Therapeutic Discovery

Impact of Intrinsic Affinity on Functional Binding and Biological Activity of EGFR Antibodies

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Abstract

A aberrant expression and activation of EGF receptor (EGFR) has been implicated in the development and progression of many human cancers. As such, targeted therapeutic inhibition of EGFR, for example by antibodies, is a promising anticancer strategy. The overall efficacy of antibody therapies results from the complex interplay between affinity, valence, tumor penetration and retention, and signaling inhibition. To gain better insight into this relationship, we studied a panel of EGFR single-chain Fv (scFv) antibodies that recognize an identical epitope on EGFR but bind with intrinsic monovalent affinities varying by 280-fold. The scFv were converted to Fab and IgG formats, and investigated for their ability to bind EGFR, compete with EGF binding, and inhibit EGF-mediated downstream signaling and proliferation. We observed that the apparent EGFR-binding affinity for bivalent IgG plateaus at intermediate values of intrinsic affinity of the cognate Fab, leading to a biphasic curve describing the ratio of IgG to Fab affinity. Mathematical modeling of antibody–receptor binding indicated that the biphasic effect results from nonequilibrium assay limitations. This was confirmed by further observation that the potency of EGF competition for antibody binding to EGFR improved with both intrinsic affinity and antibody valence. Similarly, both higher intrinsic affinity and bivalent binding improved the potency of antibodies in blocking cellular signaling and proliferation. Overall, our work indicates that higher intrinsic affinity combined with bivalent binding can achieve avidity that leads to greater in vitro potency of antibodies, which may translate into greater therapeutic efficacy. Mol Cancer Ther; 11(7); 1467–76. ©2012 AACR.

Introduction

EGF stimulates cell proliferation in normal tissue and in malignant lesions by activating EGF receptor (EGFR) tyrosine kinase signaling pathways (1–3). EGFR is frequently overexpressed in many cancers including breast, colon, head and neck, glioblastoma, gastric, and squamous cell carcinoma (4, 5) and is associated with more aggressive cancer subtypes (6, 7). Anti-EGFR monoclonal antibodies cetuximab and panitumumab are used to treat metastatic colorectal cancers as a single agent or in combination with chemotherapy (8, 9). Both cetuximab and panitumumab inhibit cancer cell proliferation by blocking EGF signaling (10–13) although they can induce immune effector functions as well (14, 15).

Antibodies are attractive candidates for cancer therapeutics not only because of their exquisite specificity to the target and ability to inhibit cell signaling, but also because of their long half-life in serum and the ability to induce tumor antigen-specific immune responses (11, 16). Previously, studies of antibody mechanism of action have revealed insights useful for guiding the engineering of therapeutic antibodies to enhance their antitumor effects, including the importance of target downregulation and signaling blockade by antibody (11, 17), the requirement of Fc to activate Fc receptors for in vivo efficacy (18), and the impact of antibody affinity and avidity on in vivo tumor targeting (19–22). With respect to antibody affinity effects on tumor targeting, studies have shown that increased binding affinity and valence leads to improved targeting of tumoral vasculature (22). However, we and others have shown that tumor targeting is maximized for antibodies with intrinsic monovalent affinities of $K_D$ in the 1 to 10 nmol/L range, regardless of whether the targeting antibody is a small single-chain Fv (scFv) or bivalent diabody or IgG (19, 21, 23–27). In fact, high affinity IgG that bind internalizing receptors may be more rapidly degraded in tumors than lower affinity IgG, limiting tumor penetration (28). These so called “binding site
barriers" (29–31) may result in higher affinity antibodies having lower quantitative tumor delivery.

Despite the presence of a number of studies examining the impact of affinity and valence on in vivo targeting, we can find no data on the impact of intrinsic and apparent affinity on biologic activities of antibodies where the antibody affinity variants bind identical epitopes on the tumor antigen. This knowledge is especially crucial for EGFR-targeting antibodies as their clinical efficacy is thought to rely substantially on the ability to inhibit EGFR-driven cell signaling and proliferation. Therefore, for this work we studied a panel of EGFR scFv antibodies, which varied 280-fold in intrinsic affinity and bound an identical EGFR epitope. By creating Fab and IgG from the scFv, we could dissect the importance of intrinsic affinity and valence-mediated avidity on tumor cell binding, EGFR phosphorylation, and inhibition of tumor cell proliferation.

Materials and Methods

Cell lines

A431, MDAMB468, and MDAMB231 cells were purchased from the American Type Culture Collection (March 2001) and cultured in RPMI 10% FBS and L15 10% FBS media, respectively. Although the cells were not authenticated and tested, mycoplasma contamination was checked regularly.

Antigen and antibodies

Recombinant EGFR-ECD was expressed and purified as previously described (32). Control antibodies include anti-EGFR antibody C225 (cetuximab, Erbitux; Lilly; ref. 11), anti-HER2 antibody C6.5 and anti-BoNT antibody 3D12 (33–35). Anti-EGFR antibody Ab11 (NeoMarkers/LabVision) was used for K_D measurement by KinExA and Ab12 (NeoMarkers/LabVision) for EGFR detection by Western blotting.

Production of anti-EGFR affinity mutant scFv, IgG, and Fab fragments

scFvs with varying affinities for EGFR (36) were subcloned from the yeast-display vector pYD2 (37) via NcoI–NotI into expression vector p5yn1 for scFv expression or for IgG expression (33). scFvs were produced in the periplasm of Escherichia coli strain TG1, and purified by osmotic shock and immobilized metal affinity chromatography, as reported previously (38). Monomeric scFv were separated from dimeric and aggregated scFv by size-exclusion chromatography (Sephadex G75; Amersham Pharmacia) with PBS as eluant. IgG proteins were secreted in the media by transfected CHO cells as described previously (33) and purified by Protein G affinity chromatography (Amersham Pharmacia). Fab fragments were generated by papain digestion (Pierce) of the corresponding IgG followed by separation using Mono S ion-exchange chromatography (GE Healthcare). The homogeneity and purity of the protein preparations were verified by SDS-PAGE stained with Coomassie blue; protein concentrations were measured by microbicinchoninic acid assay (Pierce).

Cell surface–binding measurements

Cell lines that express EGFR were grown to 80% to 90% confluence in their respective media supplemented with 10% FBS and harvested by trypsinization. Each scFv, Fab, or IgG was incubated with 5 × 10^5 cells for 16 hours at the indicated concentration. Cell binding was carried out at 4°C in PBS containing 1% FBS in adequate volume to maintain constant antibody concentration for equilibrium conditions. After 2 washes with 200 μL of PBS, bound scFv was detected by the addition of 100 μL (1 μg/mL) of PE-labeled anti-human Fc-specific F(ab‘)_2 (Jackson ImmunoResearch); bound Fab was detected by the addition of 100 μL (1 μg/mL) of PE-labeled anti-human Fab-specific F(ab‘)_2 (Jackson ImmunoResearch). After incubating 30 minutes at 4°C, the cells were washed twice and resuspended in PBS containing 4% paraformaldehyde. Fluorescence was measured by flow cytometry using a FACS LSRII (Becton Dickinson), and median fluorescence intensity (MFI) was calculated using CellQuest software (Becton Dickinson). Equilibrium constants were determined as described (39), except that values were fitted to the equation MFI = MFI_{min} + MFI_{max} × [Ab]/(K_D + [Ab]) using KaleidaGraph software.

Effects of EGF on the binding of EGFR antibodies to EGFR-overexpressing cells

The binding assay was conducted as described earlier, except that the antibodies were incubated with EGF at the indicated concentration.

Effects of anti-EGFR affinity mutants on cell proliferation

A431 cells were seeded at 3 × 10^5 cells per well in 96-well plates (Costar) in RPMI media containing 0.5% FBS and incubated for 5 days with the indicated concentrations of scFv, Fab, or IgG, as described in the figure legends. Cell proliferation was assayed as described using MTT staining (40).
The proteins were transferred to nitrocellulose and immunoblotted with antiphosphotyrosine antibody PY20 (Upstate Biotechnology). The total EGFR level in each condition was evaluated with anti-EGFR antibody Ab12—a cocktail of 4 anti-EGFR antibodies to both extracellular and intracellular epitopes.

Determination of affinity constant and binding kinetics by KinExA

Affinity studies of antibody and EGFR reaction mixtures were conducted in PBS (pH 7.4) at room temperature with 1 mg/mL bovine serum albumin (BSA) and 0.02% (w/v) sodium azide added as a preservative. Antibody solution was serially diluted into a constant concentration of EGFR sufficient to produce a reasonable signal. The range of antibody concentration was varied at least 10-fold above and below the value of the apparent K_D and antigen concentrations were set below twice the K_D to ensure a controlled experiment. Sample solutions were allowed to come to equilibrium, then each in the series of 13 dilutions was passed over a flow cell containing a 4-mm column of freshly packed NHS-activated Sepharose 4 Fast Flow beads (GE Healthcare) covalently coated with an antibody binding EGFR at the experimental epitope to capture the unbound antigen from solution. An Alexa-647–labeled antibody Ab11 binding EGFR at a separate epitope was then passed over the bead column to detect the proportion of free antigen. Each dilution was tested in duplicate. The equilibrium titration data were fit to a 1:1 reversible binding model using KinExA Pro Software (version 3.0.6; Sapidyne Instruments) to determine the K_D. Antibody–antigen-binding kinetics were also conducted using the KinExA 3000 to quantify the decrease in free antigen as mixtures of antibody and antigen came to equilibrium. A single mixture of antibody solution and EGFR solution was prepared for each interaction and measured on the instrument by being passed in 0.5 mL volumes over a fresh antigen-binding bead pack at intervals of approximately 550 seconds. Free antigen was detected with Alexa-647–labeled anti-EGFR antibody as described earlier. The exponential decrease in the concentration of free antigen as a function of time was fit to a standard bimolecular rate equation using the KinExA Pro software to determine the k_on. The k_off was calculated from the product of k_0 × k_D.

Quantification of EGFR cell surface density

EGFR cell surface density was quantified with Quantum Simply Cellular anti-human IgG bead standards (QSC beads; Bangs laboratories). The QSC beads, coated with different density of anti-Human Fc-specific antibody were stained with 1 nmol/L of C225 IgG. At the same time the reverse rate constant for each antibody was calculated as the product of the 2224 IgG rate and the ratio of measured cell-binding affinity of the relevant Fab and 2224 Fab (Supplementary Table S2).

Bivalent binding of EGFR on the cell surface is in addition described in the model by a parameter characterizing the ability of the antibody to cross-link receptors. This parameter was determined computationally, based on a least-squares optimization of the model against the experimental binding dose–response curves. All antibodies were assumed to share a common value for this cross-linking parameter. Antibody affinities were calculated for each simulation as described for the experimental cell surface–binding measurements. All computational methods were developed using MATLAB version 7.12 (The Mathworks, Inc.).

Results

EGFR antibodies studied

The human EGFR C10 scFv was previously isolated by selecting a nonimmune human scFv phage antibody library on CHO cells transfected with EGFR (42). C10 bound EGFR expressing A431 cells with a K_D of 264 nmol/L. The affinity of the C10 scFv for EGFR was increased by using yeast display (36). Seven mutant EGFR scFv antibodies (P3/5, P2/1, P2/4, P2/2, 3524, 2124, and 2224) were isolated with affinities ranging from 88.24 to 0.94 nmol/L as measured on A431 cells (Table 1). V_{I4} and V_{I5} genes from scFv were subcloned for expression as human IgG1/lambda isotype IgG, stable CHO cell lines established, and the resulting IgG purified. Fab fragments
were then enzymatically prepared from the IgG molecules. The parental C10 and higher affinity mutant scFv, Fab, and IgG molecules were used in the present studies to evaluate the impact of intrinsic and apparent antigen-binding affinity on in vitro biological effects.

Intrinsic and apparent affinities for EGFR antibody binding to EGFR-overexpressing cancer cells

Using flow cytometry, we characterized the binding affinity of scFv-derived EGFR Fab and IgG molecules for cell surface EGFR on A431 cells (Table 1). In all cases, we found that bivalent IgG had higher observed binding affinity than the cognate monovalent Fab. Previous work in our laboratory had indicated that the affinity gain upon conversion from the monovalent to the bivalent format is higher for low affinity binders (21). Here, however, we found that the ratio of IgG to Fab affinities varied in a biphasic fashion (Table 1 and Supplementary Fig. S1). The intermediate affinity Fab P2/4 showed a very large affinity gain (4,000-fold) upon conversion to bivalent IgG, whereas the Fab clones of low (C10) and high (2224) monovalent affinity showed only 100- to 200-fold affinity gains. C225, the highest affinity Fab, showed the lowest improvement as an IgG (only about 2-fold). In comparing the IgG affinities of P2/4, 2224, and C225, we found that IgG of nearly identical apparent affinity can derive from Fab with widely varying intrinsic affinities (over 3 orders of magnitude between C225 and P2/4). The ratio $K_D$(Fab)/$K_D$(IgG) represents the improvement of apparent affinity from the bivalent binding. The bivalent binding actually increases the functional concentration of antibody compared with the monovalent form, but this effect will diminish when the $k_{off}$ is slow enough for antibodies with high intrinsic affinities.

To understand this biphasic effect, we developed a simple computational model of IgG binding to free cell surface receptors to simulate the observed binding curves. The model incorporates the experimentally measured IgG binding kinetics (Supplementary Table S1) and EGFR expression levels, as well as experimental details such as experimental incubation time and antibody concentration. The computational model of IgG binding accurately described the affinities measured for the anti-EGFR IgG (Table 2, second column). In addition, the model was

### Table 1. Comparison of affinities of scFv, Fab, and IgG for binding to EGFR-overexpressing A431 cells

<table>
<thead>
<tr>
<th>Antibody</th>
<th>scFv $K_D$, nmol/L</th>
<th>Fab $K_D$, nmol/L</th>
<th>IgG $K_D$, nmol/L</th>
<th>$K_D$(Fab)/$K_D$(IgG)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C10</td>
<td>263.67</td>
<td>124.23</td>
<td>1.17</td>
<td>106</td>
</tr>
<tr>
<td>P3/5</td>
<td>88.24</td>
<td>58.24</td>
<td>0.5</td>
<td>116</td>
</tr>
<tr>
<td>P2/1</td>
<td>14.81</td>
<td>25.4</td>
<td>0.012</td>
<td>2,117</td>
</tr>
<tr>
<td>P2/4</td>
<td>15.39</td>
<td>25.2</td>
<td>0.0064</td>
<td>3,938</td>
</tr>
<tr>
<td>P2/2</td>
<td>17.01</td>
<td>18.1</td>
<td>0.0077</td>
<td>2,351</td>
</tr>
<tr>
<td>3524</td>
<td>7.47</td>
<td>15.4</td>
<td>0.012</td>
<td>1,903</td>
</tr>
<tr>
<td>2124</td>
<td>9.90</td>
<td>1.31</td>
<td>0.007</td>
<td>187</td>
</tr>
<tr>
<td>2224</td>
<td>0.94</td>
<td>1.2</td>
<td>0.007</td>
<td>171</td>
</tr>
<tr>
<td>C225</td>
<td>NA</td>
<td>0.013</td>
<td>0.006</td>
<td>2</td>
</tr>
</tbody>
</table>

**NOTE:** $K_D$ values for binding to cells were determined by fitting the data from Fig. 1 to the Lineweaver–Burk equation. Abbreviation: NA, not applicable.

### Table 2. Comparison of measured apparent IgG affinities for A431 cells versus affinities calculated by using a mathematical model

<table>
<thead>
<tr>
<th>IgG</th>
<th>Experimental $K_D$, nmol/L</th>
<th>Model $K_D$, nmol/L</th>
<th>Model $K_D$ (equilibrium), nmol/L</th>
<th>$K_D$(Fab)/$K_D$(IgG)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C10</td>
<td>1.17</td>
<td>1.5</td>
<td>1.5</td>
<td>82.8</td>
</tr>
<tr>
<td>P3/5</td>
<td>0.5</td>
<td>0.054</td>
<td>0.054</td>
<td>1,078</td>
</tr>
<tr>
<td>P2/1</td>
<td>0.012</td>
<td>0.0071</td>
<td>0.0051</td>
<td>4,980</td>
</tr>
<tr>
<td>P2/4</td>
<td>0.0064</td>
<td>0.0071</td>
<td>0.0051</td>
<td>4,941</td>
</tr>
<tr>
<td>P2/2</td>
<td>0.0077</td>
<td>0.0059</td>
<td>0.0034</td>
<td>5,323</td>
</tr>
<tr>
<td>3524</td>
<td>0.012</td>
<td>0.0057</td>
<td>0.0026</td>
<td>5,923</td>
</tr>
<tr>
<td>2124</td>
<td>0.007</td>
<td>0.0053</td>
<td>0.00033</td>
<td>3,969</td>
</tr>
<tr>
<td>2224</td>
<td>0.007</td>
<td>0.0053</td>
<td>0.00031</td>
<td>3,870</td>
</tr>
</tbody>
</table>

**NOTE:** Fab affinities from Table 1. $K_D$(Fab)/$K_D$(IgG) was corrected for nonequilibrium conditions.
used to simulate binding experiments of sufficient time (10,000-hour incubation period in the \textit{in silico} experiment) to assure equilibrium binding conditions. These simulations revealed that equilibrium constants measured for the high affinity antibodies such as 2224 and 2124 were underestimated by more than 10-fold due to insufficient incubation time at low antibody concentrations (Table 2, third column, and Supplementary Fig. S2). Thus, the mathematical modeling of antibody–receptor binding indicates that the biphasic effect is an artifact of assay limitations (Table 2, fourth column). It was not possible to further increase the incubation time of antibody with cells as the requirement for antibody to be in excess over total receptor numbers led to unacceptable cell loss due to a small number of cells in a large incubation volume.

\textbf{Impact of EGFR density on antibody apparent affinity}

To further validate the computational model of antibody binding, we measured the affinity of C10 and 2224 IgG on tumor cells expressing different levels of EGFR (Fig. 1A). We expected that both antibodies would show improvements in observed affinity on cells expressing higher levels of EGFR. EGFR density was determined for A431, MDA-MB-468, and MDA-MB-231 cells using QSC beads (Fig. 1B). For low affinity IgG C10, observed binding strength increased with the receptor density, from $K_D = 13.8 \text{ nmol/L}$ on MDA-MB231 cells to $K_D = 2.2 \text{ nmol/L}$ on A431 cells, showing a correlation between apparent affinity and receptor density. However, for high affinity IgG 2224, we observed that binding strength improved with decreasing EGFR expression: a 5-fold higher apparent affinity was obtained for MDA-MB231 cells, which have 10-fold less EGFR than A431 cells.

We found that the computational model of IgG binding was able to recreate the observed dependence of C10 and 2224 IgG binding affinity on EGFR expression level (Supplementary Table S3). Furthermore, the observed equilibrium constants for high affinity antibody 2224 were again underestimated due to insufficient incubation time in the experimental conditions (Supplementary Table S3). The equilibrium constants calculated from the mathematical model are similar for IgG 2224 on all 3 tumor cells despite the 10-fold difference in their receptor density, suggesting a broader targeting spectrum for EGFR-expressing tumors by using higher affinity antibodies.

\textbf{Impact of EGF on the binding of EGFR antibodies to EGFR-overexpressing cells}

Like EGFR antibodies cetuximab and panitumumab (12, 13), the C10 family of antibodies bind an epitope that
overlaps with the EGF-binding site. As a result, EGF in the local tumor microenvironment could inhibit antibody binding to EGFR and impact tumor targeting. We therefore determined the ability of EGF to compete with IgG antibody binding to EGFR. Using 3 mAbs in the C10 family, we tested IgG at 3 different antibody concentrations (1, 10, and 100 nmol/L) and Fab at 100 nmol/L measuring their binding to A431 cells in the presence of 0.1 to 100 nmol/L of EGF. EGF competitively inhibited the binding of the antibodies to cells (Fig. 2A–D), confirming that the C10 panel of antibodies share an epitope with EGF. For Fab (Fig. 2D), EGF inhibition of Fab binding was proportional to the Fab intrinsic affinity. Bivalent binding of IgG leading to an increased apparent affinity resulted in at least a 10-fold increase in the EGF IC$_{50}$ value for inhibition of antibody binding to cells (Fig. 2A–D). However, higher IgG intrinsic affinity also resulted in an increase in the EGF IC$_{50}$ value for inhibition of antibody binding to cells (Fig. 2A vs. B). For example, P2/4 and 2224 IgG (with similar apparent affinities but 10-fold different intrinsic affinities) differed in their EGF IC$_{50}$ values by approximately 50-fold (Fig. 2A). Thus, EGF IC$_{50}$ values could largely be explained by the relationship between the EGF K$_D$ for binding to EGFR (1 nmol/L) and the intrinsic and apparent affinity of the antibodies.

**Impact of intrinsic and apparent affinities on EGF induced EGFR phosphorylation**

Enhancement of cell proliferation by EGF occurs via phosphorylation of EGFR and downstream signaling. A previous study has shown that the phosphorylation of EGFR was inhibited more effectively by bivalent form of antibodies than by monovalent Fab fragments (11). We analyzed the antiphosphorylation effect of the C10 panel of EGFR antibodies in both monovalent (scFv) and bivalent (IgG) forms to dissect out the contribution of intrinsic versus apparent affinities. Phosphorylation of EGFR was stimulated by 10 nmol/L EGF. At low antibody concentration relative to EGF concentration (20:10 nmol/L), inhibition of EGFR phosphorylation was seen only for the IgG and in proportion to the intrinsic affinity (Fig. 3B). At higher antibody: EGF concentration ratios (400:10), inhibition was seen for both scFv and IgG, again in proportion to the intrinsic affinity (Fig. 3A). By comparing the relative inhibition of scFv to the cognate IgG, it is also clear that the increased apparent affinity of the IgG leads to greater inhibition of phosphorylation than the scFv with the same binding site (Fig. 3A, P2/4 and 2224 IgG vs. scFv). As a control, neither the scFv nor IgG of anti-HER2 antibody C6.5 inhibited EGFR phosphorylation, showing target-specific blockade of EGFR activation by anti-EGFR antibodies. No reduction in total EGFR level was observed with the blockade of EGFR activation under the tested condition.

**Impact of intrinsic and apparent affinities on antibody antiproliferative capacity**

Antibodies against EGFR inhibit the proliferation of cells that overexpress EGFR by blocking the binding of ligands to EGFR (10). To determine the impact of intrinsic and apparent affinity on EGFR antibody anti-proliferative activity, we evaluated the ability of 3 scFv and their cognate IgG to inhibit the growth of A431 cells in vitro. The lowest affinity scFv (C10) showed no significant
Inhibition of A431 proliferation whereas the P2/4 and 2224 scFv inhibited proliferation with IC_{50} values of 300 and 35 nmol/L, respectively (Fig. 4A). Thus, the inhibition of A431 cell growth by the scFv showed a strong correlation with their intrinsic affinities. Likewise, inhibition of proliferation by Fab was greatest for the highest affinity Fab. Fab potency was less than scFv potency (Fig. 4A and B), possibly due to our observation that scFv versions of these antibodies tend to form multivalent aggregates. For the IgGs, growth inhibition also correlated with the intrinsic affinity, with a 2.2 nmol/L IC_{50} value for the highest affinity IgG 2224 and more than 160 nmol/L IC_{50} value for the lowest affinity IgG C10 (Fig. 4C). Thus, while P2/4 and 2224 IgG experimentally have comparable apparent affinities, their IC_{50} values for inhibition of proliferation differ by more than 30-fold. The IC_{50} value of the 2224 IgG was also 67-fold less than that of the 2224 Fab and 16-fold lower than the 2224 scFv, which indicated a significant impact of apparent affinity (bivalent binding) on the antiproliferative effect. It is also notable that the impact of the bivalent binding was greater for the IgG with higher intrinsic affinity than the IgG with lower intrinsic affinity. The anti-EGFR antibody C225, which bound the extracellular domain of EGFR with 0.21 nmol/L IC_{50} value measured by KinExA (Supplementary Table S1), inhibited the cell proliferation to a similar extent as 2224 for IgG (Fig. 4C) and slightly better than 2224 for Fab form (Fig. 4B). As a negative control, the anti-BoNT antibody 3D12 did not show proliferative inhibition.

Discussion

One of the attractive features of antibodies as therapeutics is the ability to use the tools of antibody engineering to tune affinity, valency, and pharmacokinetics. This allows testing and identification of the optimal antibody properties for a given application. Many therapeutic antibodies in oncology function by binding to cell surface receptors at an epitope, which blocks ligand binding, thus inhibiting receptor activation. While it may seem intuitive that higher antibody affinity for antigen would be beneficial in this setting, no studies have rigorously explored the relationship between affinity, valency, and the ability of antibodies to inhibit signaling and proliferation driven by oncogenic targets like EGFR.

To address this question, we studied EGFR antibodies binding an identical epitope with intrinsic affinities differing by more than 250-fold in monovalent form. Upon conversion of these antibodies to IgG format, we surprisingly observed a biphasic relationship between intrinsic monovalent affinity and bivalent-binding strength. This is in contrast to our previous results using a set of IgG affinity variants targeting HER2 at the same epitope, where we observed a monotonic decline in the observed ratio of intrinsic monovalent K_{D} to functional bivalent K_{D} (35). In the current work, we used a computational binding model to successfully predict the observed IgG affinities, and to further show that the biphasic effect results from nonequilibrium experimental conditions that underestimate the actual functional binding strength for the higher affinity antibodies. Similar nonequilibrium effects are also likely present in the results of Tang and colleagues (35) and specific experimental conditions the cause of the different observed relationships between intrinsic and functional affinity. While it might appear that the solution to this measurement problem is to increase incubation time beyond the 16 hours used, such studies are intractable due to poor cell recovery in the large incubation volumes and small numbers of cells needed to ensure antibody excess over receptor number. Fortunately, mathematical modeling allows the prediction of true binding affinities even if experimentally infeasible to measure.

In addition to the cell-binding studies, the anti-EGFR antibodies were analyzed in both monovalent and bivalent forms for their abilities to inhibit cell proliferation and EGF signaling. Signaling and proliferation inhibition by both monomeric scFv and Fab correlated with their monovalent-binding constants, whereas the bivalent IgG exhibited much greater inhibition than comparable affinity monovalent antibodies, due to higher functional affinity. IgG derived from higher intrinsic affinity-binding sites (such as 2224) had greater inhibition of EGFR phosphorylation and cell growth than IgG
with increasing Fab intrinsic affinity, observed for the scFv. C, inhibition of proliferation of A431 cells increased with increasing IgG intrinsic affinity, but this effect was less than that observed for the scFv. C, inhibition of proliferation of A431 cells increased with increasing Fab intrinsic affinity, but this effect was less than that observed for the scFv. C, inhibition of proliferation of A431 cells increased with increasing IgG intrinsic affinity. 3D12, control anti-botulinum neurotoxin antibody.

Figure 4. Inhibition of A431 cell proliferation by anti-EGFR scFv, IgG and Fab. A, inhibition of proliferation of A431 cells increased with increasing scFv intrinsic affinity, with an IC50 value of 35 nmol/L for the highest affinity 2224 scFv. B, inhibition of proliferation of A431 cells increased with increasing Fab intrinsic affinity, but this effect was less than that observed for the scFv. C, inhibition of proliferation of A431 cells increased with increasing IgG intrinsic affinity. 3D12, control anti-botulinum neurotoxin antibody.

derived from lower intrinsic binding sites (P2/4), even though P2/4 and 2224 had comparable apparent affinities (due to assay limitations). These results indicate that for antibodies that inhibit signaling, both higher intrinsic affinity and avid bivalent binding lead to greater in vitro potency.

Whether this results in greater in vivo efficacy remains unproven. The overall efficacy of antibody therapies results not only from signaling inhibition, but also from effective tumor penetration and retention and for some targets antibody-dependent cell-mediated cytotoxicity and complement-dependent cytotoxic antibody activity. We and others have shown that for antibodies targeting solid tumors, affinities lower than Kd = 1 to 10 nmol/L do not increase quantitative tumor accumulation and may actually reduce the magnitude of tumor targeting (21, 23, 26, 28). In fact, using a series of HER2-targeted antibody variants binding the same epitope with different affinity, we have recently shown that higher affinity antibodies are more rapidly internalized and degraded in tumors than antibodies of lower affinity, limiting tumor penetration (28). This effect was particularly pronounced for the IgG with an intrinsic binding site Kd significantly less than 1 nmol/L. While the EGFR IgG studied did not have intrinsic Kd in this range, such effects are likely to occur for EGFR antibodies, and might be more pronounced due to the faster rate of EGFR internalization relative to HER2. In addition, higher intrinsic affinity could result in greater binding of shed tumor antigen in the tumor microenvironment than in antibodies of lower intrinsic affinity, interfering with cell binding and signaling inhibition. These “binding site barriers” (29–31) for tumor delivery may result in higher affinity antibodies having lower efficacy despite a higher intrinsic ability to inhibit signaling pathways. Experimental in vivo evaluation of antibodies binding identical epitopes but with varying intrinsic and functional affinities will help elucidate the impact of these interconnected factors on overall antibody efficacy.

Disclosure of Potential Conflicts of Interest

J.D. Marks has major ownership interest in Merrimack Pharmaceuticals. He also is a consultant and on the scientific advisory board of Merrimack Pharmaceuticals. No potential conflicts of interest were disclosed by the other authors.

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Analysis and interpretation of data (e.g., statistical analysis, biostatistics, computational analysis): Y. Zhou, A.L. Goenaga, B.D. Harms, F. Conrad, J.D. Marks
Writing, review, and/or revision of the manuscript: Y. Zhou, A.L. Goenaga, B.D. Harms, J. Lou, F. Conrad, G.P. Adams, B. Schoeberl, U.B. Nielsen
Administrative, technical, or material support (i.e., reporting or organizing data, constructing databases): H. Zou, F. Conrad
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