e.g., Sp1), addition of GSH or NAC enhanced...
Penfluridol induces ROS-dependent modulation of Sp1, Sp3, Sp4, and other responses in breast cancer cells. MDA-MB-231 and SKBR3 cells were treated with different concentrations of penfluridol alone or in combination with GSH for 24 hours, and whole cell lysates were analyzed for Sp1, Sp3, and Sp4 (A and B, respectively) and other responses (C and D, respectively) as outlined in the Materials and Methods. E, Penfluridol-mediated downregulation of Sp proteins was also determined in the presence or absence of catalase by Western blots of whole cell lysates. F, MDA-MB-231 and SKBR3 cells were transfected with pGL3-GC3-luc and luciferase was determined as described previously (22).
endogenous expression of these proteins. Penfluridol decreased focal adhesion kinase (FAK) phosphorylation and activated (cleaved) caspase 3 in breast cancer cells (Fig. 2C and D) as previously reported (10) and the responses were attenuated by cotreatment with GSH. Penfluridol also induced ROS-dependent caspase 3/PARP cleavage (apoptosis) in both cell lines (Fig. 2C and D). In addition, penfluridol-mediated down-regulation of Sp proteins was also inhibited after addition of catalase (Fig. 2E) and penfluridol also decreased luciferase activity in these cells transfected with a construct containing 3 consensus (Sp) binding sites (Fig. 2F), confirming that induction of apoptosis and the growth- and migration-inhibitory effects of penfluridol in breast cancer cells were ROS dependent.

Penfluridol activates the ROS → Myc → miR27a/miR20/17a → ZBTB pathway

Figure 3 illustrates the cascade of events leading to ROS-dependent downregulation of Sp TFs and this is initiated by cMyc downregulation due to rapid shifts of chromatin-modifying complexes from non-GC-rich to GC-rich (e.g., cMyc) gene promoters (13, 14, 23). Results in Fig. 3B show that penfluridol also decreased cMyc expression in MDA-MB-231 and SKBR3 cells and this response was totally reversed after cotreatment with GSH, and Fig. 3C shows that cMyc and Sp1 were rapidly decreased in MDA-MB-231 and SKBR3 cells after treatment with penfluridol. ChIP analysis showed that pol II association with the GC-rich cMyc promoter was decreased after treatment of MDA-MB-231 and SKBR3 cells with penfluridol for 3 and 6 hours, and this was
accompanied by an increase in the inhibitory H3K27me3 and a decrease in the activating H3K4me4 and H4K16Ac histone marks (Fig. 3D). The ROS-dependent decrease in cMyc was accompanied by a decrease in the cMyc-regulated miR27a and miR20a/miR17 after treatment penfluridol and this decrease was also inhibited after cotreatment with GSH (Fig. 3E). Penfluridol-mediated repression of miR27a and miR20a/miR17 (part of the miR17-92 cluster) resulted in induction of the miR-regulated Sp repressors ZBTB10 and ZBTB4, respectively, and this induction response was also attenuated in cells cotreated with GSH (Fig. 3F). These results demonstrate that the mechanism of penfluridol-induced ROS is initiated by ROS-mediated epigenetic repression of cMyc (Fig. 3A) and this has previously been observed for phenylethylisothiocyanate (PEITC) and HDAC inhibitors in pancreatic cancer and rhabdomyosarcoma (RMS) cells, respectively (13, 14).

Penfluridol induces ROS-dependent repression of integrins
Penfluridol was reported to decrease α6- and β4-integrin in breast cancer cells (10). Our results confirm this same response in MDA-MB-231 and SKBR3 cells, and cotreatment with GSH which attenuated this effect (Fig. 4A) indicating that this response was also ROS-dependent. We also observed that α5- and β1-integrin expression were decreased by penfluridol and rescued after cotreatment with GSH (Fig. 4A and B) and this was consistent with our recent studies showing that β1-integrin is an Sp-regulated gene and α5-integrin appears to be coregulated with β1-integrin (11). Confirmation that α6, α5-, β4-, and β1-integrin are Sp-regulated genes was determined by RNA interference where knockdown of one or more of Sp1 (siSp1), Sp3 (siSp3), Sp4 (siSp4), or all three Sp5 (siSp1,3,4) decreased expression of the integrins in MDA-MB-231 (Fig. 4C) and SKBR3 (Fig. 4D) cells.
Lysates for this Western blot analysis were obtained from a previous study which showed efficient knockdown of Sp1, Sp3, and Sp4 (12). Results of Sp knockdown show that α5- and β4-integrins were coregulated by two or all three of the Sp TFs; however, α6-integrin and β1-integrin were regulated only by Sp4 (MDA-MB-231) and Sp1 (SKBR3), respectively. Knockdown of cMyc by RNA decreased Sp1, Sp3, and Sp4 expression in MDA-MB-231 and SKBR3 cells (Fig. 4E), and the same treatment also decreased expression of α6-, α5-, β1-, and β4-integrin in the same cell lines (Fig. 4F). Similar results were observed with a second oligonucleotide targeting cMyc (Supplementary Fig. S1C). Thus, penfluridol activation of ROS results in cMyc downregulation, leading to decreased expression of Sp1, Sp3, Sp4, and Sp-regulated genes α6-, α5-, β1-, and β4-integrin. The critical role of cMyc was confirmed in studies showing that penfluridol-induced downregulation of Sp proteins, Sp-regulated genes, and caspase 3/PARP cleavage was rescued by Myc overexpression (Supplementary Fig. S2). Moreover, similar results were observed after knockdown of ZBTB10 (Supplementary Fig. S3).

As the α6-, α5-, β1-, and β4-integrins are Sp-regulated genes, we also investigated the effects of penfluridol on association of pol II and Sp1, Sp3, Sp4 with the GC-rich regions of the integrin gene promoters (Fig. 5A) using a ChIP assay. In addition, as we previously observed that NR4A1 and p300 act as cofactors for Sp-dependent activation of β1-integrin (11), the effects of penfluridol on their association with the integrin promoters was also determined. Figure 5B–E shows that pol II, Sp1, Sp3, Sp4, p300, and NR4A1 were all associated with the α6-, β4-, α5-, and β1-integrin gene promoters as determined in a ChIP assay. With the exception of p300, treatment of MDA-MB-231 and SKBR3 cells

Figure 5.
Penfluridol decreases interactions of Sp proteins and other nuclear factors with integrin gene promoters. A, Outline of GC-rich regions for α6-integrin (ITGA6), β4-integrin (ITGB4), α5-integrin (ITGA5), and β1-integrin (ITGB1) gene promoters and primers targeting these regions. Breast cancer cells were treated with DMSO and 5 μmol/L penfluridol, and interactions of Sp1, Sp3, Sp4, and other nuclear cofactors with the ITGA6 (B), ITGB4 (C), ITGA5 (D), and ITGB1 (E) gene promoters were determined in a ChIP assay as outlined in the Materials and Methods.
with penfluridol decreased association of Sp1, Sp3, Sp4, and NRA1 with the four integrin promoters, whereas the loss of p300 was promoter- and cell context-dependent.

**NRA1/Sp regulation of integrins**

NRA1/Sp regulate expression of β1- and α5-integrin in breast cancer cells, and knockdown of Sp or NRA1 or treatment with an NRA1 antagonist inhibits their expression (11). Therefore, we investigated the activity of penfluridol as an NRA1 ligand in Panc1 cells transfected with a GAL4-NRA1 chimera and a reporter gene containing a GAL4 response element; this assay is used for screening NRA1 antagonists (24, 25), and penfluridol did not inhibit or activate this system (Supplementary Fig. S4). Knockdown of NRA1 (siNR4A1) or treatment with the NRA1 antagonists 1,1-bis(3’-indolyl)-1-(p-hydroxyphenyl)methane (DIM-C-pPhOH) or 1,1-bis(3’-indolyl)-1-(p-carbomethoxyphenyl)methane (DIM-C-pPhCO₂Me) decreased expression of 6- and β4-integrin in MDA-MB-231 and SKBR3 cells (Fig. 6A) and similar results were previously observed for β1- and α5-integrin (11). Treatment of MDA-MB-231 and SKBR3 cells with DIM-C-pPhOH (C-DIM8) or DIM-C-pPhCO₂Me (C-DIM14) showed that these compounds induced loss of pol II from the α6- (Fig. 6C), β4- (Fig. 6D), and α5- (Fig. 6E) integrin promoters in a ChIP assay and this was previously observed for the β1-integrin promoter (11). Unlike penfluridol which decreased expression of Sp1, Sp3, and Sp4, resulting in loss of these TFs from the integrin promoters (Fig. 5), NRA1 antagonists or NRA1 knockdown did not affect Sp expression (11) and retention or loss of Sp1, Sp3, or Sp4 from the integrin promoters in ChIP assays is highly variable and gene promoter- and cell context-dependent (11). This variability was observed for p300 (Fig. 6C-E) and also in a previous study (11). Nevertheless, the results show that both penfluridol and NRA1 antagonists downregulate expression of α5, α6-, β1-, and β4-integrin by selectively targeting the Sp and NRA1 transcription factors, respectively, which are required for expression of the integrin genes (Fig. 6F).

Treatment of athymic nude mice bearing MDA-MB-231 cells in an orthotopic model with penfluridol (10 mg/kg/day) decreased tumor volume over the treatment period (Fig. 7A) and inhibited tumor growth (Fig. 7B), and toxicity (organ damage or weight loss) was not observed. Western blot analysis of lysates from control (solvent) and penfluridol-treated mice showed that penfluridol decreased expression of Sp1, Sp3, and Sp4, prototypical Sp-regulated genes (survivin and EGFR) including α6-, α5-, β1-, and β4-integrin and induced apoptosis (Fig. 7C). These effects were also quantitated (Fig. 7D) and penfluridol significantly decreased expression of Sp proteins and Sp-regulated genes or increased caspase 3/PPAR cleavage compared with control protein set at 100% and normalized to β-actin. Thus, the in vivo results complement the in vitro studies demonstrating that the anticancer activities of penfluridol are ROS-dependent and this includes NRA1/Sp-mediated downregulation of integrins.

**Discussion**

Integrins are cell surface receptors that function as α/β heterodimers, which are formed from the 18 α and 8 β subunits, and the functions of the known 24 β/α-integrin heterodimers are highly tissue/cell type specific. In cancer cells, integrins are important for migration and invasion, and overexpression of some integrins in cancer patients is a negative prognostic factor (26, 27). Integrins play a key role in breast cancer stem cells and mammary tumors, and there is extensive evidence for the role of β1-, β4-, α5-, and α6-integrins and their α/β heterodimers in breast cancer cell migration, invasion, and metastasis (28–34). Although integrin inhibitors have been developed, their applications for cancer chemotherapy have been limited (26) and this is due, in part, to β1- to β3-integrin switching in which drugs targeting β1-integrin induce β3-integrin (35, 36). We recently showed that the orphan nuclear receptor NRA1 regulates expression of β1-integrin and treatment of breast cancer cells with bis-indole-derived (C-DIMs) NRA1 antagonists such as DIM-C-pPhOH and DIM-C-pPhCO₂Me decrease both β1- and β3-integrin expression and eliminate the switching pathway (11). Both integrins are regulated by an NRA1–Sp complex, which binds to GC-rich regions of their gene promoters (Fig. 6F), suggesting that β1- and β3-integrin can be targeted by NRA1 antagonists or drugs that target Sp transcription factors.

It was previously reported that penfluridol decreased expression of α6- and β4-integrin in breast cancer cells (10). Because these cell lines highly express Sp1, Sp3, and Sp4 (37) and because there is evidence for Sp-regulation of these integrins (19, 20), we hypothesized that the mechanism of action of penfluridol may be due to repression of Sp proteins. We also investigated α5-integrin expression which also has a GC-rich gene promoter (21). Using SKBR3 and MDA-MB-231 cells as a model, it was clear that penfluridol-induced ROS and the resulting inhibition of cell growth and migration and induction of apoptosis were all inhibited after cotreatment with GSH (Fig. 1). Subsequent experiments showed that induction of ROS was the key factor in mediating downregulating Sp1, Sp3, and Sp4 through ROS-dependent epigenetic repression of chtyc, decreased expression of chtyc-regulated miR27a and miR20a/miR17, and induction of the Sp repressors ZBTB10 and ZBTB4. This ROS-dependent pathway has previously been observed for other ROS inducers such as PERTC and HDAC inhibitors (13, 14). The ROS–miRZRTB–Sp pathway (Fig. 3A) has also been observed in cancer cells treated with a nitro-aspirin derivative, celastrol, betulinic acid and curcumin; other ROS inducers including arsenic trioxide, ascorbate, hydrogen peroxide, and l-butyhydroperoxide also downregulate Sp1, Sp3, and Sp4 in cancer cell lines (15–17, 38–41). We also observed that penfluridol targeted Sp TFs and pro-oncogenic Sp-regulated genes in vivo and this was consistent with potent inhibition of tumor growth in an orthotopic model (Fig. 7) and complemented results of previous studies on the anticancer activities of penfluridol (9, 10).

This article also demonstrates that penfluridol-mediated downregulation of α5-, α6-, and β4-integrins is also ROS-dependent and Sp TFs regulate expression of these genes in breast cancer cell lines. Moreover, like β1- and β3-integrin (11), the α5-, α6-, and β4-integrins are coregulated by NRA1 and expression of this family of integrins is due to interactions of the NRA1–Sp complex with their corresponding GC-rich gene promoter elements (Fig. 6C). Penfluridol does not exhibit NRA1 antagonist activity but targets the integrins through repression of Sp TFs, whereas the C-DIM/NRA1 antagonists inactivate the coactivator-like activity of NRA1 (Fig. 6F). Nuclear receptor-mediated activation of genes through interaction with DNA bound Sp TFs has been observed for several other receptors (42) and like the integrins in this study can be targeted through inactivation of the receptor or downregulation of Sp TFs. The identification of the mechanism of action of penfluridol coupled with the reported
Figure 6.
Integrin genes targeted by penfluridol are coregulated by Sp transcription factors and NR4A1. Breast cancer cells were transfected with siNR4A1 (A) or treated with the NR4A1 antagonists DIM-C-pPhOH (C-DIM8) and DIM-C-pPhCO2Me (C-DIM14; B), and whole-cell lysates were analyzed by Western blots. Breast cancer cells were treated with DMSO and the two NR4A1 antagonists, and interactions of Sp1, Sp3, Sp4, and other nuclear cofactors with the ITGA6 (C), ITGB4 (D), and ITGA5 (E) gene promoters were determined in a ChIP assay. The same interactions with the ITGB1 promoter were previously reported (11). F, Model of NR4A1/Sp interactions with GC-rich integrin gene promoters and the differential targeting by penfluridol and NR4A1 antagonists.
Figure 7.
Penfluridol inhibits tumor growth in vivo. Penfluridol (10 mg/kg/day) inhibit tumor volume (A) and weight (B) in athymic nude mice bearing MDA-MB-231 cells in an orthotopic model. Western blot analysis of tumor lysates (C) and quantitation of various proteins (D and E) where control proteins normalized to β-actin were set at 100% and penfluridol-mediated changes (normalized to β-actin) compared with controls were determined as outlined in the Materials and Methods. Both high and low molecular weight forms of Sp3 were detected.
effectiveness of this compound as an inhibitor of breast cancer metastasis will facilitate the design of future clinical applications of this ROS inducer for breast cancer therapy. This would also include applications in treating tumors such as RMS which exhibit high endogenous ROS levels (9, 43–44).

Disclosure of Potential Conflicts of Interest
No potential conflicts of interest were disclosed.

Authors’ Contributions
Conception and design: E. Hedrick, S. Safe
Development of methodology: E. Hedrick, S. Safe
Acquisition of data (provided animals, acquired and managed patients, provided facilities, etc.): E. Hedrick, X. Li
Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): E. Hedrick, X. Li
Writing, review, and/or revision of the manuscript: E. Hedrick, X. Li, S. Safe
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): S. Safe
Study supervision: S. Safe

Grant Support
The financial assistance of the NIH (P30-ES023512, to S. Safe), Texas AgriLife Research and Sid Kyle endowment, is gratefully acknowledged. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received July 11, 2016; revised October 14, 2016; accepted October 26, 2016.
Published OnlineFirst November 3, 2016.

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Molecular Cancer Therapeutics

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