Ganetespib, a Unique Triazolone-Containing Hsp90 Inhibitor, Exhibits Potent Antitumor Activity and a Superior Safety Profile for Cancer Therapy

Weiwen Ying, Zhenjian Du, Lijun Sun, Kevin P. Foley, David A. Proia, Ronald K. Blackman, Dan Zhou, Takayo Inoue, Noriaki Tatsuta, Jim Sang, Shuxia Ye, Jamie Acquaviva, Luisa Shin Ogawa, Yumiko Wada, James Barsoum, and Keizo Koya

Abstract

Targeted inhibition of the molecular chaperone Hsp90 results in the simultaneous blockade of multiple oncogenic signaling pathways and has, thus, emerged as an attractive strategy for the development of novel cancer therapeutics. Ganetespib (formerly known as STA-9090) is a unique resorcinolic triazolone inhibitor of Hsp90 that is currently in clinical trials for a number of human cancers. In the present study, we showed that ganetespib exhibits potent in vitro cytotoxicity in a range of solid and hematologic tumor cell lines, including those that express mutated kinases that confer resistance to small-molecule tyrosine kinase inhibitors. Ganetespib treatment rapidly induced the degradation of known Hsp90 client proteins, displayed superior potency to the ansamycin inhibitor 17-allylamino-17-demethoxygeldanamycin (17-AAG), and exhibited sustained activity even with short exposure times. In vivo, ganetespib showed potent antitumor efficacy in solid and hematologic xenograft models of oncogene addiction, as evidenced by significant growth inhibition and/or regressions. Notably, evaluation of the microregional activity of ganetespib in tumor xenografts showed that ganetespib was efficiently distributed throughout tumor tissue, including hypoxic regions >150 μm from the microvasculature, to inhibit proliferation and induce apoptosis. Importantly, ganetespib showed no evidence of cardiac or liver toxicity. Taken together, this preclinical activity profile indicates that ganetespib may have broad application for a variety of human malignancies, and with select mechanistic and safety advantages over other first- and second-generation Hsp90 inhibitors.

Introduction

Hsp90 is a molecular chaperone that regulates the posttranslational folding, stability and function of its protein substrates (client proteins), many of which play critical roles in cell growth, differentiation, and survival (1, 2). As with other physiologic processes that become co-opted by tumor cells, it is now clear that the chaperoning functions of Hsp90 can become subverted during tumorigenesis to facilitate malignant progression (1). The Hsp90 machinery serves as a biochemical buffer for a number of oncogenic signaling proteins causally implicated in a variety of tumors (3, 4). Often, these oncoproteins are expressed as mutant forms that are particularly reliant on Hsp90 for stability and function (5, 6). Cancer cells contain elevated levels of the active form of the Hsp90 complex relative to normal cells, and have been shown to be selectively sensitive to Hsp90 inhibition (7, 8). Moreover, a unique characteristic of targeting Hsp90 is that inhibition results in the combinatorial blockade of multiple signal transduction cascades, thereby potentially bypassing pathway redundancies often found in cancer cells (9–11). Thus, Hsp90 represents an attractive molecular target for the development of novel cancer therapeutics (4, 11, 12).

The first class of Hsp90 inhibitors to be characterized were the benzoquinone ansamycins, including geldanamycin and its derivatives 17-allylamino-17-demethoxygeldanamycin (17-AAG), and 17-dimethylaminoethylamino-17-demethoxygeldanamycin (17-DMAG; ref. 13). However, the clinical progression of this group has been hampered because of several drawbacks including poor solubility, formulation problems, potential multidrug efflux, and hepatotoxicity (13, 14). In addition, as single agents, these inhibitors have only shown modest efficacies in the clinical setting (15, 16), indicating that they may be most effective as combination therapies. In an effort to overcome these limitations, several second-generation synthetic Hsp90 inhibitors representing
multiple drug classes are currently under development (17–23).

In this article, we describe the preclinical characterization of ganetespib (formerly known as STA-9090), a novel small-molecule inhibitor of Hsp90 with a unique triazolone-containing chemical structure. Key pharmacologic and biologic properties of ganetespib distinguish this compound from other first- and second-generation Hsp90 inhibitors with regard to potency, antitumor activity, and an improved safety profile resulting in a superior therapeutic index. Accordingly, ganetespib is currently being evaluated in multiple phase I and II clinical trials. Taken together, these results support the continued development of ganetespib as a novel therapeutic agent for a variety of human cancers.

Materials and Methods

Cell lines, antibodies, and reagents

All cell lines were obtained from the American Type Culture Collection (ATCC) and maintained according to standard techniques. The cell lines were authenticated by the routine ATCC Cell Biology Program using short tandem repeat analysis (DNA profiling) and were used within 6 months of receipt for this study. All primary antibodies were purchased from Cell Signaling Technology. Ganetespib [3-(2,4-dihydroxy-5-isopropylphenyl)-4-(1-methyl-1H-indol-5-yl)-1H-1,2,4-triazol-5(4H)-one] was synthesized by Synta Pharmaceuticals Corp. 17-AAG, 17-DMAG, and erlotinib were purchased from LC Laboratories. Purified Hsp90 protein was obtained from Stressgen and recombinant human hepatocyte growth factor (HGF) from R&D Systems.

Cell viability assays

Cells were grown in 96-well plates based on optimal growth rates determined empirically for each line. Twenty-four hours after plating, cells were treated with the indicated compounds or controls for 72 hours. AlamarBlue (Invitrogen) was added (10% v/v) to the cells, and the plates were incubated for 3 hours and, then, subjected to fluorescence detection. For the comparative viability/apoptosis assay, NCI-H1975 cells were treated with escalating concentrations of ganetespib for the indicated time periods and subjected to viability analysis via CellTiter Fluor (Promega) and apoptosis via Caspase Glo 3/7 (Promega).

Western blotting

Following treatment, tumor cells were disrupted in lysis buffer (CST) on ice for 10 minutes. Lysates were clarified by centrifugation and equal amounts of proteins resolved by SDS-PAGE before transfer to nitrocellulose membranes (Invitrogen). Membranes were blocked with 5% skim milk in Tris-buffered saline with 0.5% Tween and immunoblotted with the indicated antibodies. The antibody–antigen complex was visualized and quantitated using an Odyssey system (LI-COR).

In vivo xenograft tumor models

Female immunodeficient Crl:CD1-Foxn1\textsuperscript{nu} (nude) and CB-17/1cr-Ptkdr\textsuperscript{cld}+/Crl severe combined immunodeficient (SCID) mice (Charles River Laboratories) were maintained in a pathogen-free environment, and all in vivo procedures were approved by the Institutional Animal Care and Use Committee of Synta Pharmaceuticals Corp. NCI-H1975 and MV4-11 cells were subcutaneously implanted into SCID mice and MKN45 cells into nude mice. Mice bearing established tumors (100–200 mm\textsuperscript{3}) were randomized into treatment groups of 8 and intravenously dosed via the tail vein with either vehicle or ganetespib formulated in 10/18 DRD (10% dimethyl sulfoxide, 18% Cremophor RH 40, 3.6% dextrose, 68.4% water). In the NCI-H1975 model, studies were conducted at the highest nonseverely toxic doses of 150 mg/kg weekly; in the MV4-11 model, animals were treated with ganetespib at 100 and 125 mg/kg weekly; in the MKN45 model, animals were treated with ganetespib at 50 mg/kg 3 times a week. Tumor growth inhibition was determined as described previously (24).

Microregional activity of ganetespib in NCI-H1975 xenografts

NCI-H1975 tumor xenograft-implanted SCID mice were treated with 125 mg/kg ganetespib for 6 to 72 hours. At the end of the experiment, mice were administered bromodeoxyuridine (BrdUrd) and pimonidazole to label S-phase cells and hypoxic tumor regions and, then 5 minutes before excision, mice were administered DiOC7(3) to demarcate perfused vessels. Following tumor excision and freezing, 10-μm thick cryosections were cut and sequentially immunostained to detect markers of proliferation (BrdUrd), apoptosis (TUNEL), hypoxia (HIF-1α), and tumor vasculature (CD31). Images of CD31 fluorescence, BrdUrd, and terminal deoxynucleotidyl transferase–mediated dUTP nick-end labeling (TUNEL) staining from each section were overlaid, and areas of necrosis and staining artifacts were manually removed. Proliferation and apoptosis were plotted as a function of distance from vessels.

Hepatotoxicity assay

Male Sprague–Dawley rats were treated with repeated daily administration of escalating doses of 17-DMAG (formulated in 5% dextrose in water) or ganetespib (formulated in DRD). Blood serum was collected during necropsy at the end of the experiment in all rats treated with or without testing compound. The serum liver enzyme tests were conducted at IDEXX Laboratories according to their validated standard operation procedure. For histologic analysis, livers were removed at necropsy and formalin fixed. Paraffin-embedded sections were processed and stained with hematoxylin and eosin for routine histologic evaluation.
Langendorff assay

Briefly, hearts from male New Zealand white rabbits were used to measure the physiologic variables PQ, QRS, RR, QT, and dLVP/dt following perfusion with escalating doses of ganetespib (10⁻⁸–10⁻⁴ mmol/L; see Supplementary Materials and Methods for complete details). Mean values for each parameter were calculated for each concentration, and mean values (±SEM) were plotted against concentration for all parameters assessed, both for ganetespib-exposed and vehicle-treated hearts.

Results

Ganetespib binds to the N-terminal ATP-binding site in Hsp90

Ganetespib is a novel resorcinolic triazolone compound that is structurally distinct to the first-generation ansamycin Hsp90 inhibitors. The chemical structure is shown in Fig. 1A. With a molecular weight of 364.4, ganetespib is considerably smaller than the ansamycin class, and most of the newer, second-generation Hsp90 inhibitors. Ganetespib is relatively hydrophobic, with a cLogP value of 3.3. Ganetespib exhibits competitive binding for the ATP pocket at the N-terminus of Hsp90. There are reports that the N-terminus can be crystallized in a number of conformations including an open or closed conformation in reference to the position of the ATP-binding pocket lid (25). We have obtained the co-crystal structure of ganetespib bound to the closed conformation of the Hsp90 N-terminus (Fig. 1B); however, we anticipate that ganetespib can also access the ATP pocket in the open conformation based on computational analysis (data not shown).

The X-ray co-crystal structure of ganetespib bound to Hsp90 (Fig. 1C) confirmed important hydrogen bonding interactions, also seen in the ansamycin family, involving the resorcinol hydroxyl group with Asp⁹³ and the carbonyl group of triazolone with Lys⁵⁸. Importantly, in ganetespib, the 2-hydroxyl of resorcinol is within hydrogen bonding distance to both oxygen atoms of the carboxylic group in Asp⁹³, resulting in a substantially stronger interaction. Furthermore, the N² of triazolone forms a water-bridged hydrogen bond with Asp⁹³ to provide additional hydrogen bonding. Water-bridge hydrogen bonds between 4-hydroxyl of resorcinol and Leu⁴⁸ and Ser⁵² were found to be critical for binding efficiency in our optimization efforts. The hydrazinecarboxamide moiety of triazolone in ganetespib is of particular structural importance. In addition to the direct hydrogen bond with Lys⁵⁸, it forms a unique hydrogen bond with Gly⁹⁷, a distinguishing feature from the ansamycin analogues. Furthermore, it interacts with Thr¹⁸⁴ and Asp¹⁰² through water-bridge hydrogen bonding (Fig. 1C).

Ganetespib displays superior potency to 17-AAG in a broad range of tumor cells

The in vitro cytotoxic activity of ganetespib was determined against a panel of 57 transformed cell lines derived from both hematologic and solid tumors and compared with that of 17-AAG (Supplementary Table S1). Ganetespib was potently cytotoxic in the majority of the lines examined, typically with IC₅₀ values in the low nanomolar range. Overall, ganetespib showed a 20-fold greater potency than 17-AAG with median IC₅₀ values of 14 versus 280 nmol/L, respectively. This difference in sensitivity was more significantly pronounced in the subset of hematologic malignancies, which showed a 47.5-fold difference (median IC₅₀ values of 10 vs. 475 nmol/L). Indeed, leukemic cell lines [acute myeloid leukemia (AML), chronic myeloid leukemia (CML), B-cell lymphoma, and anaplastic large-cell lymphoma (ALCL)] manifested the greatest sensitivity to ganetespib treatment, whereas melanoma and prostate cancer represented tumor types in which ganetespib was also highly cytotoxic (Supplementary Table S1). Notably, ganetespib retained potency against cell lines expressing mutated kinases that confer resistance to kinase inhibitors that are currently used in clinical practice.

Ganetespib induces cell-cycle arrest and apoptosis

Cell-cycle analysis showed that ganetespib induced marked accumulation in the G₂–M phase within 24 hours in NCI-H1975 cells, with a concomitant loss of S phase (Supplementary Fig. S1). The viable cell population remained blocked for at least 72 hours; however, over this period, the percentage of apoptotic cells increased. To confirm this, cells were exposed to increasing concentrations of ganetespib for 6 to 72 hours. Apoptosis was
Ganetespib was assessed 72 hours following drug wash-out. RFU, relative fluorescence units.

NCI-H1975 and HCC827 cells were exposed to graded concentrations of ganetespib for 5, 15, and 60 minutes and 24 hours. Cell viability was quickly affected by ganetespib treatment and measured using activated caspase 3/7 levels and compared with cell viability (Fig. 2A). No effects were seen 6 hours after treatment. However, the marked loss of viability following exposure to ganetespib observed 24 to 48 hours posttreatment correlated with increased apoptotic induction. These results indicate that ganetespib-induced cytotoxicity is mediated by an irreversible commitment to apoptosis, which is likely subsequent to growth arrest and effects on the cell cycle (24).

Ganetespib displays potent activity against drug-resistant tumor phenotypes in vitro

In NSCLC, activating mutations in epidermal growth factor receptor (EGFR) can drive tumorigenesis and confer sensitivity to tyrosine kinase inhibitors (TKI) such as erlotinib and gefitinib (26). To examine whether ganetespib could overcome the resistant phenotype in NSCLC cells, we compared the activities of ganetespib and erlotinib (Supplementary Fig. S2) using the NCI-H1975 cell line, which expresses a mutationaly activated and erlotinib-resistant EGFR<sup>L858R/T790M</sup> mutation, and the erlotinib-sensitive HCC827 cells, which express EGFR<sup>Del E746_A750</sup> (Fig. 3A). As expected, erlotinib treatment resulted in dose-dependent cytotoxicity in HCC827 cells, but had no effect on NCI-H1975 cells. In contrast, ganetespib exhibited full potency against both cell lines, irrespective of EGFR mutational status.

In addition, resistance to EGFR inhibitors may emerge through alternative oncogenic mechanisms. HGF, a ligand of the c-MET oncoprotein, can induce TKI resistance in lung tumors with EGFR-activating mutations by independently activating and restoring phosphoinositide 3-kinase (PI3K)–AKT signaling via phosphorylation of c-MET (27, 28). Given that EGFR, c-MET, and AKT are all Hsp90 client proteins, we, therefore, determined whether ganetespib was active against c-MET–induced TKI-resistant cells. HCC827 cells were seeded in the presence or absence of HGF (50 ng/mL) and, 24 hours later, were dosed with ganetespib or erlotinib. Cell viability was assessed 72 hours after addition of the drug (Fig. 3B).
Both ganetespib and erlotinib were highly potent in non-stimulated HCC827 cells, with IC_{50} values of approximately 10 nmol/L. Importantly, whereas HGF-treated cells did not respond to erlotinib, ganetespib retained its potency in the presence of the growth factor. Similar results were observed in HCC827 cells selected for potency in the presence of the growth factor. Cells did not respond to erlotinib, ganetespib retained its potency in the presence of the growth factor. The activity of ganetespib was assessed in NCI-H1975 tumor xenografts. 

Ganetespib exhibits potency against erlotinib-resistant NSCLC tumor phenotypes

Ganetespib exhibits potency against erlotinib-resistant NSCLC tumor phenotypes (Fig. 4B).

To examine modulation of MET-driven AKT signaling by ganetespib, HCC827 cells seeded with or without HGF for 24 hours were treated with ganetespib or erlotinib at either 10 or 100 nmol/L doses (which represent the IC_{50} and IC_{100} drug concentrations, respectively). Cells were harvested at 24 hours, and levels of MET, EGFR, and their relevant effectors were examined by Western blot analysis (Fig. 3C).

Ganetespib treatment promoted the downregulation of MET, EGFR, and AKT protein levels in both the absence and presence of HGF, resulting in complete loss of AKT and extracellular signal-regulated kinase (ERK) activity. Erlotinib exposure was capable of inactivating AKT and ERK in the absence of HGF, but was ineffective in the presence of the growth factor.

**Ganetespib exhibits potent in vivo activity in both solid and hematologic xenograft models**

To determine whether the effects of ganetespib in vitro translate to antitumor efficacy in vivo, the activity of ganetespib was evaluated using a variety of doses and schedules in a series of xenograft models. Initially, SCID mice bearing NCI-H1975 NSCLC xenografts were dosed intravenously with ganetespib on a weekly schedule at its nearest CD31 endothelial cells (Fig. 5). A single dose of ganetespib at 125 mg/kg dramatically reduced cellular proliferation throughout the tumors, with the maximal effect occurring 24 hours following treatment (Fig. 5A).

Furthermore, a concomitant induction of tumor cell apoptosis occurred within 24 hours (Fig. 5B). Moreover, at 6 hours, apoptosis was preferentially induced near vessels and then became uniformly induced throughout the tissue by the 48-hour time point (data not shown). Increased HIF-1α staining as a function of distance confirmed the hypoxic gradient that existed within the tumors and, importantly, this Hsp90 client protein was potently altered associated with AML (29). This cell line is highly sensitive to ganetespib in vitro (IC_{50}: 4 nmol/L; Supplementary Table S1). Ganetespib was administered intravenously to MV4-11 tumor–bearing SCID mice once weekly at 100 and 125 mg/kg. As shown in Fig. 4C, these 2 treatment regimens resulted in significant tumor regression (85% and 94%, respectively). Moreover, tumors were undetectable in 37.5% of ganetespib–treated animals at the end of the 3-week dosing period.

Amplification of the c-MET receptor tyrosine kinase occurs in approximately 20% of gastric carcinomas (30). We, therefore, used the human c-MET amplified MKN45 gastric carcinoma cell line as an additional xenograft model of oncogene addiction to examine the antitumor activity of ganetespib (Fig. 4D). Ganetespib treatment was again highly efficacious in this study, with a 50 mg/kg dose 3 times per week resulting in 92% inhibition of tumor growth.

Ganetespib penetrates hypoxic regions of tumors in vivo

To evaluate tumor penetration, the microregional activity of ganetespib was assessed in NCI-H1975 tumor xenografts. Immunohistochemical markers of proliferation (BrdUrd), apoptosis (TUNEL), and hypoxia (HIF-1α) in tumors were mapped in relation to distance from the nearest CD31 endothelial cells (Fig. 5). A single dose of ganetespib at 125 mg/kg dramatically reduced cellular proliferation throughout the tumors, with the maximal effect occurring 24 hours following treatment (Fig. 5A). Furthermore, a concomitant induction of tumor cell apoptosis occurred within 24 hours (Fig. 5B). Moreover, at 6 hours, apoptosis was preferentially induced near vessels and then became uniformly induced throughout the tissue by the 48-hour time point (data not shown). Increased HIF-1α staining as a function of distance confirmed the hypoxic gradient that existed within the tumors and, importantly, this Hsp90 client protein was potently altered associated with AML (29). This cell line is highly sensitive to ganetespib in vitro (IC_{50}: 4 nmol/L; Supplementary Table S1). Ganetespib was administered intravenously to MV4-11 tumor–bearing SCID mice once weekly at 100 and 125 mg/kg. As shown in Fig. 4C, these 2 treatment regimens resulted in significant tumor regression (85% and 94%, respectively). Moreover, tumors were undetectable in 37.5% of ganetespib–treated animals at the end of the 3-week dosing period.

Amplification of the c-MET receptor tyrosine kinase occurs in approximately 20% of gastric carcinomas (30). We, therefore, used the human c-MET amplified MKN45 gastric carcinoma cell line as an additional xenograft model of oncogene addiction to examine the antitumor activity of ganetespib (Fig. 4D). Ganetespib treatment was again highly efficacious in this study, with a 50 mg/kg dose 3 times per week resulting in 92% inhibition of tumor growth.
suppressed in vivo following treatment with ganetespib (Fig. 5C). These results provide strong evidence that ganetespib efficiently distributed within the extravascular compartment, including the hypoxic regions >150 μm from the microvasculature, resulting in sustained inhibition of proliferation and induction of apoptosis throughout the tumors.

**Ganetespib exhibits a favorable safety profile**

The hepatotoxicity profile of ganetespib was evaluated in male Sprague–Dawley rats on the basis of changes in the liver enzymes aspartate aminotransferase (AST) and alanine aminotransferase (ALT; Fig. 6A). Animals were treated with repeated administration of ganetespib at 25, 50, and 75 mg/kg/d for 5 days, or 17-DMAG (Supplementary Fig. S2) at 2, 4, and 6 mg/kg/d for 4 days. No changes in the levels of either enzyme were observed in the ganetespib-treated animals, even at the highest dose of 75 mg/kg, which is higher than the efficacious dose range used to treat the tumors.

Cardiovascular effects of escalating doses ganetespib on electrophysiologic (PQ, QRS, RR, and QTc) and mechanical (left ventricular developed pressure) properties were evaluated in isolated New Zealand white rabbit hearts. Ganetespib exerted no significant physiologic effects other than a minimal reduction in atrioventricular conduction (lengthening PQ interval) and a minor reduction in heart rate (increased RR interval) over the concentrations 10⁻⁸ to 10⁻⁵ mol/L (data not shown). There was no change in the QTc(F) intervals at concentrations of ganetespib between 10⁻⁸ and 10⁻⁵ mol/L when compared with baseline or vehicle (Fig. 6C). Similarly, there was no change in the QRS duration after exposure to concentrations of ganetespib ranging from 10⁻⁸ to 10⁻⁵ mol/L, when compared with baseline or vehicle; however, an increase in the duration of the QRS was noted after exposure to the 10⁻⁵ mol/L concentration (Fig. 6C). At 10⁻⁴ mol/L, the highest concentration tested, ganetespib caused lengthening of PQ interval and QRS duration; however, this concentration was approximately 3,000-fold higher than the unbound Cmax in the 125 mg/kg dose in the NCI-H1975 tumor-penetration studies (Fig. 5). Other cardiac electrophysiologic parameters and mechanical properties, including left ventricular developed pressure, were not significantly altered following exposure to ganetespib.
ganetespib, whereas expected physiologic changes with the positive control quinidine were observed (data not shown).

Discussion

In this article, we provide the first preclinical characterization of ganetespib, a novel and potent inhibitor of Hsp90 that offers considerable promise as a new targeted cancer therapeutic agent. Structurally, ganetespib is distinct from the first-generation ansamycin Hsp90 inhibitors, with a unique scaffold that is considerably smaller than these geldanamycin analogues. Hsp90 chaperone activity is associated with an ATP-driven conformational change within the N-terminal domain (31, 32). Our data show that ganetespib is able to enter the ATP-binding pocket in the so-called closed conformation. In contrast, due to their larger size, the ansamycin analogues can only occupy the ATP-binding pocket in the open conformation. For clarity, this discussion of open and closed conformation of the Hsp90 N-terminal refers to the positioning of the ATP pocket lid, which is different from the open and closed concept in the Hsp90 chaperoning cycle involving dimerization (33). This lack of restriction for binding to the Hsp90 ATP pocket may be one of the reasons that ganetespib shows higher in vitro potency than the geldanamycin analogues. In addition, our structural analysis has identified a series of additional hydrogen bond interactions due to the presence of the triazolone moiety that predict for superior binding affinities between ganetespib and Hsp90, further distinguishing this compound from the geldanamycin class.

Human cancers are typically characterized by a variety of genetic alterations that collectively contribute to the transformed state; however, a subset is now believed to be dependent on single, definable oncogenic pathways for their genesis, proliferation, and/or survival. This phenomenon is known as oncogene addiction (34) and, because a large number of the addicting oncoproteins are known Hsp90 clients, this has important implications for the development of targeted therapeutics. The panel of solid and hematologic tumor lines found to be sensitive to the ATP pocket lid, which is different from the open and closed concept in the Hsp90 chaperoning cycle involving dimerization (33). This lack of restriction for binding to the Hsp90 ATP pocket may be one of the reasons that ganetespib shows higher in vitro potency than the geldanamycin analogues. In addition, our structural analysis has identified a series of additional hydrogen bond interactions due to the presence of the triazolone moiety that predict for superior binding affinities between ganetespib and Hsp90, further distinguishing this compound from the geldanamycin class.

Human cancers are typically characterized by a variety of genetic alterations that collectively contribute to the transformed state; however, a subset is now believed to be dependent on single, definable oncogenic pathways for their genesis, proliferation, and/or survival. This phenomenon is known as oncogene addiction (34) and, because a large number of the addicting oncoproteins are known Hsp90 clients, this has important implications for the development of targeted therapeutics. The panel of solid and hematologic tumor lines found to be sensitive to...
ganetespib was derived from diverse tissue origins. Notably, many of the most acutely sensitive lines harbored activating mutations or amplifications of these oncoproteins, including EGFR, c-MET, BCR-ABL, B-RAF, c-KIT, and HER2, consistent with the hypothesis that these oncogenic drivers are more reliant on Hsp90 for their stability and function (35). The in vitro activity of ganetespib translated to potent antitumor efficacy in a series of xenograft models selected for their dependence on such oncogenic pathways for growth. In addition, ganetespib exhibited robust cellular potency against drug-resistant tumor phenotypes, overcoming molecular alterations in NSCLC lines that confer clinical resistance to a number of small-molecule TKIs. Furthermore, we discovered that even brief exposure to ganetespib (as little as 5 to 60 minutes) resulted in potent cytotoxic responses in NSCLC cells with IC50 values readily achievable in vivo. Emerging evidence from the use of small-molecule TKIs in a variety of human cancers indicates that transient, potent oncoprotein inhibition can be sufficient to induce clinically relevant effects on cellular viability (36, 37). Taken together, these findings strongly indicate that ganetespib is likely to

![Graph A](image1.png)

**Figure 6.** Liver and cardiovascular effects profile of ganetespib. A, male Sprague-Dawley rats (n = 3–5 rats per group) were treated with repeated administration of 17-DMAG at 2, 4, and 6 mg/kg for 4 days (qd × 4) or ganetespib at 25, 50, and 75 mg/kg for 5 days (qd × 5). Plasma levels of the liver enzymes aspartate aminotransferase (AST) and alanine aminotransferase (ALT) are presented. B, hematoxylin and eosin (H&E) staining of liver cross sections from vehicle control, 17-DMAG-, or ganetespib-treated animals. Original magnification, ×200 (second panel, ×400).

**C.** effects of escalating doses of ganetespib and vehicle on the QTc (F) interval (left) and QRS interval (right) in male rabbit hearts. Ganetespib values presented as the mean (±SEM); n = 6, except n = 2 at 10^-4 mol/L and n = 4 at 10^-6 mol/L. No measurements were possible at 10^-7 mol/L due to arrhythmias. Because of differences in formulation procedure, 4 hearts were perfused with concentrations of 10^-7 to 10^-4 mol/L of ganetespib. The remaining 2 hearts were perfused with concentrations of 10^-8 to 10^-6 mol/L of ganetespib. Therefore, the vehicle time points in parentheses in the figure correspond to when the 4 test hearts differ from the 2 test hearts. Vehicle values are presented from 1 animal.
have broad therapeutic use in a variety of human malignancies and imply that the durable response property of ganetespib may support the use of intermittent dosing schedules in the clinic.

With its unique chemical structure, ganetespib exhibited several key pharmacologic and biologic properties sufficient to account for the potent antitumor responses observed. The compound has relatively high lipophilicity that, along with its smaller size, would be expected to facilitate transport across lipid membranes and into cells. In xenograft bearing animals, ganetespib showed selective retention in tumor tissue with similar kinetics as those reported for other resorcinolic Hsp90 inhibitors (38). The most important observation, however, was that the physicochemical properties of the compound resulted in extensive penetration and distribution of ganetespib throughout tumors, including hypoxic regions distal to the nearest blood vessels. From a clinical perspective, the effectiveness of many anticancer agents can be compromised by limited drug distribution, as efficient penetration is necessary to reach the target population and in concentrations sufficient to exert a therapeutic effect (39). In this regard, ganetespib treatment rapidly and dramatically reduced proliferation and induced apoptosis in xenograft tissues independent of the distance from the microvasculature. The capacity of ganetespib to penetrate the extracellular compartment of solid tumors combined with its extended tumor retention and cellular potency clearly predicts for maximal efficacy.

Overall unfavorable safety profiles, including hepatotoxicity, have hampered the clinical application of the ansamycin class of Hsp90 inhibitors. There the chemical reactivity of the benzoquinone group appears accountable for the observed elevation of liver enzymes and associated liver toxicity in the clinical setting (4). Consistent with the findings for other rationally designed second-generation inhibitors, ganetespib showed no evidence of liver toxicity in the preclinical assessment of changes in liver enzymes or histopathology. In addition, cardiac toxicity is a potential risk factor for many classes of drugs, in part, due to adverse effects on critical ion channels that regulate the beating of the heart (37). Our cardiovascular analysis revealed that the 2 predominant effects attributable to exposure to ganetespib in the Langendorff assay were a slight dose-dependent lengthening of the PQ interval and a minor change in QRS duration at the 10^{-5} mol/L concentration. Moreover, the observation that the QTc interval did not lengthen is consistent with ganetespib having no effect on ventricular repolarization. Together with no significant alterations in other electrophysiologic and mechanical parameters, ganetespib, therefore, exhibits a favorable cardiotoxic profile. Importantly, our safety studies were carried out to maximally tolerated doses and no other adverse events were seen to indicate that ganetespib manifests any additional toxicities. Notably, ocular toxicities have recently emerged as an undesirable side-effect for the newer synthetic Hsp90 inhibitors (40). To date, more than 400 patients have been treated with ganetespib, and an absence of ocular toxicity is evident (<3%; W. Ying, unpublished observation). Currently a comprehensive profiling of ganetespib and other Hsp90 inhibitors for potential CNS or ocular toxicity risks is underway.

In summary, we have developed and characterized a unique small-molecule Hsp90 inhibitor that exhibits potent and sustained antitumor effects in a broad range of malignancies both in vitro and in vivo. Importantly, ganetespib retained its potency against tumor phenotypes that confer drug resistance to agents currently in use in the clinic. In addition, ganetespib displays optimal pharmacologic properties including high tumor penetration and a favorable safety profile that predict for a superior therapeutic index. Accordingly, ganetespib represents an exciting new targeted agent for the treatment of human cancers.

Disclosure of Potential Conflicts of Interest

All authors are current or former employees of Synta Pharmaceuticals Corp.

Acknowledgments

The authors thank Tim Koburt, Donald Smith, Chaohua Zhang, and Yu Liu for their excellent technical assistance and dedication to the project and Richard Bates who provided drafts and editorial assistance during preparation of the manuscript.

Grant Support

All research work was funded by Synta Pharmaceuticals Corp. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received September 20, 2011; revised November 22, 2011; accepted November 22, 2011; published OnlineFirst December 5, 2011.
Molecular Cancer Therapeutics

Ganetespib, a Unique Triazolone-Containing Hsp90 Inhibitor, Exhibits Potent Antitumor Activity and a Superior Safety Profile for Cancer Therapy

Weiwen Ying, Zhenjian Du, Lijun Sun, et al.

*Mol Cancer Ther* 2012;11:475-484. Published OnlineFirst December 5, 2011.

Updated version  Access the most recent version of this article at: doi:10.1158/1535-7163.MCT-11-0755

Supplementary Material  Access the most recent supplemental material at: http://mct.aacrjournals.org/content/suppl/2011/12/08/1535-7163.MCT-11-0755.DC1

Cited articles  This article cites 38 articles, 14 of which you can access for free at: http://mct.aacrjournals.org/content/11/2/475.full#ref-list-1

Citing articles  This article has been cited by 21 HighWire-hosted articles. Access the articles at: http://mct.aacrjournals.org/content/11/2/475.full#related-urls

E-mail alerts  Sign up to receive free email-alerts related to this article or journal.

Reprints and Subscriptions  To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at pubs@aacr.org.

Permissions  To request permission to re-use all or part of this article, contact the AACR Publications Department at permissions@aacr.org.