Effect of Niclosamide on Basal-like Breast Cancers

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Abstract

Basal-like breast cancers (BLBC) are poorly differentiated and display aggressive clinical behavior. These tumors become resistant to cytotoxic agents, and tumor relapse has been attributed to the presence of cancer stem cells (CSC). One of the pathways involved in CSC regulation is the Wnt/β-catenin signaling pathway. LRP6, a Wnt ligand receptor, is one of the critical elements of this pathway and could potentially be an excellent therapeutic target. Niclosamide has been shown to inhibit the Wnt/β-catenin signaling pathway by causing degradation of LRP6. TRA-8, a monoclonal antibody specific to TRAIL death receptor 5, is cytotoxic to BLBC cell lines and their CSC-enriched populations. The goal of this study was to examine whether niclosamide is cytotoxic to BLBCs, specifically the CSC population, and if in combination with TRA-8 could produce increased cytotoxicity. Aldehyde dehydrogenase (ALDH) is a known marker of CSCs. By testing BLBC cells for ALDH expression by flow cytometry, we were able to isolate a nonadherent population of cells that have high ALDH expression. Niclosamide showed cytotoxicity against these nonadherent ALDH-expressing cells in addition to adherent cells from four BLBC cell lines: 2LMP, SUM159, HCC1187, and HCC1143. Niclosamide treatment produced reduced levels of LRP6 and β-catenin, which is a downstream Wnt/β-catenin signaling protein. The combination of TRA-8 and niclosamide produced additive cytotoxicity and a reduction in Wnt/β-catenin activity. Niclosamide in combination with TRA-8 suppressed growth of 2LMP orthotopic tumor xenografts. These results suggest that niclosamide or congeners of this agent may be useful for the treatment of BLBC. Mol Cancer Ther; 13(4); 800–11. ©2014 AACR.

Introduction

Breast cancer is the second leading cause of cancer-related deaths in North American women. Basal-like breast cancer (BLBC), one of four subtypes of breast cancer, accounts for 13% of all breast carcinomas. Although not as common as the other subtypes, BLBC is the most deadly and aggressive type (1, 2). This aggressiveness is due to the fact that it is also the most chemoresistant of the breast cancer subtypes (3). Biologically, it is characterized by a unique mRNA profile with CK5/6 expression and inactivation of BRCA1. In addition, 70% to 90% of BLBC tumors are characterized as triple-negative breast cancer (TNBC) because they lack progesterone and estrogen receptor expression and HER-2 amplification (1, 4, 5). The poor prognosis is linked to its enrichment for tumor-initiating cells known as cancer stem cells (CSCs; refs. 6–8). CSCs engage in self-renewal, induce tumors at low-cell density, and produce tumors with differentiated and heterogeneous cell profiles. Moreover, they exhibit gene expression profiles that diverge from their more differentiated cancer cell counterparts. Breast CSCs form tumorspheres in vitro, they are more tumorigenic in mice, and more resistant to standard chemotherapy and radiation than differentiated cells (9, 10). In BLBC, CSCs are identified by their extracellular expression of CD44+/CD24− and elevated enzymatic activity of aldehyde dehydrogenase (ALDH; refs. 11, 12). These CSCs are also identified based on the aberrant regulation of their self-renewing pathways, including Wnt, Hedgehog, and Notch signaling (11, 13).

One promising approach to prevent BLBC recurrence and metastasis is to target pathways that regulate CSCs such as the Wnt/β-catenin pathway (3, 14). The cell surface receptor LRP6, essential for Wnt/β-catenin signaling, is a potential target as its expression is upregulated in 20% to 36% of human breast cancers and most significantly in the BLBC subtype. Suppression of LRP6 has been proved to be sufficient in inhibiting the Wnt/β-catenin signaling pathway in breast cancer; therefore,
it is an excellent potential target for the treatment of BLBC (14–17).

Wnt proteins activate the Wnt/β-catenin pathway by binding to its surface receptors LRP5/6. This binding induces the receptors to interact with the transmembrane receptor, frizzled, which leads to the subsequent phosphorylation of LRP5/6 (18). This leads to a build-up of β-catenin, an intracellular signal transducer, in the cytoplasm. β-Catenin can then translocate to the nucleus, in which it interacts with T-cell factor (TCF). This interaction triggers the transcription of the Wnt pathway target genes, which include survivin, Axin2, and cyclin D1. The expression of these genes leads the cell to undergo proliferation, self-renewal, and survival. In the absence of a Wnt ligand, β-catenin is tagged for degradation by the “destruction complex” comprised of adenomatous polyposis coli, Axin, and GSK3β, thereby rendering the β-catenin target genes transcriptionally inactive.

The Wnt/β-catenin pathway can be inhibited at the extracellular level by secreted inhibitors such as Dkkopf proteins or secreted frizzled-related proteins (19, 20). Chemically, this inhibition can be achieved by salinomycin or niclosamide, which both are able to inhibit the binding of a Wnt ligand to LRP5/6 receptors (21, 22). Inhibitors of Wnt/β-catenin signaling, such as niclosamide, are reported to stimulate frizzled internalization and promote LRP6 degradation, thus preventing proliferation and causing apoptosis (22–24). Niclosamide (trade name Niclocide) is a teniacide in the antihelminth family. It is a safe, inexpensive drug that has been used in humans for nearly 50 years (25). Niclosamide has also been shown to be cytotoxic against prostate cancer, colorectal cancer, myelogenous leukemia, and ovarian cancer; in ovarian cancer, it has been specifically shown to suppress CSCs (24, 26–28).

Wnt/β-catenin signaling is also inhibited by TRAIL, specifically by promoting caspase-3 and –8-mediated cleavage of β-catenin (29, 30). TRAIL also preferentially induces apoptosis in BLBC (31). TRA-8 is an agonistic monoclonal antibody (mAb) to TRAIL death receptor 5 (DR5; refs. 32, 33). We have previously shown that TRA-8 can kill both parental and CSCs from BLBC (34, 35). Furthermore, niclosamide has been shown to reduce the expression of the transcription factor STAT3 (36). This is important for our studies because STAT3 plays a key role in many cellular processes such as cell growth and apoptosis, and in breast cancer, STAT3 has been shown to be constitutively activated (37). More recently, it has been described that in breast CSCs, the STAT3 pathway plays a critical role in the conversion of non-CSCs into CSCs through regulation of OCT4 gene expression (38). On the basis of this information, we hypothesized that the suppression of canonical Wnt/β-catenin and STAT3 activity by niclosamide would have cytotoxic potential alone and would sensitize BLBC stem cells to treatment with TRA-8. In this study, we explored whether BLBC cell lines, nonadherent ALDH-enriched (NAAE) cells, and cells isolated from BLBC patient pleural effusion samples were sensitive to niclosamide alone or in combination with TRA-8.

Materials and Methods

Drugs and antibodies

Niclosamide was purchased from Sigma-Aldrich. Niclosamide for in vitro use was dissolved in dimethyl sulfoxide (DMSO) at a 4.8-mmol/L concentration and stored at 4°C until further use. For animal studies, niclosamide was dissolved in DMSO until a homogeneous suspension was observed at which time Cremophor was added to make a final solution of 25% DMSO and 75% Cremophor. The liquid was slowly inverted to obtain a clear orange solution, which was stored at 4°C.

Niclosamide was purchased from Sigma-Aldrich. Monoclonal anti–phosphorylated-LRP6, Axin2, cyclin D1, p(Tyr705)-STAT3, and STAT3 were purchased from Cell Signaling Technology, Inc.. Monoclonal anti–β-catenin antibody was obtained from Santa Cruz Biotechnology. Survivin antibody for Western blots was purchased from Southern Biotechnology Associates. ALDEFLUOR Kit including diethylamino benzaldehyde was obtained from StemCell Technologies. The 2LMP subclone of the human breast cancer cell line MDA-MB-231 was obtained from Dr. Marc Lippman (University of Miami, Miami, FL) and maintained in improved minimum essential medium (MEM) supplemented with 10% FBS (HyClone). BLBC cell lines, HCC1187 and HCC1143, were obtained from American Type Culture Collection (ATCC) and cultured according to supplier’s directions. SUM159 was obtained from Asterand and grown according to supplier’s recommendation. MCF10A immortalized, nontransformed epithelial cells were obtained from ATCC. Cell lines were obtained 3 years ago, frozen in aliquots, which were passed no more than 15 times, and then a new aliquot was used. They were not authenticated. All cell lines were maintained in antibiotic-free medium at 37°C in a 5% CO2 atmosphere and routinely screened for mycoplasma contamination.

With Institutional Review Board approval, pleural effusion fluid was collected from patients with advanced-stage TNBC. Cells were washed in PBS (HyClone) and isolated via centrifugation at 400 rpm × 5 minutes. Cells were then plated in ultralow attachment plates (Corning) and incubated in mammary epithelial cell growth medium (MEGM; Lonza).

Generation and characterization of tumorspheres and NAAE cells

BLBC cells were seeded in 96-well ultralow attachment plates (Corning) at 40,000 cells/mL in mammary epithelial
basal medium (MEBM) supplemented with bullet kit (bovine pituitary extract, insulin, human recombinant epidermal growth factor, and hydrocortisone); complete media is called MEGM (Lonza). These nonadherent cells then formed tumorspheres. Cells were tested for ALDH activity at 6, 12, 24, 48, and 72 hours after plating by flow cytometry as previously described (34). In addition, cells were sorted for the ALDH+ population and retention of ALDH was analyzed at the same time points. Both sorted and nonsorted cells were found to be maximally enriched for ALDH at 12 hours, but then decreased at later time points. The NAAE cells obtained at the 12-hour time point (20,000 cells) were injected into the mammary fat pad (MFP) of nonobese diabetic/severe combined immunodeficient (NOD/SCID) mice. The nonadherent cells at 72 hours after plating (20,000 cells) were also injected into the MFP of mice. To evaluate the difference between the tumorigenicity of the NAAE cells (12 hours after plating) and the nonadherent cells (72 hours after plating), the tumors were measured twice weekly until mice were euthanized in accordance to Institutional Animal Care and Use Committee (IACUC) regulations.

**Generation of LRP6 knockdown cell line**

LRP6 protein expression was knocked down in the adherent 2LMP cell line using Mission Lentiviral transduction particles SHCI-LNV, MOI of the TRCN000003405 (Lot 01091309MN, 1.5 × 10^5 TU/mL). We used Mission transduction particles for nonmammalian short hairpin RNA (shRNA) as a control, SHCI002V, MOI 3 (Lot 04301208MN, 2.6 × 10^5 TU/mL; Sigma-Aldrich). Cells were selected using ready-made solution puromycin dihydrochloride (Sigma-Aldrich). LRP6 knockdown was confirmed using Western blot analysis of LRP6.

**Western blotting**

To evaluate the difference in Wnt/β-catenin pathway signaling in NAAE cells and adherent cells, adherent and NAAE 2LMP cells were seeded (1 × 10^5 cells/well) in 6-well plates. Cells were lysed in 0.5 mL of lysis buffer (PBS containing 1% Triton X-100 and 1 mmol/L phenylmethylsulfonylfluoride) at 4°C for 15 minutes. Equal quantities of protein were subjected to SDS-PAGE under reducing conditions. Following transfer to immobilon-P membrane, successive incubations with anti-LRP6, anti–p-LRP6, anti–free β-catenin, anti–total β-catenin or actin, and horseradish peroxidase–conjugated secondary antibody were carried out for 60 to 120 minutes at room temperature. The immunoreactive proteins were then detected using the ECL system (PerkinElmer). Films showing immunoreactive bands were scanned by Kodak Digital Science DC120 Zoom Digital Camera (Kodak). Cytosolic free β-catenin analysis was done with GST–E-cadherin binding assay (22). Uncomplexed cytosolic free β-catenin in 100 mg of total cell lysate was subjected to SDS-PAGE and detected using the mAb to β-catenin.

For treatment studies, 2LMP adherent and NAAE cells, and patient samples were seeded (1 × 10^6 cells/well) in 6-well plates and treated for 24 hours with niclosamide (0, 0.5, 0.25, 0.125 μmol/L). Additional proteins detected were Axin 2, survivin, cyclin D1, p(Tyr705)-STAT3, and STAT3. 2LMP and HCC1187 adherent and NAAE cells were seeded in the same manner and treated with TRA-8 alone (25 and 125 ng/mL) for 2 hours. Anti–total β-catenin and anti-actin antibodies were used.

**TOPflash luciferase reporter assay**

NAAE cells from all four BLBC cell lines were plated at 40,000 cells per well. The next day, the cells were transfected with 0.05 mg of TCF/lymphoid enhancer factor (LEF) activity reporter (TOPflash; plasmid from Dr. Randall Moon’s laboratory, Upstate Biotechnology). Cells were transfected using Lipofectamine 2000 (Invitrogen) in Opti-MEM (Gibco/Invitrogen) as per the manufacturer’s instructions. Six hours after transfection, cells were treated with 0.25 μmol/L niclosamide for 24 hours. Adherent BLBC cells were plated and transfected in the same manner. In one experiment, adherent cells were treated with 0.25 μmol/L niclosamide for 48 hours and 0.25 ng/mL TRA-8 for 24 hours. In another experiment, adherent SUM159, HCC1143, and HCC1187 cells were treated with 1 μmol/L niclosamide and 2LMP cells with 0.25 μmol/L niclosamide for 24 hours. Each cell line was also concurrently transfected with Wnt3A (5 ng/mL; R&D Systems) alone and in combination with niclosamide.

Patient samples were plated in the same manner and transfection was attempted with the plasmid, but obtaining a consistently high transfection efficiency for the analysis proved challenging. Therefore, the method was revised to use lentivirus pCignal Lenti TCF/LEF Reporter at multiplicity of infection (MOI) of 25 (Qiagen) to transfect the cells. Twenty-four hours after transfection with the lentivirus, cells were treated with 4 μmol/L of niclosamide for 24 hours. Total protein was harvested 24 hours after treatment and luciferase activity was measured using a Turner 20/20 luminometer (Promega). The luciferase reading was normalized to the total protein concentration as reported previously (39). The luciferase activity was normalized to untreated control and represented as a mean ± SE for a minimum of three replicates.

**Tumorsphere inhibition assay**

NAAE cells for all four BLBC cell lines were plated at 2,000 cells/well in 50 μL media, then treated with niclosamide (0.25–1.0 μmol/L) for 24 hours, followed by treatment with TRA-8 (1 ng/mL 2LMP, 0.5 ng/mL SUM159, 25 ng/mL HCC1143, and 5 ng/mL HCC1187) for an additional 24 hours. Two independent investigators used a reticle eyepiece to visually count tumorspheres. Mean tumorsphere inhibition was calculated relative to no treatment. Samples were run in quadruplicate in three independent experiments and error bars represent ± SE.

**In vitro cytotoxicity studies**

2LMP, SUM159, HCC1143, HCC1187, NAAE and attached cells, MCF10A, LRP6 knockdown, or shRNA-
attached cells were seeded at 2,000 cells/50 μL of media. Adherent cells were plated in optically clear tissue culture–treated black plates (Costar). NAAE cells and patient samples were plated in low-attachment plates in MEGM media. To assess the viability of cells exposed to niclosamide or TRA-8, all cells were treated with niclosamide (0.125–8 μmol/L) or TRA-8 (0.1–500 ng/mL) for 24 or 48 hours. In combination treatment studies, after 24 hours exposure to niclosamide, TRA-8 (0.25–500 ng/mL) was added for an additional 24 hours before the cells were lysed and analyzed for viability using the APLite luminescence-based assay (PerkinElmer) as previously described (40). Alternatively, cells were treated concurrently with niclosamide and TRA-8 for 48 hours. TRA-8 and niclosamide were diluted in culture medium immediately before use, and the final concentration of DMSO was always ≤0.001%. All samples were assayed in triplicate or quadruplicate and are reported as the mean ± SE.

**In vivo therapy and toxicity studies**

Female athymic nude mice at 4 weeks of age were purchased from Harlan. *In vivo* generation of tumors was accomplished by resuspending 2 × 10^5 2LMP cells in 100 μL (1:1 Matrigel and PBS) and injecting cells into the MFP of athymic nude mice (*n* = 5). Mice were randomly assigned into four groups and treated when the tumors reached approximately 16 mm^2 (7 days after implantation). Mice were intraperitoneally injected twice weekly with TRA-8 (200 μg), niclosamide (30 mg/kg; 50% cremophore EL:50% DMSO) 5 days a week for 3 weeks. Tumor size (surface area) was calculated by a × b, in which *a* is the largest diameter and *b* is the diameter perpendicular to *a*. Tumors were measured with a Vernier caliper 2 times a week until mice were euthanized in accordance with the IACUC regulations.

To assess tumor and normal tissue toxicity, a separate animal study was conducted with three mice per group and treated with vehicle control 5 days a week, niclosamide (20 mg/kg) 5 days a week, TRA-8 (200 μg) twice weekly, and the combination for a total of 2 weeks. All animals were monitored daily and weighed twice a week. After 2 weeks of treatment, animals were euthanized and tumor tissue, large and small intestines, kidney, spleen, and liver were dissected and fixed in 10% neutral buffered formalin at room temperature and then processed to paraffin blocks. The tissues from each of the four groups were coded and hematoxylin and eosin stains were performed on 4-μm tissue sections. The board-certified diagnostic pathologist who evaluated the tissues for toxicity was blinded about the treatment group. In addition, portions of the tumors were evaluated for changes in protein expression by Western blot analysis.

**Immunohistochemistry**

Patient pleural effusion samples were stained by immunohistochemistry for LRP6, Moc31, and calretinin (Cell Signaling Technology, Inc.). Mouse tumor tissue was also analyzed for p-STAT3 and β-catenin expression. Cells were harvested into histogel and embedded into a paraffin block. For each sample, the staining intensity (0, 1+, 2+, and 3+) and the percentage of cells staining positive (0%–100%) was determined by a blinded board-certified pathologist. An H-score was calculated as the product of the intensity and the percentage of cells with positive staining.

**Statistical analysis**

*T* tests were used to compare means when appropriate. General linear models were used to make comparisons over time. Enhanced combination effect was further confirmed by a concentration–effect curve using the nonlinear regression method and isobologram methods (data not shown; ref. 41). Experimental animal treatment groups were composed of 5 animals each to provide evidence of substantial tumor sensitivity to TRA-8 therapy with or without niclosamide. For xenograft models, tumor doubling time (TDT) was estimated for each animal using empirical distribution, and median TDT between treatment groups was compared using the Kruskal–Wallis nonparametric statistical test.

**Results**

**Analysis of cells**

In this study, we investigated four BLBC cell lines: 2LMP, SUM159, HCC1143, and HCC1187. All cell lines were analyzed as adherent cells in normal tissue culture plates with media that contained serum. In addition, cells were plated in ultralow attachment plates with MEGM media and were noted to form tumorspheres, which are composed of nonadherent cells. Nonadherent cells were further analyzed for the stem cell marker, ALDH, at various time points and compared with cells that were sorted for ALDH. Both types of cells were noted to have elevated levels of ALDH 12 hours after being plated, which then decreased with time. The nonsorted and nonadherent aldehyde-enriched cells at the 12-hour time point were termed NAAE.

**Characterization of NAAE cells**

Cells were tested for ALDH activity at 6, 12, 24, 48, and 72 hours for the 2LMP and SUM159 cell lines and at 6, 12, and 24 hours for the HCC1143 cells. These cells were compared with cells that were sorted for the ALDH population and subsequently analyzed for ALDH expression at the same time points. Both sorted and nonsorted cells were found to be enriched for ALDH at the 12-hour time point, but subsequently both populations returned to the presorted expression level; therefore, the NAAE cells represent a CSC-enriched population at the 12-hour time point (Fig. 1A). To verify enhanced tumorigenicity of the NAAE cells (12 hours after plating), they were injected into mice and compared with cells that were cultured for 72 hours before injection, which had reduced ALDH activity. The NAAE cells formed aggressive large tumors,
indicating their enrichment for CSCs compared with tumors formed by cells from the 72-hour time point ($P = 0.01$). NAAE xenografts developed earlier and formed larger tumors over time, indicating enrichment for tumor-initiating CSCs (Fig. 1B).

2LMP NAAE cells were also analyzed by Western blot for p-LRP6, LRP6, free and total β-catenin (Fig. 1C) and compared with adherent 2LMP cells. The NAAE cells demonstrated higher free β-catenin and lower LRP6 and p-LRP6 expression. Similar levels of total β-catenin were observed in both NAAE and adherent populations.

**Niclosamide induces cytotoxicity to BLBC cell lines in vitro**

We examined the cytotoxicity of niclosamide against both adherent cells and NAAE cells in BLBC cell lines. All four BLBC adherent cell lines were sensitive to niclosamide with an IC$_{50}$ range of 0.33 to 1.9 μmol/L (Supplementary Table S1). NAAE cell populations from BLBC cell lines were more sensitive to niclosamide than their corresponding adherent cell populations with IC$_{50}$ values of 0.17 to 0.29 μmol/L (Supplementary Table S1). Statistical significance was not met in the 2LMP NAAE compared with the adherent cells (0.29 vs. 0.44; $P > 0.05$), but the SUM159, HCC1187, and HCC1143 cell lines had a statistically significant difference in the adherent and the NAAE IC$_{50}$ values. Adherent MCF10A (nontumorigenic) mammary epithelial cells showed less than 10% niclosamide-mediated cytotoxicity (Supplementary Fig. S1).

**Niclosamide inhibits Wnt/β-catenin and STAT3 signaling in BLBC cell lines**

To characterize the effect of niclosamide on the Wnt/β-catenin pathway, we performed the signaling TOP-flash luciferase reporter assay, which measures the level of nuclear β-catenin, on NAAE cells and adherent cells. Wnt3A ligand was added to the four adherent BLBC cell lines to upregulate the Wnt/β-catenin pathway. Nuclear β-catenin was decreased by niclosamide treatment in all four adherent cell lines (0.25 μmol/L in the more sensitive cell line, 2LMP, and 1 μmol/L in the other three cell lines) after upregulation by Wnt3A ligand ($P < 0.05$). Three of the four cell lines had niclosamide-mediated Wnt/β-catenin inhibition in the absence of the Wnt3A ligand ($P < 0.05$; Supplementary Fig. S2). NAAE cells from all four BLBC cell lines showed significant inhibition of Wnt/β-catenin signaling following 24 hours of treatment with 0.25 μmol/L niclosamide without the addition of the Wnt3A ligand ($P < 0.01$; Fig. 2A).

Western blot analysis of Wnt/β-catenin signaling proteins (LRP6, pLRP6, free and total β-catenin), three target genes (Axin2, survivin, and cyclin D1), and two STAT3 pathway proteins [phosphorylated (tyr705) and total STAT3] after 24 hours treatment with niclosamide (0.125-0.5 μmol/L) showed a dose–response inhibition of the Wnt/β-catenin and STAT3 pathways in both the adherent and NAAE 2LMP cells (Fig. 2B). Cell death was also calculated for these doses to assure that Western results were not a consequence of cell death. The highest
dose of 0.5 \( \mu \text{mol/L} \) did not induce more than 60% kill of the adherent or the NAAE cells when cells were plated according to the Western blotting protocol (Supplementary Fig. S3). The 0.125-\( \mu \text{mol/L} \) dose caused almost no cell kill, yet did produce decreases in the Wnt/\( \beta \)-catenin target genes, suggesting that the Wnt/\( \beta \)-catenin pathway is downregulated at noncytotoxic doses of niclosamide.

**Effect of niclosamide and TRA-8 treatment in vitro**

Our laboratory has previously published on TRA-8 cytotoxicity to BLBC cells (35) and their CD44\(^+\)/CD24\(^-\)/ALDH\(^+\) cell-sorted populations (34); therefore, we hypothesized that the combination of two CSC-targeting agents, TRA-8 and niclosamide, would produce increased cytotoxicity of NAAE cells. NAAE cells were pretreated with niclosamide for 24 hours followed by 24 hours treatment with TRA-8, and the number of tumorospheres was compared with single agents or no treatment. Tumorsphere formation was inhibited in all four cell lines with niclosamide and TRA-8 alone, but the combination of the two agents produced a significant increase in tumosphere inhibition compared with either agent alone (\( P = 0.03 \); Fig. 3A). Cytotoxicity of niclosamide and TRA-8 was also tested against NAAE cells (Fig. 3B). The cytotoxicity results correlated with the tumosphere inhibition results in all four cell lines.

**Effect of niclosamide and TRA-8 on Wnt/\( \beta \)-catenin signaling**

The TCF/LEF plasmid reporter TOPFlash assay was used to evaluate the combination of niclosamide and TRA-8 on the Wnt/\( \beta \)-catenin signaling pathway in BLBC adherent cells. The TOPFlash assay was also attempted for the combination treatment on NAAE cells, but sufficient transfection was never achieved. All four BLBC cell lines were treated for 48 hours with 0.25 \( \mu \text{mol/L} \) of niclosamide. After the first 24 hours, TRA-8 at 0.25 ng/mL was added. Wnt/\( \beta \)-catenin pathway inhibition occurred after treatment with niclosamide or TRA-8 alone as compared with no treatment (Fig. 4A).
Combination treatment produced significantly greater inhibition of the Wnt/β-catenin reporter than single agents in three of the cell lines.

TRA-8, unlike niclosamide, was previously not known to influence Wnt/β-catenin signaling. Western blots were used to examine the effect of TRA-8 on β-catenin. Total β-catenin levels decreased (low exposure) with evidence of fragmentation (high exposure) of the protein after a 2-hour TRA-8 treatment of 2LMP adherent and NAAE cells. However, with HCC1187, TRA-8 treatment only produced a decrease in total β-catenin in NAAE cells (Fig. 4B).

Given our observation that niclosamide’s inhibition of the Wnt/β-catenin pathway resulted in reduced expression of survivin, we hypothesized that decreasing LRP6 would enhance the cytotoxicity of the apoptotic agent TRA-8. Knockdown of LRP6 using Mission Lentiviral transduction particles shRNA on adherent 2LMP cells produced enhanced TRA-8–mediated cytotoxicity (Fig. 4C). LRP6 knockdown was confirmed by Western blot, which also showed reduced survivin expression compared with a vector control (Supplementary Fig. S4).

Effect of niclosamide and TRA-8 on established 2LMP tumors

Given the impressive in vitro observations of niclosamide and TRA-8 cytotoxicity with BLBC cell lines, we carried out a combination therapy study in an orthotopic BLBC animal model. Animals received niclosamide (30 mg/kg 5 days a week), or TRA-8 (200 μg twice weekly), or both treatments for a total of 3 weeks and were compared with mice that received a vehicle control. The mice that received TRA-8 alone had a significant inhibition of tumor growth (P < 0.05), similar to what had been previously reported by our laboratory (35). Animals treated with niclosamide had similar tumor growth inhibition compared with those treated with TRA-8, whereas animals treated with the combination of agents had significant tumor growth inhibition compared with untreated controls (P < 0.01) and to either single agent alone (P < 0.05; Fig. 5).

To further investigate the potential toxicity of niclosamide and TRA-8 in the mice, we performed a 2-week in vivo treatment of both single agents and the combination. Although this treatment regimen of niclosamide was 20 mg/kg, there was still a significant decrease in tumor	

Figure 3. The effect of TRA-8, niclosamide, and the combination on tumorsphere formation and cytotoxicity in NAAE cells. A, NAAE cells were cultured with media alone, pretreated with niclosamide (0.25–1.0 μmol/L) for 24 hours, followed by TRA-8 (1 ng/mL 2LMP, 0.5 ng/mL SUM159, 25 ng/mL HCC1143, and 5 ng/mL HCC1187) for 24 hours, for a total of 48 hours treatment. Tumorsphere formation was determined in four replicates and three separate experiments. (P < 0.05 treatment vs. control; P < 0.05 combination vs. either agent alone). Additive drug interaction against tumorsphere inhibition was observed for all four cell lines (P = 0.03). B, cell viability of drug combination was also tested on 2LMP, SUM159, HCC1143, and HCC1187 NAAE cells. 2LMP and SUM159 cells were pretreated for 24 hours with (0.25–1.0 μmol/L) niclosamide, followed by TRA-8 (1 ng/mL 2LMP and 0.5 ng/mL SUM159 for 24 hours, for a total of 48 hours). HCC1143 and HCC1187 were treated concurrently with (0.25 μmol/L) niclosamide, and TRA-8 (25 ng/mL HCC1143 and 5 ng/mL HCC1187), for a total of 48 hours treatment.
growth in both the single-agent treatment and further inhibition with the combination treatment. The weight of the treated mice, both with a single agent and the combination did not decrease over time (Supplementary Fig. S5). In addition, the tissue architecture for the kidney, spleen, liver, small and large intestines was analyzed and found to be normal. Furthermore, the tumors in the mice were analyzed by immunohistochemistry, which showed that treated mice had reduced expression of p-STAT3, cytosolic, and nuclear β-catenin, and increased expression of cleaved caspase-3 with niclosamide, TRA-8, and the combination treatment (Supplementary Table S2). This shows that niclosamide, TRA-8, and the combination repress tumor growth by inhibiting the Wnt/β-catenin pathway, the STAT3 pathway, and by causing apoptosis. Western blot analysis of tumor tissue resulted in reduced expression of survivin in niclosamide-treated mice and further reduction with combination treatment (results not shown). These results further demonstrate the combination effect of TRA-8 and niclosamide.

Primary BLBC patient samples response to niclosamide and TRA-8

The combination of niclosamide and TRA-8 was further investigated on patient BLBC tumor cells collected from pleural effusions from triple-negative treatment-resistant patients. All four samples (UAB01, UAB03, UAB04, and UAB05) stained positive for LRP6 by immunohistochemistry (data not shown). All four patient samples showed significant cytotoxicity when treated with niclosamide for 48 hours (1–8 μmol/L; Fig. 6A). TOPflash assay using lentiviral reporter was performed on UAB03 and UAB05 because these two samples had adequate number of cells available, whereas UAB01 and UAB04 had too few cells. The decrease in percentage luciferase activity in the TCF/LEF viral reporter after 24 hours of treatment with 4 μmol/L niclosamide showed Wnt/β-catenin pathway inhibition in these two samples (Fig. 6B). Patient sample UAB03 also had enough cells for Western blot assay to be performed. Niclosamide demonstrated dramatic inhibition of Wnt/β-catenin pathway proteins (LRP6 and β-catenin) and cyclin D1, as well as reduced p-STAT3 and total STAT3 expression in this patient sample when treated for 24 hours with 4 μmol/L niclosamide (Fig. 6C). When niclosamide and TRA-8 treatment was combined in two patient samples, UAB05 and UAB03, tumor cell cytotoxicity was enhanced compared with TRA-8 alone for patient sample UAB05, and a modest combination effect was seen in patient sample UAB03 (Fig. 6D). The higher concentrations of niclosamide that were needed for cytotoxic, Western, and TOPflash effects in the patient samples compared with the cell lines may reflect the drug-resistance history and metastatic nature of the patient samples. Cells collected from two BLBC patient pleural effusion samples
Discussion

Most cancer treatment regimens assume that all cancer cells have equal malignant potential and respond similarly to therapy. More recently, therapies are being designed to target both bulk tumor cells and CSCs with the goal to prevent recurrence and metastasis. Therefore, therapies that target both the CSC maintenance pathways and induce apoptosis in the bulk tumor cell population could potentially result in increased chances for survival. We previously described that TRA-8 kills BLBC cell lines both in vitro and in xenograft mouse models and preferentially kills their CSC population. In an effort to improve on this strategy, this study demonstrated that niclosamide, a potent inhibitor of Wnt/β-catenin signaling, produced similar cytotoxicity with bulk tumor cells (attached cells), CSC-enriched cell populations (NAAE cells), and primary tumor cells from patients with BLBC both alone and in combination with TRA-8.

Because CSCs represent a small fraction of the total tumor cell population, it is difficult to run mechanistic studies with these cells especially by techniques such as Western blot and the TOPflash reporter assay that require a large number of cells. NAAE cells were discovered as a tool to analyze the effect of drugs on CSC-enriched populations. Our findings imply that sorting may not be necessary to enrich for CSC characteristics if treatments are conducted within a short time period after plating. Our results also indicate that treatments targeted toward CSCs should not be carried out on sorted populations after 48 hours from sorting due to depletion of initial enrichment for CSC marker expression. In vivo tumorigenicity studies functionally validated the enhanced tumor take and growth rate of NAAE cells. Future studies will characterize NAAE cells and look at epithelial–mesenchymal transition and mesenchymal–epithelial transition markers that could potentially influence observed dynamic CSC marker expression (42).

The involvement of Wnt/β-catenin signaling in BLBC and CSCs has been well documented (14). High-throughput screening studies identified niclosamide as a potent Wnt/β-catenin pathway inhibitor (43). Multiple publications by various groups have shown that niclosamide is a potent inhibitor of Wnt/β-catenin signaling (22–24). Like many inhibitors of signaling pathways, niclosamide also affects other pathways, such as NF-κB (26), Notch (44), reactive oxygen species (26), mammalian target of rapamycin complex 1 (36), and STAT3 (45). Niclosamide has been shown to be cytotoxic in cell lines from several cancers such as colon (46), ovarian (28), osteosarcoma (23), myeloma (47), and breast cancer (22). Our objective was to investigate niclosamide’s cytotoxicity in BLBC cell lines and BLBC patient pleural effusion samples. NAAE cells were generated from BLBC cell lines to specifically test the sensitivity of CSCs to niclosamide. The Wnt/β-catenin pathway is known to play an important role in CSCs; the sensitivity of NAAE cells to niclosamide treatment further supports the role of the Wnt/β-catenin pathway in cells enriched for stem cell markers. Furthermore, Western blot analysis of NAAE cells showed that they have reduced levels of LRP6, thus their enhanced sensitivity to niclosamide is not due to overexpression of LRP6. NAAE cells did show elevated levels of free β-catenin, which could contribute to their increased sensitivity to niclosamide. Given there was no elevation in LRP6 or p-LRP6 in the NAAE cells, other pathways such as STAT3 could be the mechanism by which nuclear β-catenin is increased. Prior studies have shown that activation of the STAT3 pathway leads to increased levels of β-catenin (48).

The BLBC cells isolated from patient samples that showed cytotoxicity with niclosamide treatment were previously resistant to in vitro treatment with chemotherapy and Notch inhibitors (data not shown). The dose of niclosamide that was needed to cause in vitro cytotoxicity of these patient samples was within the range of blood concentrations (0.76–18.35 μmol/L) achieved when healthy patients were given 2 g of niclosamide orally once for the treatment of tapeworms (25).

Both TOPflash results and Western blot analysis showed functional inhibition of the Wnt/β-catenin signaling pathway in BLBC cell lines and patient samples treated with niclosamide. Niclosamide also caused a
decrease in p-STAT3, which shows that it affects other pathways, in addition to the Wnt/b-catenin pathway involved in tumor initiation and progression. The ability of niclosamide to deplete survivin expression could be the mechanism by which there is an increase in apoptosis when BLBC cells are treated with TRA-8. Survivin normally inhibits caspase-9 activation within the apoptosome, a key step in the apoptotic cascade (49). The 2LMP LRP6 knockdown cells had enhanced sensitivity to TRA-8, and reduced survivin activity, further validating our hypothesis that inhibition of canonical Wnt/b-catenin signaling can sensitize cells to death receptor–mediated apoptosis.

The extrinsic apoptotic pathway has been shown to regulate b-catenin activity by TRAIL’s ability to decrease full-length b-catenin expression (30, 50). Although TRAIL can induce apoptosis in tumor cells, many cancer cells develop resistance, which could be overcome by combination treatment. b-Catenin can be targeted to sensitize apoptosis-resistant cancer cells. TRA-8 and niclosamide led to reduction of b-catenin expression, coinciding with maximal apoptosis. The ability of TRA-8 to reduce b-catenin was confirmed by TOPflash reporter assay and Western blot analysis of cleaved b-catenin. Future studies will focus on characterizing the mechanism by which TRA-8 can inhibit b-catenin activity and the results could help further elucidate how TRA-8 preferentially induces apoptosis in CSCs.

Overall, this study provides new insights into the treatment of CSCs from BLBC. It further supports existing literature that niclosamide’s inhibition of Wnt/b-catenin signaling and reduction in p-STAT3 produces enhanced cytotoxicity against CSCs. In addition, the results demonstrate that the combination of niclosamide and TRA-8 targets the CSC subpopulation and potentially prevents recurrence and metastasis in patients with BLBC. The combination of these two agents or similar compounds can potentially be translated into clinical trials, helping patients with BLBC who have very few treatment options available to them.
Disclosure of Potential Conflicts of Interest

A.F. LoBuglio has ownership interest in anti-DR5. D.J. Buchsbaum has ownership interest in U.S. Patent Nos. 7,279,160, 7,476,383, and 7,704,502. No potential conflicts of interest were disclosed by the other authors.

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