Antitumoral Effects of Calcitriol in Basal Cell Carcinomas Involve Inhibition of Hedgehog Signaling and Induction of Vitamin D Receptor Signaling and Differentiation

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Abstract

Activation of the Hedgehog (Hh)-signaling pathway due to deficiency in the Hh receptor Patched1 (Ptch) is the pivotal defect leading to formation of basal cell carcinoma (BCC). Recent reports provided evidence of Ptch-dependent secretion of vitamin D3-related compound, which functions as an endogenous inhibitor of Hh signaling by repressing the activity of the signal transduction partner of Ptch, Smoothened (Smo). This suggests that Ptch-deficient tumor cells are devoid of this substance, which in turn results in activation of Hh-signaling. Here, we show that the physiologically active form of vitamin D3, calcitriol, inhibits proliferation and growth of BCC of Ptch mutant mice in vitro and in vivo. This is accompanied by the activation of the vitamin D receptor (Vdr) and induction of BCC differentiation. In addition, calcitriol inhibits Hh signaling at the level of Smo in a Vdr-independent manner. The concomitant antiproliferative effects on BCC growth are stronger than those of the Hh-specific inhibitor cyclopamine, even though the latter more efficiently inhibits Hh signaling. Taken together, we show that exogenous supply of calcitriol controls the activity of 2 independent pathways, Hh and Vdr signaling, which are relevant to tumorigenesis and tumor treatment. These data suggest that calcitriol could be a therapeutic option in the treatment of BCC, the most common tumor in humans. Mol Cancer Ther; 10(11); 2179–88. ©2011 AACR.

Introduction

The Hedgehog (Hh)-signaling pathway regulates cell differentiation, organ patterning, and cell proliferation (1). In the absence of a ligand, the activity of the Hh-pathway is inhibited due to an interaction between the Hh receptor Patched1 (Ptch) and its partner Smoothened (Smo). Binding of Hh to Ptch, inactivating Ptch mutations, or activating Smo mutations may suspend this inhibition, resulting in the transcription of target genes including Gli1 (1). Thus aberrant (e.g., mutation driven) Hh signaling results in tumor formation (2). One prominent example is basal cell carcinoma (BCC), the most frequent tumor entity in humans, with active Hh-signaling due to mutations in Ptch.

Today, the inhibition of the Hh pathway is considered to be a promising strategy in the treatment of these tumors. Thus specific Smo inhibitors such as cyclopamine and GDC0449 have been tested in several small, nonrandomized clinical trials (2). GDC0449 was recently shown to elicit antitumoral effects in 18 of 33 patients with locally advanced or metastatic BCC (3). It remains to be elucidated if these promising results will be confirmed in a prospective, randomized, and controlled study and if Hh targeting alone will be sufficient. In this context, a recent report describes resistance to GDC0449 due to a therapy-associated Smo mutation (4). As with other tumors, therapy of those associated with abnormal Hh signaling may require targeting of additional signaling pathways.

Vitamin D3 and its derivatives (e.g., EB1089; Paricalcitol) are known to have antitumoral effects on different cancer types (5) including squamous cell carcinoma (6, 7) or on hyperproliferative skin diseases such as psoriasis (8, 9). These effects comprise G0−G1 arrest, cellular differentiation, induction of apoptosis, and modulation of inflammation or of different signaling pathways in tumor cells, as well as inhibiting tumor angiogenesis (5). Until now, the antitumoral effects of vitamin D3 were explained by binding of the biologically active form of vitamin D3, calcitriol (1α,25-dihydroxy vitamin D3), to the vitamin D receptor (Vdr) and the subsequent regulation of Vdr-bound genes.

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Calcitriol is produced from vitamin D₃ by 2 hydroxylation steps in the liver and kidney, respectively, and to a lesser extent in other organs and in tumor cells (5). Through binding to Vdr, calcitriol regulates the transcription of Vdr target genes (5). This so-called genomic calcitriol/Vdr signaling regulates a variety of physiologic processes including cellular differentiation, especially in the skin (10, 11), proliferation, and apoptosis, and can be monitored by measuring transcription of the calcitriol metabolizing enzyme 24-hydroxylase (5). In addition, calcitriol elicits rapid, so-called nongenomic (i.e., transcription independent) effects such as calcium influx (5).

Recent data suggest a cross-talk between vitamin D₃ and Hh signaling. Through medium transfer experiments Bijlsma and colleagues provided first evidence for a Ptch-dependent secretion of vitamin D₃ compounds. They also showed that vitamin D₃ inhibits the Hh-pathway at the level of Smo (12). This suggests that, in addition to or instead of a direct protein–protein interaction, Ptc may repress Smo via secretion of a vitamin D₃ derivative (12).

The possibility of a Ptch-dependent secretion of a vitamin D₃ derivative with Smo-inhibitory properties opens new perspectives for therapies of tumors that arise due to mutations in Ptc. One would expect that inactivation of Ptc results in a disrupted secretion of this derivative. This should result in a potential deprivation of the vitamin D₃ derivative, and in lack of Smo inhibition. If this hypothesis is true, the concerted action of active Hh and inactive Vdr signaling may be the driving force leading to enhanced cell proliferation, compromised differentiation, and ultimately to tumor formation. It follows that it should be possible to revert or at least to impede these processes by administration of the respective vitamin D₃ derivative.

Here, we investigated the effect of the biologically active vitamin D₃ derivative calcitriol on Vdr- and Hh-signaling, growth, apoptosis, and differentiation of Ptch-deficient BCC cells in vitro and in vivo using the Ptchlox/lox Rosa26CreERT2+/− (Ptchlox/lox ERT2+/−) mouse model for BCC (13). In addition, we compared its effects with those of cyclopamine and sought to unravel the molecular mechanisms underlying the calcitriol-mediated effects on Hh signaling.

Materials and Methods

Compounds
Calcitriol (Sigma-Aldrich) and cyclopamine (Toronto Research Chemicals Inc.) were dissolved in ethanol (EtOH). Final concentrations for in vitro experiments are indicated in the respective experiments and correspond to those normally used in cell culture (5, 14, 15). For in vivo use, calcitriol was diluted individually for each animal in 20 μL EtOH/1,200 μL sterile sunflower oil (Sigma-Aldrich) to obtain a final concentration of 40 or 100 ng/kg in 50 μL.

Animals and treatment of tumor-bearing Ptchlox/lox ERT2+/−, mice with calcitriol
Conditional Ptchllox/lox ERT2+/− mice were randomized into 2 groups and BCC was induced in all animals by intramuscular injection of 100 μg tamoxifen as described (13, 16). Tumors of this animal model lack the expression of wt Ptc alleles and are therefore deficient in Ptc (13, 17). Starting points of the daily intraperitoneal treatment with 100 ng/kg calcitriol or vehicle were day 0 or 60 after BCC induction. Treatment of each cohort was conducted until day 90 after BCC induction. For RNA isolation and histologic examinations, skin samples were collected from tails 24 hours after the last calcitriol injection.

Mice were fed with calcium- and phosphate-reduced and vitamin D₃-free food (ssniff Spezialdiaeten, E15312-14; ref. 18) 1 week before and during the injection period. All animals were treated and housed in accordance with the German animal protection law.

Measurement of tumor size
BCC size was measured on hematoxylin and eosin (H&E)-stained sections using the area calculation tool of the software CellF (Olympus Soft Imaging Solutions GmbH).

Analysis of calcium blood serum values
Amounts of 100 μL of blood were collected from the retroorbital plexus. Serum calcium concentrations were measured using an O-cresolphthaleine–based assay (cobas, Roche Diagnostics GmbH).

Histopathology and immunohistochemistry
BCC and normal skin from the tail were embedded in paraffin for histologic analyses or were used for isolation of total RNA. The identity of BCC was confirmed by examination of H&E-stained sections. Paraffin sections were stained using an anti-Ki67- and anti-active caspase 3-antibody as described (19).

Cell lines and primary cell culture of BCC
The fibroblast cell line Ptchllox/lox ERT2+/− was established from dermis of a Ptchllox/lox ERT2+/− mouse. Ptch−/− cells are stable Ptch-deficient cells derived from tamoxifen-treated Ptchllox/lox ERT2+/− fibroblasts. Smo−/− and Vdr−/− fibroblasts have been described in Ma and colleagues (20) and Sun and colleagues (21), respectively. All fibroblast cell lines were maintained in Dulbecco’s Modified Eagle Medium (DMEM), 10% fetal calf serum, and 1% penicillin/streptomycin (PS).

The murine BCC cell line ASZ001 was established from UV-induced BCC of Ptchm22−/− mice and maintained as described (22). All cell lines have been tested and authenticated before using by genotyping PCRs on genomic DNA as described (13, 20, 21).

Skin punches were isolated from BCC-bearing Ptchllox/lox ERT2+/− mice 30, 40, and 60 days after tumor induction.
initiation by tamoxifen. Punches were maintained for 10 days in culture as reported (16).

**Cell culture experiments**

For gene expression analysis or 5-bromo-2′-deoxyuridine (BrdU) incorporation and caspase assays, 100,000 or 4,000 cells/well were seeded in 6-well and 96-well plates, respectively. For short interfering RNA (siRNA)-mediated knockdown of Vdr expression, Pelch\textsuperscript{fluox}\textsuperscript{fluox} ERT2\textsuperscript{+/−} cells were transfected with Vdr-specific or control siRNA as described below. After 24 hours, the cells were washed and incubated for additional 48 hours with medium supplemented with calcitriol, cyclospamine, or EtOH as indicated in the respective experiments.

Cell proliferation was measured after BrdU-pulsing for the last 22 hours using a Cell Proliferation BrdU ELISA (Roche Diagnostics GmbH).

Activity of caspase 3 and 7 was measured using the Caspase-Glo 3/7 Assay (Promega) and a microplate reader (Biotek instruments Inc.) according the manufacturer’s instructions.

Shh-N-conditioned medium (Shh-N-CM) or respective control medium were obtained from HEK293-Shh (HEK293-Shh express one of the 3 mammalian Hh proteins Sonic hedgehog (Shh)) or HEK293 cells, respectively, as described (14).

Knockdown of Vdr expression in Pelch\textsuperscript{−/−} and Pelch\textsuperscript{fluox}\textsuperscript{fluox} ERT2\textsuperscript{+/−} cells was achieved by using a Vdr-specific siRNA (5′-CAGCGGAAGCATGAAGCGCAA-3′). Scrambled siRNA (AllStars negative, Qiagen) was used as control siRNA. 75 ng siRNA and 4.5 μL HiPerFect (Qiagen) were mixed with 100 μL DMEM and incubated for 10 minutes. Simultaneously, 15,000 cells/well were seeded in DMEM/10% fetal calf serum/2% penicillin/streptomycin in 24-well plates. Subsequently, the siRNA/HiPerFect-mix was added to the cells. After 24 hours, the cells were incubated with the respective media (as indicated in the experiments) supplemented with vehicle or 10 nmol/L calcitriol. After additional 48 hours, the cells were collected and used for subsequent experiments.

For activation of Hh-signaling pathway in Smo\textsuperscript{−/−} fibroblasts the cells were transfected with a plasmid-expressing hSmo (15, 23) using RotiFect following the manufacturer’s instructions. After 6 hours, the cells were incubated with media containing 10 nmol/L calcitriol, 5 μmol/L cyclospamine, or EtOH for 48 hours.

Data shown are representative for at least 3 independent experiments each conducted in triplicate.

**Reverse transcription PCR and quantitative real-time PCR analyses**

Total RNA was extracted using TriReagent (Sigma-Aldrich). For skin samples, the RNeasy fibrous tissue mini kit (Qiagen) was used. Synthesis of cDNA and primer combinations for amplification of 18S rRNA, Gli1 and keratin10 (K10) transcripts used for quantitative real-time PCR (qRT-PCR) were described previously (16, 19). Primer pairs used for qRT-PCR analysis of Gli2, Vdr, Cyp24a1, and transglutaminase 1 (Tgm1) were 5′-GCAAAGTCTAA-GACTGAGCCTGA-3′/5′-GCTGCTCTTGCTGCTCATAT-3′/5′-AGAAACTGTGCTGCTCATAGG-3′/5′-CTACCTTGCCGTAGGCCGTGCTGCTGCT; 5′-GTTGGC- CAAGGCGCAACACGCT-3′/5′-CCGTGACAGGCGGTATACAGT-3′; and 5′-GCAGTGGTGTTAATGCAGCTGG-3′/5′-ATGAGGAGCTCAAGGGCAATGC-3′, respectively.

Amplification of 18S RNA was done as an endogenous control for the normalization of target gene expression. The amount of target and endogenous reference was determined using the relative standard curve method. Each sample was measured in triplicates. Graphs represent the mean value of all measurements.

Glycerol-3-phosphate dehydrogenase (mGapd), wild-type (wt) Pelch transcripts (derived from the nonrecombined Pelch\textsuperscript{fluox} locus) and Pelch\textsuperscript{fluox} were detected by RT-PCR as described in Zibat and colleagues (13, 24).

**Protein extraction and Western blot**

Pelch\textsuperscript{−/−} fibroblasts were transfected with Vdr-specific siRNA or control siRNA as described. After 72 hours, the cells were harvested. Nuclear fractions of the transfected cells and of Vdr\textsuperscript{−/−} fibroblasts were isolated using the kit NE-PER Nuclear and Cytoplasmatic Extraction Reagents (Thermo Scientific) according to the manufacturer’s instructions. Protein concentrations were measured by Pierce BCA Protein Assay Kit (Thermo Scientific) and 16.5 μg of each protein sample of the nuclear fraction were analyzed by Western blot using a rabbit anti-Vdr antibody (1:100; clone 9A7; Abcam) and a mouse anti-heat shock 70 kDa protein 8 (HSC-70; 1:10,000; Santa Cruz) in combination with horseradish peroxidase-conjugated goat anti-rat IgG (1:10,000; Thermo Scientific) and rabbit antimonozyme IgG (1:5,000; GE Healthcare), respectively. Signals were detected by using ECL reagent (GE Healthcare).

**Statistics**

Mann–Whitney U testing was done to determine the significance of the results.

**Results**

Calcitriol activates the Vdr pathway, suppresses Hh signaling and inhibits proliferation of cultured Pelch mutant BCC cells

As human BCC (5, 25–27), BCC from Pelch\textsuperscript{fluox}\textsuperscript{fluox} ERT2\textsuperscript{+/−} mice express elevated Vdr levels compared with normal murine skin (Fig. 1A).

To test the response of Pelch-deficient murine BCC cells to calcitriol in vitro, BCC-bearing skin punches were collected 30, 40, or 60 days after BCC initiation in Pelch\textsuperscript{fluox}\textsuperscript{fluox} ERT2\textsuperscript{+/−} mice (see Materials and Methods). The punches were cultured with 10 nmol/L calcitriol or with 10 μmol/L cyclospamine for 10 days (the structures of cyclospamine and calcitriol are provided in Fig. 1B). Gli1 expression levels served to monitor...
Figure 1. Calcitriol inhibits proliferation and the Hh-signaling pathway of BCC-bearing skin punches and of the BCC cell line ASZ001. A, Vdr expression levels of BCC \((n = 3)\) of Ptchflox/floxERT2\(^{+/-}\) mice 90 days after tumor induction compared with NS \((n = 3)\). B, chemical structures of calcitriol and cyclopamine. C, Gli1 and Cyp21a1 expression levels of cultured BCC-bearing skin punches isolated from Ptchflox/floxERT2\(^{+/-}\) mice 30, 40, or 60 days after BCC induction. D, Ki67\(^+\) BCC cells of the respective punches. Gli1 and Cyp21a1 expression levels and caspase 3/7 activities of ASZ001 cells. Cells treated with 500 nmol/L staurosporin served as positive controls. The punches and the ASZ001 cells were incubated with vehicle (EtOH), calcitriol, or cyclopamine (CP) as indicated in the text. Gli1 and Cyp21a1 expression levels (E) and BrdU incorporation (F) in ASZ001 cells. G, caspase 3/7 activities of ASZ001 cells. Cells treated with 500 nmol/L staurosporin served as positive controls. The punches and the ASZ001 cells were incubated with vehicle (EtOH), calcitriol, or cyclopamine (CP) as indicated in the text. Gli1 expression and caspase 3/7 activities are shown in relation to the respective vehicle-treated controls. Ki67\(^+\) BCC cells and BrdU-incorporation are represented as percentage of respective vehicle-treated controls. **, \(P < 0.05\); error bars: mean \pm SD.
Calcitriol inhibited Hh signaling downstream of PtcH but upstream of Gli1

Vitamin D₃ has been shown to inhibit Hh signaling at the level of Smo (12, 35, 36). To investigate whether the mechanism of Hh-pathway inhibition mediated by calcitriol is similar to that of Vitamin D₃, we made use of PtcHlox/loxERT2²⁺/⁻ and Smo⁻/⁻ fibroblasts. PtcHlox/loxERT2²⁺/⁻ fibroblasts normally express wt PtcH transcripts from the PtcHlox locus. Upon tamoxifen-induced activation of ERT2 recombine the exons 8 and 9 of the floxed PtcH alleles are excised. This results in the expression of an aberrant, nonfunctional PtcH transcript (Fig. 3A).

In PtcHlox/loxERT2²⁺/⁻ fibroblasts both, the treatment with Shh-N-CM or the tamoxifen-induced PtcH mutation result in activation of Hh signaling as revealed by induction of Gli1 expression (Fig. 3B). Coincubation of Shh- or tamoxifen-treated PtcHlox/loxERT2²⁺/⁻ fibroblasts with calcitriol significantly inhibited Gli1 expression (Fig. 3B). Similar results were obtained with the validated Smo inhibitor cyclopamine (Fig. 3B). These data show that similar to cyclopamine, calcitriol inhibits Hh signaling downstream of PtcH.

The effects of calcitriol were also tested in Smo⁻/⁻ cells, which express basal Gli1 levels. Due to lack of Smo, Hh-signaling cannot be activated in these cells by incubation with Shh-conditioned medium (37). However, in these cells Hh-pathway activity can be restored by transfection with a hSMO expression plasmid.

As shown in Fig. 3C, calcitriol did not influence basal Gli1 expression levels in Smo⁻/⁻ cells (Fig. 3C). However, calcitriol efficiently inhibited Hh signaling after
restoration of Hh-pathway activity upon transfection with hSMO. These results are similar to those obtained with cyclopamine (Fig. 3C) and show that calcitriol normally inhibits Gli1 expression at the level of Smo.

**Calcitriol inhibits Hh signaling in a Vdr-independent manner**

Next, we tested whether calcitriol-mediated inhibition of the Hh-signaling pathway is independent of Vdr-signaling. For this purpose, we used Vdr<sup>−/−</sup> fibroblast (21). In these cells Vdr signaling is completely abrogated, since incubation with calcitriol does not result in the induction of the Vdr-target Cyp24a1 (Fig. 4A). As revealed by Gli1 expression, incubation of the cells with Shh-N-CM resulted in induction of Hh-pathway activity (Fig. 4A), which was significantly inhibited by addition of calcitriol or by the control substance cyclopamine (Fig. 4A). Similar results were achieved in Ptch<sup>−/−</sup> or Shh-N-CM-treated Ptch<sup>−/−</sup> ERT2<sup>+/−</sup> cells after siRNA-mediated Vdr knockdown. Efficient downregulation of Vdr expression was verified by Western blot (Fig. 4B), qRT-PCR and by a significantly lower Cyp24a1 expression after calcitriol treatment in comparison with the controls (Fig. 4C, D). Whereas Vdr knockdown per se did not significantly attenuate Hh-pathway activity, treatment with calcitriol resulted in a downregulation of Gli1 expression (Fig. 4C and D). These data show that calcitriol-mediated inhibition of the Hh-signaling pathway occurs independently of the Vdr.

Taken together, these data show that calcitriol inhibits Hh-pathway activity downstream of Ptch at the level of Smo in a Vdr-independent manner.

**Discussion**

The active form of vitamin D<sub>3</sub>, calcitriol, is rapidly gaining importance in oncology due to its antiproliferative and differentiation-inducing effects combined with its low toxicity (5).

Our work extends the range of calcitriol tumor targets to those induced by an abnormal activity of the Hh-signaling pathway (summarized in Fig. 5). Specifically, we show that calcitriol inhibits growth of BCC induced by deletion of the Hh receptor Ptch. This is accompanied by...
the expression of relevant cell differentiation markers. Our data suggest that calcitriol could be a valuable supplement or even alternative to the established treatments of BCC, the most common tumor in humans, associated with aberrant Hh-pathway activity.

As assessed by reduced Gli1 transcription, calcitriol inhibits canonical Hh signaling independently of Vdr signaling and downstream of Ptch (Fig. 5). An obvious molecular target of this Vdr-independent effect of calcitriol is Smo, because Smo-deficient cells (unlike those reconstituted with Smo or Ptch-deficient ones) show no decreased Gli1 transcription in response to this substance. A similar observation has been made for the inactive form of calcitriol, vitamin D₃ (12). Nevertheless, whether calcitriol directly binds to Smo should be addressed in future studies.

Besides inhibition of Hh-signaling pathway, calcitriol inhibits proliferation, and increases the expression of skin differentiation marker in BCC. The latter effects are also induced by calcitriol in skin of patients with hyperproliferative skin diseases such as psoriasis (6–9, 34, 38). Calcitriol treatment specifically results in inhibition of proliferation of psoriatic skin and induction of differentiation of keratinocytes (9, 39, 40). Similarly to BCC, psoriatic skin also expresses high levels of the Hh-target gene Gli1 (41). These facts raise the question whether calcitriol mediates its antiproliferative effects and differentiation stimuli via activation of Vdr signaling or rather via inhibition of the Hh-signaling pathway. A clue might come from our present study: Our in vitro study shows that calcitriol has a significantly stronger antiproliferative effect on BCC than the pure Smo inhibitor cyclopamine, even though the latter substance inhibits Hh signaling more efficiently (Fig. 1D and F). Together with the fact that both calcitriol (present study) and cyclopamine (42) inhibit BCC proliferation in vivo, it is possible that calcitriol exerts its antiproliferative effects via both signaling pathways (e.g., by inhibition or activation of Hh or Vdr signaling, respectively). On the other hand, differentiation of calcitriol-treated BCC is probably unrelated to inhibition of the canonical Hh-signaling pathway, because cyclopamine never has been reported to induce the expression of keratinocyte differentiation markers in BCC. More likely, calcitriol induces BCC differentiation via Vdr signaling, which is supported by the increased expression of the Vdr-target genes Cyp24a1 and Tgm1 (Fig. 2F).

Antiproliferative and Hh-signaling inhibitory properties have also been described for the inactive form of calcitriol, vitamin D₃, on murine BCC in vitro and in vivo (35). In contrast to calcitriol, vitamin D₃ did not affect
differentiation in BCC, although it was topically applied at high concentrations. Supposedly the treatment period using vitamin D3 (i.e., 30 days) may have been too short to induce Vdr signaling and thus a differentiation response. This suggestion is based on a comparison with our study in which a 30 days calcitriol application also had no significant effects on these processes (see BCC treated for days 60 to 90, Fig. 2B and E).

Figure 4. Calcitriol inhibits the Hh-signaling pathway in a Vdr-independent manner. A, Gli and Cyp24a1 expression of Vdr−/− fibroblasts after treatment with vehicle (EtOH), calcitriol, or cyclopamine (CP) and Shh-N-CM or control medium (CoM). B, analyses of the Vdr protein level of nuclear extracts of si-Vdr or si-control transfected Ptch−/− fibroblasts by Western blot. Detection of HSC-70 protein served as control. C and D, Gli, Cyp24a1, and Vdr expression of Ptch−/− and Shh-stimulated Ptchfloxt/floxErt2+/− cells are shown in relation to the respective vehicle-treated control. **, $P < 0.05$; error bars: mean ± SD.

Figure 5. Model for the dual function of calcitriol in Ptch-associated BCC. Normally Ptch inhibits its signaling partner Smo, thereby regulating the activity of the Hh-signaling pathway (normal cell). Mutations of Ptch lead to a constitutive activation of the Gli transcription factors, which resulted in cell proliferation and tumor formation (BCC cell). The known Smo-inhibitor cyclopamine inhibits Hh-pathway in the Ptch-mutant cells and thus Hh-related processes involved in tumor growth (cyclopamine-treated BCC cell). In contrast, calcitriol inhibits Hh-pathway activity and additionally activates Vdr signaling (calcitriol-treated BCC cell). Consequently, calcitriol not only inhibits tumor-relevant processes mediated by Hh signaling, but also induces antiproliferative effects and differentiation processes via the Vdr-signaling pathway.
Finally, a new model of tumorigenesis driven by Pch-deficiency may emerge from our study. According to a recent work, Pch might function as an efflux pump for vitamin D₃-related compounds with Hh-inhibitory properties (12). A deficiency of this compound due to Pch inactivation would pathologically activate Hh-pathway and reduce Vdr signaling. Vice versa application of this compound should result in inhibition of Hh-pathway and activation of Vdr signaling (Fig. 5). Whether this vitamin D₃-related compound is calcitriol remains to be analyzed in the future.

Taken together the application of calcitriol holds promises as an effective anticancer drug in the treatment of BCC. Due to its dual effects on both Vdr and Hh signaling, it may be superior to substances that solely target the Hh-signaling pathway. Calcitriol treatment may also be superior to application of vitamin D₃, which has to be metabolized before activating Vdr signaling. The benefits of topical application of calcitriol in treatment of BCC have to be tested in the future.

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Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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